



Swansea University
Prifysgol Abertawe



Cronfa - Swansea University Open Access Repository

This is an author produced version of a paper published in:
Antimicrobial Agents and Chemotherapy

Cronfa URL for this paper:
<http://cronfa.swan.ac.uk/Record/cronfa38876>

Paper:

Colley, T., Sehra, G., Chowdhary, A., Alanio, A., Kelly, S., Kizawa, Y., Armstrong-James, D., Fisher, M., Warrilow, A., et. al. (2018). In Vitro and In Vivo Efficacy of a Novel and Long Acting Fungicidal Azole, PC1244 on *Aspergillus fumigatus* Infection. *Antimicrobial Agents and Chemotherapy*, AAC.01941-17
<http://dx.doi.org/10.1128/AAC.01941-17>

Released under the terms of a Creative Commons Attribution 4.0 International license (CC-BY).

This item is brought to you by Swansea University. Any person downloading material is agreeing to abide by the terms of the repository licence. Copies of full text items may be used or reproduced in any format or medium, without prior permission for personal research or study, educational or non-commercial purposes only. The copyright for any work remains with the original author unless otherwise specified. The full-text must not be sold in any format or medium without the formal permission of the copyright holder.

Permission for multiple reproductions should be obtained from the original author.

Authors are personally responsible for adhering to copyright and publisher restrictions when uploading content to the repository.

<http://www.swansea.ac.uk/library/researchsupport/ris-support/>

Revision, AAC01941-17 (Research Article, T.Colley/G.Sehra)

1 Title

2 ***In Vitro* and *In Vivo* Efficacy of a Novel and Long Acting Fungicidal Azole,**
3 **PC1244 on *Aspergillus fumigatus* Infection**

4

5 Thomas Colley^{1¶}, Gurpreet Sehra^{1¶}, Anuradha Chowdhary², Alexandre Alanio^{3,4,5}, Steven L.
6 Kelly⁶, Yasuo Kizawa⁷, Darius Armstrong-James^{8,9}, Matthew C. Fisher¹⁰, Andrew G.S.
7 Warrilow⁶, Josie E. Parker⁶, Diane E. Kelly⁶, Genki Kimura⁷, Yuki Nishimoto⁷, Mihiro Sunose¹¹,
8 Stuart Onions¹¹, Damien Crepin¹¹, Franz Lagasse¹¹, Matthew Crittall¹¹, Jonathan Shannon¹¹,
9 Matthew McConville¹¹, John King-Underwood¹², Alan Naylor¹³, Stéphane Bretagne^{3,4,5}, John
10 Murray¹, Kazuhiro Ito^{1*}, Pete Strong¹ and Garth Rapeport¹.

11

12 ¹Pulmocide Ltd, London, UK, ²Department of Medical Mycology, Vallabhbai Patel Chest
13 Institute, University of Delhi, Delhi, India, ³Institut Pasteur, CNRS, Molecular Mycology Unit,
14 French National Reference Center for Invasive Mycoses & Antifungals, URA3012, Paris, ⁴Paris
15 Diderot, Sorbonne Paris Cité University, Paris, France, ⁵Parasitology-Mycology Laboratory,
16 Saint Louis Hospital, Assistance Publique-Hôpitaux de Paris (APHP), Paris, France, ⁶Centre for
17 Cytochrome P450 Biodiversity, Institute of Life Science, Swansea University Medical School,
18 Wales, UK, ⁷School of Pharmacy, Nihon University, Chiba, Japan, ⁸Department of
19 Microbiology, Royal Brompton Hospital, London, UK, ⁹National Heart and Lung Institute,
20 Imperial College School of Medicine, London, UK, ¹⁰Department of Infectious Disease
21 Epidemiology, Imperial College School of Public Health, St Mary's Campus, London, UK,
22 ¹¹Sygnature Discovery Ltd, Nottingham, UK, ¹²CompChem Resource, Ledbury, UK. ¹³Alan
23 Naylor Consultancy Ltd., Royston, UK.

Revision, AAC01941-17 (Research Article, T.Colley/G.Sehra)

24

25 Running title: Antifungal activity of novel inhaled azole PC1244

26 Key words: *Aspergillus fumigatus*, azole, inhalation, CYP51, azole resistant

27 # Address correspondence to Kazuhiro Ito: kaz@pulmocide.com

28 ¶ These authors contributed equally to this work.

29

Revision, AAC01941-17 (Research Article, T.Colley/G.Sehra)

30 **ABSTRACT**

31 The antifungal effects of the novel triazole, PC1244, designed for topical or inhaled
32 administration, against *A. fumigatus* have been tested in a range of *in vitro* and *in vivo* studies.
33 PC1244 demonstrated potent antifungal activities against clinical *A. fumigatus* isolates (N=96)
34 with a MIC range of 0.016–0.25 µg/ml, whereas the MIC range for voriconazole was 0.25–0.5
35 µg/ml. PC1244 was a strong tight-binding inhibitor of recombinant *A. fumigatus* CYP51A and
36 CYP51B (sterol 14 α -demethylase) enzymes and strongly inhibited ergosterol synthesis in *A.*
37 *fumigatus* with an IC₅₀ of 8 nM. PC1244 was effective against a broad spectrum of pathogenic
38 fungi (MIC ranged from <0.0078~2 µg/ml), especially on *Aspergillus terreus*, *Trichophyton*
39 *rubrum*, *Candida albicans*, *Candida glabrata*, *Candida krusei*, *Cryptococcus gattii*,
40 *Cryptococcus neoformans* and *Rhizopus oryzae*. PC1244 also proved to be quickly absorbed into
41 both *A. fumigatus* hyphae and bronchial epithelial cells, producing persistent antifungal effects.
42 In addition, PC1244 showed fungicidal activity (MFC, 2 µg/ml), which was 8-fold more potent
43 than voriconazole. *In vivo*, once daily intranasal administration of PC1244 (3.2 ~ 80µg/mL) to
44 temporarily neutropenic, immunocompromised mice 24h after inoculation with itraconazole-
45 susceptible *A. fumigatus* substantially reduced fungal load in the lung, galactomannan in serum
46 and circulating inflammatory cytokines. Furthermore, 7 days extended prophylaxis with PC1244
47 showed superior *in vivo* effects when compared against 1 day of prophylactic treatment,
48 suggesting accumulation of the effects of PC1244. Thus, PC1244 has the potential to be a novel
49 therapy for the treatment of *A. fumigatus* infection in the lungs of humans.
50

Revision, AAC01941-17 (Research Article, T.Colley/G.Sehra)

51 INTRODUCTION

52 The incidence of fungal infections has increased substantially over the past two decades and
53 invasive forms are leading causes of morbidity and mortality, especially amongst
54 immunocompromised or immunosuppressed patients. In addition, chronic lung infections with
55 *Aspergillus*, such as a previous infection with tuberculosis (1) or pulmonary inflammatory
56 diseases, can leave patients with poor lung function, and extensive and permanent lung structural
57 change (2-4).

58 Systemic triazole therapy is the basis for treating infections with pathogenic fungi but the
59 adverse effects of itraconazole (ITC), voriconazole (VRC) and posaconazole (POS) are well
60 characterised and thought to be a consequence of the pharmacological effects of the compounds
61 in host tissues (5-9). It has been observed that up to 15% of patients treated with voriconazole
62 experience raised transaminase levels in the liver, a site of triazole toxicity (10, 11). Serious
63 unwanted effects in other organs have been reported after oral or systemic VRC and POS
64 treatment, and exposure of the liver also results in significant drug interactions arising from
65 triazole inhibition of hepatic P450 enzymes (12, 13), although recent azoles isavuconazole and
66 VT1161 showed better risk-benefit profiles in clinical or preclinical tests (14, 15).

67 Administration of triazoles orally can lead to wide variations in patient response due to
68 variable plasma concentrations, leading to compromised individual efficacy (16). Furthermore
69 notable drug interactions for voriconazole due to the inhibition of hepatic P450 enzymes make
70 clinical use challenging and indeed the variability in exposure of the triazoles via the oral route
71 necessitates the need for close therapeutic drug monitoring and limits the use of triazole therapy
72 prophylactically in at risk groups (13, 16). In addition, structural changes in the lung architecture,
73 caused by chronic pulmonary disease or infection with tuberculosis, can lead to *Aspergillus*

Revision, AAC01941-17 (Research Article, T.Colley/G.Sehra)

74 colonisation of pre-existing cavities, limiting the efficacy of orally administered compounds
75 which often struggle to penetrate into the pulmonary epithelial lining fluid (17). It is
76 acknowledged that targeted administration to the lung, the primary point of infection, would
77 prolong lung tissue residence and reduce systemic exposure, to display a better risk-benefit ratio.
78 Recently, existing antifungal medications such as AMB, VRC and ITC have been repurposed in
79 this manner to effectively prevent invasive disease (18-20).

80 In this report we disclose the *in vitro* and *in vivo* activities of a newly discovered azole
81 class, antifungal agent: 4-(4-(4-(((3*R*,5*R*)-5-((1*H*-1,2,4-triazol-1-yl)methyl)-5-(2,4-
82 difluorophenyl)tetrahydrofuran-3-yl)methoxy)-3-methylphenyl)piperazin-1-yl)-*N*-((1*S*,2*S*)-2-
83 hydroxycyclohexyl)benzamide, (referred to here as PC1244, Fig 1) (21). The compound
84 demonstrates activities comparable to POS and superior to VRC against both ITC susceptible
85 and resistant strains, and it has been designed to have physicochemical properties suitable for
86 topical administration to the lung and promote long lasting tissue residency.

AAC (Research Article, T.Colley/G.Sehra)

87 **RESULTS**

88 ***In vitro* antifungal activity against laboratory adopted strains of *A. fumigatus*.** The
89 antifungal activity of test compounds against *A. fumigatus* strains (Itraconazole (ITC)
90 susceptible-NCPF2010, AF294, AF293; ITC-resistant-AF72, AF91) were calculated from
91 growth curves generated by spectrophotometric analysis and compared to positive and negative
92 controls.

93 It was observed that significantly lower concentrations of PC1244 were needed for endpoints
94 (50% inhibition (IC₅₀ [OD]) and 90% inhibition (IC₉₀ [OD]) than those of all reference
95 compounds, including voriconazole (VRC), posaconazole (POS) and itraconazole (ITC), against
96 ITC-susceptible *A. fumigatus* laboratory strains (NCPF2010, AF294, AF293, Table 1) (22, 23).
97 In addition, PC1244 was the most active test agent against known ITC-resistant *A. fumigatus*
98 strains (AF72, AF91) (24, 25) (Table 1).

99

100 ***In vitro* antifungal activity against clinically isolated *A. fumigatus*.** The antifungal activity of
101 PC1244 was further evaluated in 96 clinical isolates [obtained from the Saint Louis Hospital,
102 Paris, France (50 isolates) and NW Mycology Centre, Manchester, UK (46 isolates)]. In this
103 study, PC1244 was found to be 6.2-fold more potent than VRC and demonstrated comparable
104 effects to POS based on their geometric means (Table 2, Fig 2). During this assay, the quality
105 control strain *A. fumigatus* ATCC204305 was used for validation and posaconazole showed a
106 MIC of 0.25 µg/ml, within the range set by the EUCAST guidelines.

107

108 ***In vitro* assessment of antifungal activity using CLSI methodology.** *A. fumigatus* growth
109 inhibition by PC1244 was confirmed by the CLSI method as well as the EUCAST microdilution

AAC (Research Article, T.Colley/G.Sehra)

110 method. Following the CLSI methodology guidelines (M38-A (26)), the growth of four ITC-
111 susceptible laboratory *A. fumigatus* strains was assessed visually. PC1244 generated a MIC
112 (0.063 $\mu\text{g/ml}$) which was comparable to POS (0.047 $\mu\text{g/ml}$) and 8-fold more potent than VRC
113 (0.5 $\mu\text{g/ml}$).

114

115 **CYP51 binding properties.** Both PC1244 and POS produced type II difference spectra when
116 titrated against purified recombinant *A. fumigatus* (AF293) CYP51A and CYP51B enzymes.
117 PC1244 bound with similar affinities to the two isoenzymes as POS (CYP51A, K_d values 0.74
118 and 0.96 μM for PC1244 and POS, respectively; CYP51B, K_d values of 0.018 and 0.012 μM ,
119 respectively) (Fig 3A, B). The low-end accuracy limit for K_d determinations using the modified
120 Morrison equation is ~0.5 to 1% of the enzyme concentration (27), i.e. 0.020 to 0.040 μM in this
121 study. Consequently, calculated K_d values below 0.020 μM should be treated numerically as
122 <0.020 μM .

123

124 **Inhibitory activity against *A. fumigatus* CYP51 enzyme.** The inhibitory activities of PC1244
125 against *A. fumigatus* sterol 14 α -demethylases were determined using 0.5 μM *A. fumigatus*
126 CYP51A and 0.5 μM CYP51B in the membrane fraction prepared from *Escherichia coli*
127 expression clones, and compared to those of posaconazole. Both PC1244 and POS were strong
128 tight-binding inhibitors of CYP51A and CYP51B *in vitro* activity (CYP51A IC_{50} values for
129 PC1244 and POS: 0.27 and 0.16 μM ; and CYP51B IC_{50} values of 0.23 and 0.17 μM ,
130 respectively), suggesting $K_{i,\text{app}}$ values below 1 nM (27), with PC1244 being equally as effective
131 as POS (Fig 3C, D). This data suggests that both agents share the same mode of action; by
132 directly coordinating as the sixth axial ligand of the CYP51 heme iron.

AAC (Research Article, T.Colley/G.Sehra)

133

134 **Cell based *A. fumigatus* sterol composition and CYP51 assay.** The sterol composition of *A.*
135 *fumigatus* (NCPF2010) was determined for cells treated with 0, 0.0001, 0.001, 0.01, 0.1, and 1
136 $\mu\text{g/ml}$ PC1244 and posaconazole. Sterols were extracted by saponification with KOH followed
137 by extraction with n-hexane and TMS-derivatisation prior to analysis by GC/MS. Azole
138 treatment of *A. fumigatus* resulted in the dose dependent accumulation of the 14 α -methylated
139 sterols (lanosterol and eburicol) and the corresponding depletion of the final sterol product:
140 ergosterol (Table 3), characteristic of cellular CYP51 inhibition.

141 CYP51 enzyme inhibitory activity was also measured in a cell-based assay, as described
142 previously (28). In this plate-based ergosterol quantification experiment, oxidation of ergosterol
143 by cholesterol oxidase was determined by observing the conversion of the weakly fluorescent
144 resazurin to the highly fluorescent resorufin, and was normalised using crystal violet staining
145 (indicating cell number). Mirroring the inhibitory activity observed in the cell-free model of
146 CYP51 and the sterol profiles of treated cells, PC1244 strongly inhibited ergosterol production
147 ($\text{IC}_{50} = 0.0055 \mu\text{g/ml}$; $0.0080 \mu\text{M}$) and was 12-fold more potent than VRC ($\text{IC}_{50} = 0.067 \mu\text{g/ml}$;
148 $0.19 \mu\text{M}$) and 2.2-fold more potent than POS ($\text{IC}_{50} = 0.012 \mu\text{g/ml}$; $0.017 \mu\text{M}$).

149

150 ***In vitro* determination of persistence of action.** The duration of action of test agents within the
151 hyphae of *A. fumigatus* has been determined using a resazurin-based microtiter assay (28). *A.*
152 *fumigatus* hyphae were exposed to test agents for 16 h and the inhibition of fungal growth was
153 measured, and the efficacy was compared with that obtained after contact with drug for only 20
154 minutes, followed by washout and incubation for the same period. As seen in Table 4, it was
155 observed that PC1244 (IC_{50} : $0.00011 \mu\text{g/ml}$) was 100 and 4.1-fold more potent than VRC and
156 POS, respectively, at inhibiting hyphal *A. fumigatus* growth. In addition, the potency of VRC and

AAC (Research Article, T.Colley/G.Sehra)

157 POS diminished markedly, after short contact and washout, by factors of >93-fold and 4.9-fold,
158 respectively. In contrast, washout produced only a 2.4-fold reduction in the activity of PC1244
159 compared with continuous contact in this experimental paradigm (Table 4, Fig 4A and B).

160 In a second system, the persistence of action of the same three agents on *A. fumigatus*-
161 infected bronchial epithelial cells was quantified using GM production in the culture supernatant
162 as an index of fungal growth. BEAS2B cells were infected with *A. fumigatus*, and the effects of a
163 24 h washout period (media change) prior to infection were examined. A one hour contact time
164 with PC1244 followed by 24 h washout resulted in a 5.4-fold loss of potency against *A.*
165 *fumigatus*, compared with the control where there was no washout. POS showed a greater loss of
166 its activity on washout (14.7-fold) and it was particularly notable that VRC was ineffective under
167 the same test conditions (Table 4, Fig 4C and D). The pattern of effects mirror those seen in *A.*
168 *fumigatus* hyphae (above) and imply that only a short contact period of bronchial epithelial cells
169 with PC1244 would be required for the agent to exert a long duration of therapeutic action.

170
171 ***In vitro* fungicidal activity against *A. fumigatus*.** The MFC for each compound was calculated
172 48 h after supernatants from the MIC assay were transferred to agar plates, and determined to be
173 the lowest concentration of compound that yielded 3 colonies or less (CFU-MFC). PC1244
174 exhibited the greatest CFU-MFC of all compounds tested (2 µg/ml), and was 2 and 8-fold
175 stronger than POS and VRC, respectively. The ratios of CFU-MFC versus MIC were 32, 32 and
176 9.6 for PC1244, POS and VRC, respectively (Table 5).

177 In addition, the fungicidal effect of each compound was determined using an XTT based
178 quantitative colorimetric analysis (XTT-MFC). Absorbance was measured at OD₄₅₀₋₆₂₀, 24 h after
179 supernatant including compound was removed from the wells used for broth microdilution MIC
180 assay. Again, PC1244 exhibited the greatest level of inhibition of all compounds tested with a

AAC (Research Article, T.Colley/G.Sehra)

181 XTT-MFC of 0.14 $\mu\text{g/ml}$. In this system, PC1244 was 3-fold more potent than POS and >229-
182 fold more potent than VRC. The maximum inhibition of fungicidal activities for PC1244, POS
183 and VRC were 99.9% at 1 $\mu\text{g/ml}$, 100% at 2 $\mu\text{g/ml}$ and 69.2% at 32 $\mu\text{g/ml}$, respectively. In
184 addition, the ratios of XTT-MFC versus MIC were 2.2, 3.4 and >19 for PC1244, POS and VRC,
185 respectively (Table 5, Fig 5).

186

187 ***In vivo* antifungal activity on ITC susceptible *A. fumigatus* infection.** To assess the *in vivo*
188 activity of PC1244, temporarily neutropenic mice infected with *A. fumigatus* (ATCC13073
189 [strain: NIH 5233]) were used. MIC values of PC1244, POS, VRC and ITC against this strain
190 were 0.063, 0.125, 0.5 and 0.5 $\mu\text{g/ml}$, respectively. An aqueous suspension of PC1244 in
191 isotonic saline (0.0032, 0.016 and 0.08 mg/ml, 35 μl , please see the table 6 for conversion to
192 mg/mouse or approximately mg/kg) was dosed by intranasal injection once daily for 3 days post
193 infection with ITC-susceptible *A. fumigatus*. This “late intervention” regimen was found to
194 strongly inhibit fungal load (CFU) in the lung, the highest dose (0.08 mg/ml) administered
195 exhibiting 97% inhibition, when compared to vehicle (Fig 6A). In comparison, POS, given at the
196 same level of 0.08 mg/ml, achieved only 39% inhibition of lung fungal load and the $\text{ID}_{\text{Log}10}$ value
197 (the dose to reduce 1 log₁₀ of CFU/g) was 2.0 mg/ml (70 $\mu\text{g/mouse}$) which was 143-fold higher
198 than that of PC1244 ($\text{ID}_{\text{Log}10}$: 0.014 mg/ml (0.49 $\mu\text{g/mouse}$)). PC1244 also decreased GM
199 concentrations in BALF in a dose-dependent manner, showing ID_{50} value of 0.032 mg/ml (1.1
200 $\mu\text{g/mouse}$), which was 6.7fold lower than that of POS (ID_{50} : 0.21 mg/ml (7.4 $\mu\text{g/mouse}$)) (Fig
201 6B). PC1244 decreased GM concentrations in serum in a dose-dependent manner, too (Fig 4C).
202 Notably, 0.08 mg/ml (2.8 $\mu\text{g/mouse}$) of PC1244 produced marked inhibition (82% inhibition) of
203 GM in serum, whereas POS, at the same dose, did not show any effect (-11% inhibition). In pilot
204 study, we also measured *A. fumigatus* PCR products in lung tissue, and consistent with these data

AAC (Research Article, T.Colley/G.Sehra)

205 above, PC1244 inhibited the accumulation of PCR product(Supplement Fig 1B). In addition,
206 PC1244 also reduced *A.fumigatus* infection dependent increase in CXCL1 in BALF (Fig 6D),
207 IL-6 (Fig 6E) and TNF α levels (Fig 6E4D) in serum.

208 Extended prophylaxis with PC1244 [0.0032 mg/ml (0.11 μ g/mouse); -7/0] achieved a
209 high level of inhibitory effects on fungal load and biomarkers when compared to that observed in
210 the late intervention study (Day 1-3 treatment) at the same dose (Fig 6A vs. 7A, Fig 6C vs. Fig
211 7B). Furthermore, a marked difference between extended prophylaxis (-7/0) and a shorter period
212 of treatment (-1/0) was also observed on CFU in lung, galactomannan in serum and MDA
213 (malondialdehyde, an oxidative stress marker) in BALF (Fig 7A, B and C).

214 As we indicated persistent action of PC1244 in the *in vitro* system eralier, the persistent
215 action was also evaluated *in vivo*. As the MIC value on the *A. fumigatus* strain (ATCC13073)
216 used in this study was 2-fold lower in PC1244 than POS, PC1244 at 0.4 mg/ml (14 μ g/mouse)
217 and POS at 0.8 mg/ml (28 μ g/mouse) were intranasally administered 16 h before *A. fumigatus*
218 inoculation, and the lungs were collected for GM and CFU assessment 8 h after *A. fumigatus*
219 inoculation. As observed in Fig. 8A and B, PC1244 showed significant inhibition on both GM
220 and CFU in the lung, but POS did not despite of administration at a 2-fold higher dose. Thus, in
221 the *in vivo* system, persistent action of PC1244 was confirmed.

222

223 **Antifungal activity against non-*A. fumigatus* species.** The *in vitro* activity of PC1244 was
224 compared with VRC, and POS against 23 pathogenic fungi (1~2 isolates each) and the results are
225 displayed in Table 7. In all non-*Aspergillus* and non-*Candida* species tested, PC1244 was more
226 potent or comparable in potency to POS and VRC. Of particular note, PC1244 was effective
227 (MIC, 0.25 – 2 μ g/ml) against species in which VRC and POS had no effect within the
228 concentration range tested (MIC, >8 μ g/ml), including *Gibberella zeae* (*Fusarium*

AAC (Research Article, T.Colley/G.Sehra)

229 *graminearum*), *Lichtheimia corymbifera*, *Mucor circinelloides*, *Rhizomucor pusillus* and
230 *Rhizopus oryzae*. PC1244 was found to have antifungal activity against *A. flavus*, *A. niger*, and *A.*
231 *terreus*, albeit with less potency than POS. Against *Aspergillus carbonarius*, PC1244 was
232 equally potent to POS and more potent than VRC. Against all *Candida* species tested (*Candida*
233 *albicans*, *Candida glabrata*, *Candida krusei* and *Candida parapsilosis*), PC1244 was more
234 potent than VRC, and stronger or comparable in potency to POS in its inhibitory activity.

235

236 **DISCUSSION**

237 In this report, we present data demonstrating that: 1. the novel triazole PC1244 possesses both
238 potent and persistent antifungal activity and significant fungicidal activity against ITC
239 susceptible and/or ITC resistant *A. fumigatus in vitro*, 2. the antifungal activity was confirmed in
240 clinical isolates from two geographical areas, 3. intranasal once-daily PC1244 treatment
241 exhibited potent antifungal effects against *A. fumigatus in vivo*, in temporarily neutropenic mice,
242 4. PC1244 showed a broad range of antifungal activity when screened against a panel of
243 pathogenic fungal organisms.

244 The proposed mechanism of action of PC1244 is inhibition of sterol 14 α -demethylase
245 (CYP51A1), the enzyme required to convert eburicol to 14-demethylated eburicol, an essential
246 step in the ergosterol biosynthesis pathway in fungi. Type II binding spectra, which display an
247 A_{\max} at 423-430 nm and a broad trough at 386-412 nm arise through a specific interaction in
248 which the triazole *N*-4 nitrogen (posaconazole) or the imadazole ring *N*-3 nitrogen coordinates as
249 the sixth axial ligand with the heme iron to form a low-spin CYP51-azole complex (29, 30).
250 PC1244 produced type II difference spectra when titrated against purified recombinant ITC-
251 susceptible *A. fumigatus* (AF293) CYP51A and CYP51B, and bound with a similar affinity to

AAC (Research Article, T.Colley/G.Sehra)

252 both enzymes as posaconazole. Furthermore, the strong inhibition of CYP51A activity observed
253 with both PC1244 and posaconazole, characteristic of tight-binding inhibitors (IC_{50} value
254 approximately half that of the enzyme concentration present), exceeded that predicted by the
255 calculated K_d values from ligand binding studies using recombinant CYP51A, suggesting that the
256 conformation of purified CYP51A in solution differs from that in cell membranes.

257 In sterol composition determinations, treatment with increasing concentrations of either
258 PC1244 or posaconazole, from 0 to 1 $\mu\text{g/ml}$, resulted in an accumulation of the 14-methylated
259 sterols, lanosterol and eburicol, and depletion of the final sterol product, ergosterol; this pattern
260 of effect is consistent with CYP51 inhibition being the key pharmacological activity of both
261 agents. In addition, a cell-based assay of ergosterol biosynthesis in *A. fumigatus* demonstrated
262 that PC1244 was 12 and 2.2-fold more potent at inhibiting ergosterol production than
263 voriconazole and posaconazole, respectively. Thus, the mechanism of action of PC1244, as for
264 other triazole antifungals, is the inhibition of fungal sterol 14α -demethylase, resulting in the
265 depletion of ergosterol in the fungal membrane so disrupting membrane structure and function
266 and inhibiting growth of the pathogenic organism (31).

267 A highly desirable feature of topical medicines is a long duration of action ensuring that
268 the desired therapeutic activity is maintained throughout the inter-dose period. This is
269 particularly relevant to the treatment of pulmonary infection with *A. fumigatus*, which germinates
270 in both extracellular environments and intracellular compartments. The duration of action of
271 PC1244 was therefore considered an important property and was evaluated in a variety of *in vitro*
272 systems. In *A. fumigatus* hyphae, the IC_{50} value measured for PC1244 following a 20 minute
273 contact period and washout for 16 h was reduced only 2.4-fold relative to that obtained following
274 continuous contact with the drug for the same period without washout. Furthermore, in the
275 BEAS2B cell line, washout for 24 h, after a 1 h contact period, resulted in only an approximate

AAC (Research Article, T.Colley/G.Sehra)

276 5-fold loss of potency against *A. fumigatus* compared with control. These observed properties of
277 rapid cellular penetration and persistence of action, in the absence of the pathogen, may be
278 particularly valuable characteristics, which enhance the potential use of PC1244 in prophylaxis.
279 The persistent action of PC1244 was also confirmed in the *in vivo* system when administered 16
280 h before *A. fumigatus* inoculation (Figure 5).

281 In the *in vivo* system, intranasal treatment of PC1244 showed better effects than
282 posaconazole despite comparable MIC values in *in vitro* testing. We speculate that, firstly,
283 persistence of the drug substance on bronchial cells, as shown in Figure 4, is likely to be a
284 contributory factor to the amplification of the antifungal effects of PC1244 *in vivo* seen in the
285 current once daily treatment regimen. Secondly, we have demonstrated here that a 7 day
286 extended prophylactic treatment (using very low doses) produced much greater anti-*Aspergillus*
287 activity than prophylactic treatment for 1 day, and also that the effects of 7 day prophylactic
288 treatment are maintained if treatment ceased when *Aspergillus* is inoculated on day 0 (Fig. 7).
289 This is powerful pharmacodynamic evidence that the effects of PC1244 accumulate on daily
290 dosing in mice and are maintained when dosing is terminated. Thirdly, Baistrocchi and
291 colleagues published evidence of the accumulation of posaconazole in granulocyte type cells and
292 demonstrated enhanced synergic antifungal effects (by exposure of *Aspergillus* to cellular
293 posaconazole during phagocytosis) (32). Considering the persistent action of PC1244, it is likely
294 that granulocytes/macrophages containing PC1244 contributed to further enhancement of the
295 antifungal effect.

296 Determining whether an antifungal compound is “fungistatic” or “fungicidal” is complex
297 and the clinical utility of such characterisation is the subject of much debate. Fungal infections of
298 body compartments that are not easily accessed by host defences require agents that are
299 fungicidal in nature, and this is especially true in immunocompromised patients (33). For an

AAC (Research Article, T.Colley/G.Sehra)

300 antibiotic to qualify as bactericidal its minimum bactericidal concentration (MBC) must be no
301 more than 2× to 4× the MIC, but the definition of “fungicidal” is yet to be standardised (33).
302 MFC determination in filamentous fungi is not standardised either, but studies have shown that
303 reproducible MFCs can be obtained by following standardised broth microdilution methods for
304 MIC determination, followed by subculture onto agar (33, 34). Defining an MFC as the lowest
305 drug dilution to yield less than 3 colonies to obtain 99% - 99.5% killing activity, Espinel-Ingroff
306 *et al* determined the MFC₉₀ range of itraconazole (0.2 – 4 µg/ml), voriconazole (0.5 – 4 µg/ml)
307 and posaconazole (0.06 – 2 µg/ml), in a number of *A. fumigatus* isolates (34). However, it is
308 worth noting that the agar subculture methodology tests for fungicidal activity on planktonic Af
309 growth only. Here we have used a combination of different methodologies to attempt to
310 determine the fungicidal activity of PC1244 and clinically used triazoles accurately. Using
311 subculture on agar (CFU-MFC) gave an MFC of 2 µg/ml for PC1244 with an MFC/MIC ratio of
312 32, and similar results were seen with posaconazole (MFC = 4 µg/ml, MFC/MIC = 32), whilst
313 voriconazole exhibited a higher MFC (16 µg/ml) but a lower MFC/MIC ratio (9.6). As discussed
314 above, this data would suggest voriconazole is a more fungicidal compound as it exhibits a lower
315 MFC/MIC ratio. However, recent studies have shown that this technique likely overestimates the
316 fungicidal activity of a compound as it does not factor in viable conidia attached to the base of
317 test wells (35). To account for this phenomenon, a colorimetric method for assessing the
318 fungicidal activity of a compound against sessile *A. fumigatus* was used (35). MFC determination
319 by this microbroth colorimetric method (XTT-MFC) gave an MFC of 0.14 µg/ml for PC1244
320 with an MFC/MIC ratio of 2.2, which was superior to both posaconazole (MFC = 0.42 µg/ml,
321 MFC/MIC = 3.4) and voriconazole (MFC = >32 µg/ml, MFC/MIC = >19). Therefore, with these
322 data we provide evidence that PC1244 is a fungicidal compound with a similar or improved
323 degree of potency to posaconazole.

AAC (Research Article, T.Colley/G.Sehra)

324 As with any study there are limitations, especially the *in vivo* study. Firstly, the delivery
325 system does not mimic clinical use. The advantage of intranasal instillation is being able to
326 confirm that all the solution is delivered into the body, but we do not control the level of lung
327 exposure or the exposure site (same as aerosolization). However, we carefully optimised the
328 intranasal dosing volume, as it has been shown that approximately 60% of the administered dose
329 will be deposited in the lung after intranasal treatment (36) and also we confirmed trachea/lung
330 deposition after intranasal administration of 35 μ L of methylene blue solution to A/J mice (data
331 not shown). Aerosolization with close drug monitoring at the exposure site (rather than systemic)
332 is ideal, but this is not easily achieved as special imaging equipment is required, as Miller *et al*
333 demonstrated elegantly using whole-Animal Luminescent Imaging (37). Secondly, CFUs were
334 determined in only whole left lobe. This will cause location bias of fungal load, and ideally we
335 should test in homogenate from the whole lung. However, when we determined CFU and GM in
336 right lobe and left lobe, we did not find any significant difference of these biomarkers between
337 right lobe and left lobe (data not shown). In addition, to avoid this bias, we determined GM in
338 BALF and serum. Thirdly, there was a lack of pharmacokinetic measurement of PC1244 in mice
339 used for the *in vivo* study. PC1244 has been optimised for topical treatment to the lung to
340 maximize local exposure and minimise systemic exposure. Systemic concentrations of drug are
341 therefore not a useful surrogate marker to help explain the different antifungal efficacy of
342 compounds. However, we have some data demonstrating that measurable levels of systemic
343 exposure do occur. In preliminary studies using non-infected mice dosed intra-tracheally with 40
344 μ L of a 2 mg/ml aqueous suspension of PC1244, it was shown that the plasma concentrations of
345 PC1244 ranged from 111 ~ 303 ng/ml 2 hours post dose, and 249 ~ 339 ng/ml 8 hours post dose.
346 But this had been reduced to 41.5 ~ 50.7 ng/ml 24 hours post dose despite decent *in vivo* effects
347 after once daily treatment. Under the same conditions, the plasma concentrations achieved with

AAC (Research Article, T.Colley/G.Sehra)

348 posaconazole were 15.6 ~ 125 ng/ml at 24 hours post dose, which was more variable but not too
349 different from that of PC1244, although the *in vivo* activity of PC1244 was superior to that of
350 posaconazole. As PC1244 has much less oral availability compared with posaconazole
351 (unpublished data), the exposure results from absorption through the respiratory tract (not by
352 accidental ingestion of compound during dosing). Furthermore, all compounds are water
353 insoluble, and administered topically (exposed to respiratory tract directly). We do not, therefore,
354 believe that water solubility is a key factor explaining the *in vivo* efficacies or topical exposure
355 levels. Finally, we tested only limited isolates of azole resistant strains. Further study will be
356 required with a wide range of clinical azole resistant isolates with different genotypes to
357 determine its potency against recent *A. fumigatus* strains with TR₃₄/L98H and
358 TR₄₆/Y121F/T289A mutations or other genetic cause(s) underlying resistance.

359 Thus, due to its superior or comparable activity against both azole susceptible and azole
360 resistant *A. fumigatus*, persistent action, extended retention within the lung after topical treatment
361 and broad repertoire of fungal targets, PC1244 has the potential to become a valuable new
362 therapeutic agent for the treatment of *A. fumigatus* and other, difficult to treat, fungal infections
363 in man.

AAC (Research Article, T.Colley/G.Sehra)

364 **MATERIALS & METHODS**

365 **Antifungal agents.** PC1244 was synthesised by Sygnature Discovery Ltd (Nottingham, UK),
366 and voriconazole (Tokyo Chemical Industry UK Ltd., Oxford, UK), posaconazole (Apichem
367 Chemical Technology Co., Ltd., Zhejiang, China), itraconazole (Arkopharma, Carros, France),
368 amphotericin B (Selleckchem, Munich, Germany) and caspofungin (Selleckchem, Munich,
369 Germany) were procured from commercial sources. For *in vitro* antifungal assays, stock
370 solutions of test agents were prepared in DMSO (2000 µg/ml). For *in vivo* studies, solid materials
371 of test agents were directly suspended in physiological saline at 10 mg/ml, and diluted with
372 physiological saline after sonication.

373

374 ***A. fumigatus* CYP51 binding assay and enzyme inhibitory activity.** *A. fumigatus* CYP51
375 binding properties were determined as previously reported (28, 38). Test agents were titrated
376 against 4 µM recombinant *A. fumigatus* (AF293 strain) CYP51A or CYP51B proteins and
377 binding saturation curves were constructed from the change in the absorbance between the
378 spectral peak and the trough. A rearrangement of the Morrison equation was used to determine
379 the dissociation constant (K_d) values when ligand binding was tight (39).

380 A CYP51 reconstitution assay system was used to determine 50% inhibitory (IC_{50})
381 concentrations (40). Test agent was added to a mixture of 0.5 µM CYP51, 1 µM *A. fumigatus*
382 cytochrome P450 reductase isoenzyme 1 (AfCPR1), 50 µM eburicol, 4% (w/v) 2-hydroxypropyl-
383 β-cyclodextrin, 0.4 mg/ml isocitrate dehydrogenase, 25 mM trisodium isocitrate, 50 mM NaCl, 5
384 mM MgCl₂ and 40 mM 3-(*N*-morpholino) propanesulfonic acid (MOPS; pH ~7.2). The mixtures
385 were then incubated at 37°C for 10 minutes prior to initiation with 4 mM β-NADPHNa₄
386 followed by shaking for 20 minutes at 37°C. Sterol metabolites were recovered by extraction

AAC (Research Article, T.Colley/G.Sehra)

387 with ethyl acetate followed by derivatisation with 0.1 ml *N,O*-
388 bis(trimethylsilyl)trifluoroacetamide : trimethylchlorosilane (99:1) and 0.3 ml anhydrous
389 pyridine prior to analysis by gas chromatography mass spectrometry.

390

391 ***A. fumigatus* sterol analysis.** A working suspension of *A. fumigatus* spores (NCPF2010) was
392 prepared in filter-sterilised MOPS RPMI-1640 (RPMI-1640 containing 2 mM L-glutamine, 2%
393 glucose, 0.165 M MOPS, buffered to pH 7 with NaOH) at a final concentration of 8×10^6 spores
394 ml^{-1} . To each 100 mm Petri dish, 10 ml of the working suspension was added and the dishes
395 were incubated for 4 h at 35°C and 5% CO_2 . Samples for baseline determinations were collected
396 by scraping, pelleted by centrifugation at 2000 rpm for 5 minutes and stored at -80°C. Test
397 compounds or DMSO (50 μl) were added to the remaining dishes, which were subsequently
398 gently rocked by hand to disperse the compounds. Dishes were incubated for 2 h at 35°C and 5%
399 CO_2 . Samples were collected and processed as described above. Posaconazole and PC1244
400 concentrations of 0.0001, 0.001, 0.01, 0.1 and 1 $\mu\text{g ml}^{-1}$ were tested. These samples were
401 prepared in the laboratory at Pulmocide Ltd., and sent to the laboratory in the Centre for
402 Cytochrome P450 Biodiversity, Institute of Life Science, Swansea University Medical School,
403 for analysis.

404 Non-saponifiable lipids were extracted as previously reported (31) and were derivatised
405 with 0.1 ml *N,O*-bis(trimethylsilyl)trifluoroacetamide : trimethylchlorosilane (99:1) and 0.3 ml
406 anhydrous pyridine (2 h at 80°C) prior to analysis by gas chromatography mass spectrometry
407 (41). Sterol composition was calculated using peak areas from the gas chromatograms and the
408 mass fragmentation patterns compared to known standards were used to confirm sterol identity.
409 The sterol content of *A. fumigatus* (basal) and treated *A. fumigatus* (either DMSO, posaconazole
410 or PC1244) were determined in three biological replicates.

AAC (Research Article, T.Colley/G.Sehra)

411

412 ***A. fumigatus* cell based ergosterol assay.** Growth medium (RPMI-1640, 2 mM L-glutamine,
413 2% glucose, 0.165 M MOPS, 0.5% BSA, pH 7.0) was added across a 96-well plate and test
414 agents were added in duplicate. *A. fumigatus* (NCPF2010) conidia were added across the plate at
415 a final concentration of $1 \times 10^4 \text{ ml}^{-1}$. After incubation for 24 h at 35°C, media was removed from
416 all wells and replaced with reaction buffer (Amplex red cholesterol assay kit, ThermoFisher,
417 A12216) and Amplex red solution. Plates were incubated for 30 minutes at 37°C, protected from
418 light, after which fluorescence was quantified using a spectrophotometer. Media was removed
419 from all wells and replaced with crystal violet solution (1% v/v), and plates were incubated at
420 room temperature on a shaker for 30 minutes. Plates were washed three times with PBS, and
421 sodium dodecyl sulphate solution (0.1% v/v) was added across the plate to lyse the cells. After
422 incubation at room temperature for 1 h, absorbance was measured at OD₅₉₀ using a
423 spectrophotometer.

424

425 ***In vitro* antifungal activity against *A. fumigatus*.** Assessment of antifungal activity against a
426 selection of *A. fumigatus* laboratory/clinical strains (NCPF2010 [National Collection of
427 Pathogenic Fungi (NCPF), Bristol, UK], AF72 [NCPF, Bristol, UK], AF91 [NCPF, Bristol, UK],
428 AF293 [NCPF, Bristol, UK], AF294 [NCPF, Bristol, UK]) was performed using EUCAST
429 methodology as previously reported (28), in a 384-well plate format as quadruplicates, in three
430 independent experiments. Growth medium (RPMI-1640, 2 mM L-glutamine, 2% glucose, 0.165
431 M MOPS, 0.5% BSA, pH 7.0) was added across the plate, test agents were added in
432 quadruplicate and the DMSO concentration was identical across the plates. Conidia were added
433 across the plate at a final concentration of $1 \times 10^5 \text{ ml}^{-1}$. Plates were incubated for 48 h at 35°C
434 after which turbidity was assessed by measuring optical density (OD) at 530 nm using a

20

AAC (Research Article, T.Colley/G.Sehra)

435 spectrophotometer, and the IC₅₀ and IC₉₀ values were calculated from the concentration-response
436 curve generated for each test compound using a four-parameter logistic equation (Dotmatics,
437 Bishops Stortford, UK). *A. fumigatus* ATCC204305 was used as the assay control.
438 Determination of antifungal activity against 50 *A. fumigatus* clinical isolates from St Louis
439 Hospital (Paris, France) was performed with 96-well plates using the EUCAST method shown
440 above (28) in duplicate. Antifungal susceptibility testing for 46 *A. fumigatus* isolates [obtained
441 from the North West England Mycology Reference Centre] was performed as singlicate by
442 Evotec (UK) Ltd (Manchester, UK) according to EUCAST guidelines. Assessment of the
443 antifungal activity of four of the *A. fumigatus* strains (ATCC1028, ATCC10894, ATCC13073,
444 and ATCC16424) was performed as singlicate according to M38-A methodology described by
445 the Clinical and Laboratory Standards Institute (CLSI) (26) by Eurofins Panlabs Taiwan Ltd.
446 (Taipei, Taiwan).

447

448 ***In vitro* antifungal activity against other fungal species.** For the measurement of activity
449 against *C. gattii*, the method described in EUCAST definitive document EDef 7.2 was used and
450 assay plates were incubated statically at 37°C in ambient air for 24 h (\pm 2 h) unless poor growth
451 necessitated further incubation to 36 or 48 h (42). Antifungal potency against *Aspergillus flavus*,
452 *Aspergillus niger* and *A. terreus*, was determined as set out in EUCAST definitive document
453 EDef 9.2 and assay plates were incubated at 37°C for 48 h (43). These tests were conducted at
454 Evotec (UK) Ltd (Manchester, UK). Measurement of activity against other fungi was performed
455 by Eurofins Scientific according to methodology described by the Clinical and Laboratory
456 Standards Institute (CLSI) (CLSI M38-A (26) or M27-A2 (44), www.eurofinspanlabs.com). The
457 source or strain name of each fungus species was indicated in Table 6. The MIC against *C.*

AAC (Research Article, T.Colley/G.Sehra)

458 *albicans*, *C.parapsilosis* and *C.glabrata* were determined using an azole endpoint, which
459 indicates 50% inhibition of fungus growth.

460

461 ***In vitro* fungicidal activity of PC1244 against *A. fumigatus*.** The antifungal activity of PC1244
462 against *A. fumigatus* [NCPF2010] was determined in 96-well plates using the methodology
463 described above as duplicates, in three independent experiments. After the MIC for each
464 compound was recorded, fungicidal activity was determined as previously described (35).
465 Briefly, media from each well (100 μ l/well) was removed after pipetting up and down five times
466 and sub-cultured onto 4% Sabouraud dextrose agar plates. The plates were incubated (35°C with
467 ambient air) for 48 h and the colony forming units (CFU) were counted for each compound
468 concentration. The minimum fungicidal concentration (MFC) was determined as the lowest
469 compound concentration yielding 3 colonies or less.

470 After removal of media for culture-based CFU testing, the contents of all wells were
471 carefully aspirated and warm PBS (200 μ l/well) was added. After gentle agitation, the contents of
472 all wells were aspirated and fresh medium added (200 μ l/well). The plates were incubated (35°C
473 with ambient air) for 24 h. A working solution of 0.5 mg/ml XTT and 125 μ M menadione was
474 prepared in PBS and added across the plate (50 μ l/well). The plates were incubated (35°C with
475 ambient air) for 2 h, after which plates were agitated gently for 2 min. The optical density (OD)
476 of each well at 450 nm and 620 nm was measured using a multi-scanner (Clariostar: BMG,
477 Buckinghamshire, UK). The MFC (XTT-MFC) was calculated from the concentration-response
478 curve generated using a cut-off of 99% inhibition.

479

480 ***In vitro* determination of persistence of action on *A. fumigatus* hyphae.** The persistence of
481 action of test agents was determined in *A. fumigatus* hyphae (NCPF2010) as previously reported

AAC (Research Article, T.Colley/G.Sehra)

482 (28). Briefly, conidia diluted in growth media (RPMI-1640, 2 mM L-glutamine, 2% glucose,
483 0.165 M MOPS, pH 7.0) were added across a 384-well plate at a final concentration of
484 1×10^3 /well. After incubation at 35°C for exactly 6 h, test and reference articles or neat DMSO (as
485 vehicle) (0.5 μ l/well) were added to the appropriate wells to give a final concentration of 0.5%
486 DMSO. The plates were incubated for exactly 20 minutes at 35°C and 5% CO₂. After the
487 incubation time had elapsed all wells on the designated washout plate were aspirated and growth
488 media (100 μ l/well) was added across the plate. For the non-washout plate, after compounds
489 were added to hyphae, no media change was applied. Resazurin (0.04% diluted in growth
490 media) was added to all wells of both non-washout and washout plates (5 μ l/well) to give a final
491 concentration of 0.002% resazurin. The plates were incubated at 35°C and 5% CO₂ for 16 h.
492 Subsequently fluorescence in each well was measured at $\lambda_{\text{ex}}/\lambda_{\text{em}}$ 545/600 nm using a
493 multiscanner (Clariostar: BMG, Buckinghamshire, UK). The percentage inhibition for each well
494 was calculated and the IC₅₀ value was calculated from the concentration-response curve
495 generated for each test compound using a four-parameter logistic equation (Dotmatics, Bishops
496 Stortford, UK). This study was conducted in quadruplicates, in three independent experiments.

497

498 ***In vitro* determination of persistence of action on bronchial epithelial cells.** The persistence
499 of action of test agents was evaluated in immortalised, bronchial, epithelial cells (BEAS2B) as
500 previously reported (28). Each experiment consisted of one non-washout plate (96-well) and a
501 parallel washout plate into which BEAS2Bs were seeded at a concentration of 3×10^4 cells/well in
502 growth media (RPMI-1640, 2 mM L-glutamine, 10% FCS), and incubated for 24 h at 37°C, 5%
503 CO₂. Test and reference articles or neat DMSO (as vehicle) (0.5 μ l/well) were added to the
504 appropriate wells of the washout plate to give a final concentration of 0.5% DMSO. The plate
505 was incubated for exactly 1 h at 37°C and 5% CO₂. After the incubation time had elapsed all

AAC (Research Article, T.Colley/G.Sehra)

506 wells on the washout plate were aspirated and growth media (100 μ l/well) was added across the
507 plate. After 24 h incubation at 37°C, test and reference articles or neat DMSO (as vehicle) (0.5
508 μ l/well) were added to the appropriate wells of the non-washout plate to give a final
509 concentration of 0.5% DMSO. The plate was incubated for exactly 1 h at 37°C and 5% CO₂ after
510 which *A. fumigatus* conidia were added across both plates at a final concentration of 1 x 10³/well.
511 Fungal growth was determined after a further 24 h incubation at 35°C, 5% CO₂, by measuring
512 galactomannan (GM) concentrations, using Platelia GM-EIA kits (Bio-Rad Laboratories, 62794).
513 The percentage inhibition for each well was calculated and the IC₅₀ value was calculated from
514 the concentration-response curve generated for each test compound using a four-parameter
515 logistic equation (Dotmatics, Bishops Stortford, UK). This study was conducted in triplicates, in
516 three independent experiments.

517

518 ***In vivo* antifungal activity against *A. fumigatus* infection.** As previously reported (28), we
519 tested antifungal effects of test articles on *A. fumigatus* infected, temporarily neutropenic mice.
520 Specific pathogen-free A/J mice (male, 5 weeks old) were used for *A. fumigatus* infection as they
521 have been described to be more susceptible to *A. fumigatus* infection previously (45). Animals
522 (N=6 per group) were then dosed with hydrocortisone (Sigma H4881, 125 mg/kg,
523 subcutaneously) on days 3, 2 and 1 before infection, and with cyclophosphamide (Sigma C0768;
524 250 mg/kg, intraperitoneally) two days before infection to induce temporary neutropenia. Both
525 hydrocortisone and cyclophosphamide were diluted with physiological saline. To avoid bacterial
526 infection during immunosuppression, drinking water was supplemented with tetracycline
527 hydrochloride (Sigma T7660; 1 μ g/ml) and ciprofloxacin (Fluka 17850; 64 μ g/ml). Conidia of *A.*
528 *fumigatus* (ATCC13073 [strain: NIH 5233]) were aseptically dislodged from the malt agar plates

AAC (Research Article, T.Colley/G.Sehra)

529 and suspended in sterile distilled water with 0.05% Tween 80 and 0.1% agar. On the day of
530 infection, 30 μ l (15 μ l in each nostril) of the conidia suspension (1.67×10^8 /ml in physiological
531 saline) was administered intranasally under 3% isoflurane.

532 Test agents, suspended in physiological saline, were administered intranasally (35 μ l,
533 approximately 17.5 μ l each in each nostril), once daily, on days 1, 2 and 3 post infection. To
534 investigate extended prophylaxis, PC1244 was administered intranasally once daily, on days -7
535 to 0 and the effects were compared with treatment on days -1 to 0. As the injection volume was
536 fixed and body weight was changed every day, especially after infection, the accurate dose unit
537 was μ g/mouse. However, as the average body weight after immunosuppression and just before
538 infection was 20 g, we also calculated estimated dose as mg/kg. Therefore, 35 μ l injections of
539 0.0032, 0.016, 0.08, 0.4, 2 mg/ml were equivalent to 0.11, 0.56, 2.8, 14, 70 μ g/mouse,
540 respectively, which were approximately 0.0056, 0.028, 0.14, 0.7, 3.5 mg/kg, respectively (Table
541 6).

542 A body weight loss of > 20%, compared with an animal's weight on day 1, or a mouse death,
543 were both defined as "drop-out" events. Animals that lost > 20% of their initial body weight
544 were sacrificed. Animals were terminally anaesthetised 6 h after the last dose of drug was
545 administered on day 3. The volume inserted intranasally is reported to achieve almost 60%
546 deposition into the lung (36).

547 BALF was collected through cannulated tracheas using physiological saline (46), blood was
548 then collected via cardiac puncture, and lung tissue was removed for homogenate preparation.
549 The *Aspergillus* GM concentration in serum was determined with Platelia GM-EIA kits (Bio-Rad
550 Laboratories, 62794). The value was provided as a "cut-off index" (COI) which was calculated
551 by the formula: COI = OD in sample / OD in cut-off control, provided by the kit. For tissue
552 fungal load, 100 mg of whole left lobe of lung tissue was removed aseptically and homogenized

AAC (Research Article, T.Colley/G.Sehra)

553 in 0.2 ml of 0.1% agar in sterile distilled water as previously reported (28). We confirmed that
554 the CFU level was not significantly different between the right lung and left lung. Serially
555 diluted lung homogenates were plated on malt agar plates (50 µl/plate), and incubated at 24 ±
556 1°C for 72 to 96 h. The colonies of *A. fumigatus* on each plate were counted and the fungal titre
557 presented here as CFUs (x10³) per gram of lung tissue.

558 Measurement of TNFα and IL-6 in serum and CXCL1 in BALF was performed using
559 Quantikine® mouse ELISA kit (R&D systems, Inc., Minneapolis, MN, USA). MDA
560 (malondialdehyde) analysis was also performed using OxiSelect® TBARS Assay Kit (MDA
561 Quantitation; Cell Biolabs Inc, San Diego, CA, USA). For quantitative PCR, DNA amplification
562 was performed with Premix Ex Taq™ (Takara Bio, Kusatsu, Japan) and analysed in 96-well
563 optical reaction plates, using the standard curve method. *A. fumigatus* 18S rRNA gene fragments
564 were amplified with the primer pair; 5'-GGCCCTTAAATAGCCCGGT-3' and 5'-
565 TGAGCCGATAGTCCCCCTAA-3', and hybridization probe; 5'-FAM-
566 AGCCAGCGGCCCGCAAATG-TAMRA-3'. Each 25 µl reaction solution contained 50 ng of
567 DNA from mice lungs and 200 nM of probe. The PCR protocol was as follows: incubation at
568 50°C for 2 min and 95°C for 10 min; followed by 55 cycles of 65°C for 1 min and 95°C for 15
569 sec. The fluorescence was monitored at the end of each cycle to obtain a measure of the amount
570 of PCR product formed. The cycle numbers at which each sample reached the threshold were
571 determined and the amounts of *A. fumigatus* DNA in 50 ng of mice lung DNA was evaluated
572 from the standard curve with the cycle numbers and log₂ concentrations of 0.05-50,000 pg of
573 DNA from *A. fumigatus*. All animal studies were approved by the Ethics Review Committee for
574 Animal Experimentation of Nihon University. *A. fumigatus* studies were approved by the
575 Microbial Safety Management Committee of Nihon University School of Pharmacy (E-H25-
576 001).

AAC (Research Article, T.Colley/G.Sehra)

577

578 **Statistical analysis.** Results are expressed as means \pm standard error of the mean (SEM). For
579 comparison between groups either the ordinary one-way ANOVA with Tukey's *post hoc*
580 comparison or the Kruskal-Wallis ANOVA with Dunn's *post hoc* comparison test were
581 performed. Statistical significance was defined as $P < 0.05$.

582

AAC (Research Article, T.Colley/G.Sehra)

583 **ACKNOWLEDGEMENTS**

584 We are grateful to Mr. Marcus Hull and the Engineering and Physical Sciences Research Council
585 National Mass Spectrometry Service Centre at Swansea University for assistance in GC/MS
586 analyses. This work was in part supported by the European Regional Development Fund / Welsh
587 Government funded BEACON research program (Swansea University). We are also grateful to
588 Mr. Takahiro Nakaoki (Nihon University) for assistance in the *in vivo* study.

589

590

AAC (Research Article, T.Colley/G.Sehra)

591 **REFERENCES**

- 592 1. **Denning DW, Pleuvry A, Cole DC.** 2011. Global burden of chronic pulmonary
593 aspergillosis as a sequel to pulmonary tuberculosis. *Bull World Health Organ* **89**:864-
594 872.
- 595 2. **Agbetile J, Fairs A, Desai D, Hargadon B, Bourne M, Mutalithas K, Edwards R,**
596 **Morley JP, Monteiro WR, Kulkarni NS, Green RH, Pavord ID, Bradding P,**
597 **Brightling CE, Wardlaw AJ, Pashley CH.** 2012. Isolation of filamentous fungi from
598 sputum in asthma is associated with reduced post-bronchodilator FEV1. *Clin Exp Allergy*
599 **42**:782-791.
- 600 3. **Bafadhel M, McKenna S, Agbetile J, Fairs A, Desai D, Mistry V, Morley JP,**
601 **Pancholi M, Pavord ID, Wardlaw AJ, Pashley CH, Brightling CE.** 2014. *Aspergillus*
602 *fumigatus* during stable state and exacerbations of COPD. *Eur Respir J* **43**:64-71.
- 603 4. **Woolnough KF, Richardson M, Newby C, Craner M, Bourne M, Monteiro W,**
604 **Siddiqui S, Bradding P, Pashley CH, Wardlaw AJ.** 2017. The relationship between
605 biomarkers of fungal allergy and lung damage in asthma. *Clin Exp Allergy* **47**:48-56.
- 606 5. **Thompson GR, 3rd, Lewis JS, 2nd.** 2010. Pharmacology and clinical use of
607 voriconazole. *Expert Opin Drug Metab Toxicol* **6**:83-94.
- 608 6. **Xiong WH, Brown RL, Reed B, Burke NS, Duvoisin RM, Morgans CW.** 2015.
609 Voriconazole, an Antifungal Triazol That Causes Visual Side Effects, Is an Inhibitor of
610 TRPM1 and TRPM3 Channels. *Invest Ophthalmol Vis Sci* **56**:1367-1373.
- 611 7. **Tucker RM, Haq Y, Denning DW, Stevens DA.** 1990. Adverse events associated with
612 itraconazole in 189 patients on chronic therapy. *J Antimicrob Chemother* **26**:561-566.
- 613 8. **Raad, II, Graybill JR, Bustamante AB, Cornely OA, Gaona-Flores V, Affif C,**
614 **Graham DR, Greenberg RN, Hadley S, Langston A, Negrone R, Perfect JR,**
615 **Pitisuttithum P, Restrepo A, Schiller G, Pedicone L, Ullmann AJ.** 2006. Safety of
616 long-term oral posaconazole use in the treatment of refractory invasive fungal infections.
617 *Clin Infect Dis* **42**:1726-1734.
- 618 9. **Willems L, van der Geest R, de Beule K.** 2001. Itraconazole oral solution and
619 intravenous formulations: a review of pharmacokinetics and pharmacodynamics. *J Clin*
620 *Pharm Ther* **26**:159-169.
- 621 10. **Levin MD, den Hollander JG, van der Holt B, Rijnders BJ, van Vliet M, Sonneveld**
622 **P, van Schaik RH.** 2007. Hepatotoxicity of oral and intravenous voriconazole in relation
623 to cytochrome P450 polymorphisms. *J Antimicrob Chemother* **60**:1104-1107.
- 624 11. **Lat A, Thompson GR, 3rd.** 2011. Update on the optimal use of voriconazole for
625 invasive fungal infections. *Infect Drug Resist* **4**:43-53.
- 626 12. **Wexler D, Courtney R, Richards W, Banfield C, Lim J, Laughlin M.** 2004. Effect of
627 posaconazole on cytochrome P450 enzymes: a randomized, open-label, two-way
628 crossover study. *Eur J Pharm Sci* **21**:645-653.
- 629 13. **Jeong S, Nguyen PD, Desta Z.** 2009. Comprehensive in vitro analysis of voriconazole
630 inhibition of eight cytochrome P450 (CYP) enzymes: major effect on CYPs 2B6, 2C9,
631 2C19, and 3A. *Antimicrob Agents Chemother* **53**:541-551.
- 632 14. **Natesan SK, Chandrasekar PH.** 2016. Isavuconazole for the treatment of invasive
633 aspergillosis and mucormycosis: current evidence, safety, efficacy, and clinical
634 recommendations. *Infect Drug Resist* **9**:291-300.

AAC (Research Article, T.Colley/G.Sehra)

- 635 15. **Yates CM, Garvey EP, Shaver SR, Schotzinger RJ, Hoekstra WJ.** 2017. Design and
636 optimization of highly-selective, broad spectrum fungal CYP51 inhibitors. *Bioorg Med*
637 *Chem Lett* **27**:3243-3248.
- 638 16. **Bruggemann RJ, Donnelly JP, Aarnoutse RE, Warris A, Blijlevens NM, Mouton**
639 **JW, Verweij PE, Burger DM.** 2008. Therapeutic drug monitoring of voriconazole. *Ther*
640 *Drug Monit* **30**:403-411.
- 641 17. **Rodvold KA, Yoo L, George JM.** 2011. Penetration of anti-infective agents into
642 pulmonary epithelial lining fluid: focus on antifungal, antitubercular and miscellaneous
643 anti-infective agents. *Clin Pharmacokinet* **50**:689-704.
- 644 18. **Hoeben BJ, Burgess DS, McConville JT, Najvar LK, Talbert RL, Peters JI,**
645 **Wiederhold NP, Frei BL, Graybill JR, Bocanegra R, Overhoff KA, Sinswat P,**
646 **Johnston KP, Williams RO.** 2006. In Vivo Efficacy of Aerosolized Nanostructured
647 Itraconazole Formulations for Prevention of Invasive Pulmonary Aspergillosis.
648 *Antimicrob Agents Chemother* **50**:1552-1554.
- 649 19. **Hilberg O, Andersen CU, Henning O, Lundby T, Mortensen J, Bendstrup E.** 2012.
650 Remarkably efficient inhaled antifungal monotherapy for invasive pulmonary
651 aspergillosis. *Eur Respir J* **40**:271-273.
- 652 20. **Drew RH, Dodds Ashley E, Benjamin DK, Jr., Duane Davis R, Palmer SM, Perfect**
653 **JR.** 2004. Comparative safety of amphotericin B lipid complex and amphotericin B
654 deoxycholate as aerosolized antifungal prophylaxis in lung-transplant recipients.
655 *Transplantation* **77**:232-237.
- 656 21. **Colley T, Ito K, Strong P, Sunose M, McConville M.** 2016. Antimycotic
657 compound.WO2016185225.
- 658 22. **Denning DW, Venkateswarlu K, Oakley KL, Anderson MJ, Manning NJ, Stevens**
659 **DA, Warnock DW, Kelly SL.** 1997. Itraconazole resistance in *Aspergillus fumigatus*.
660 *Antimicrob Agents Chemother* **41**:1364-1368.
- 661 23. **Atherton G.** 12 January 2014, posting date. *Aspergillus fumigatus* isolate AF293 (NCPF
662 7367). <http://www.aspergillus.org.uk/content/aspergillus-fumigatus-isolate-af293-ncpf-7367>.
- 663 24. **EUCAST.** 2015. EUCAST Antifungal agents breakpoint tables for interpretation of
664 MICs. EUCAST.
- 665 25. **Diaz-Guerra TM, Mellado E, Cuenca-Estrella M, Rodriguez-Tudela JL.** 2003. A
666 Point Mutation in the 14 α -Sterol Demethylase Gene *cyp51A* Contributes to Itraconazole
667 Resistance in *Aspergillus fumigatus*. *Antimicrob Agents Chemother* **47**:1120-1124.
- 668 26. **Institute CaLS.** 2008. M38-A; Reference method for broth dilution antifungal
669 susceptibility testing of filamentous fungi; approved standard-second edition.
- 670 27. **Copeland RA.** 2005. Evaluation of enzyme inhibitors in drug discovery. A guide for
671 medicinal chemists and pharmacologists. *Methods Biochem Anal* **46**:1-265.
- 672 28. **Colley T, Alanio A, Kelly SL, Sehra G, Kizawa Y, Warrilow AG, Parker JE, Kelly**
673 **DE, Kimura G, Anderson-Dring L, Nakaoki T, Sunose M, Onions S, Crepin D,**
674 **Lagasse F, Crittall M, Shannon J, Cooke M, Bretagne S, King-Underwood J,**
675 **Murray J, Ito K, Strong P, Rapeport G.** 2017. In vitro and in vivo antifungal profile of
676 a novel and long acting inhaled azole, PC945, on *Aspergillus fumigatus* infection.
677 *Antimicrob Agents Chemother* doi:10.1128/aac.02280-16.
- 678 29. **Jefcoate CR, Gaylor JL, Calabrese RL.** 1969. Ligand interactions with cytochrome P-
679 450. I. Binding of primary amines. *Biochemistry* **8**:3455-3463.
- 680

AAC (Research Article, T.Colley/G.Sehra)

- 681 30. **Jefcoate CR.** 1978. Measurement of substrate and inhibitor binding to microsomal
682 cytochrome P-450 by optical-difference spectroscopy. *Methods Enzymol* **52**:258-279.
- 683 31. **Kelly SL, Lamb DC, Corran AJ, Baldwin BC, Kelly DE.** 1995. Mode of action and
684 resistance to azole antifungals associated with the formation of 14 alpha-methylergosta-
685 8,24(28)-dien-3 beta,6 alpha-diol. *Biochem Biophys Res Commun* **207**:910-915.
- 686 32. **Baistrocchi SR, Lee MJ, Lehoux M, Ralph B, Snarr BD, Robitaille R, Sheppard DC.**
687 2016. Posaconazole-Loaded Leukocytes as a Novel Treatment Strategy Targeting
688 Invasive Pulmonary Aspergillosis. *J Infect Dis* doi:10.1093/infdis/jiw513.
- 689 33. **Pfaller MA, Sheehan DJ, Rex JH.** 2004. Determination of Fungicidal Activities against
690 Yeasts and Molds: Lessons Learned from Bactericidal Testing and the Need for
691 Standardization. *Clin Microbiol Rev* **17**:268-280.
- 692 34. **Espinel-Ingroff A, Fothergill A, Peter J, Rinaldi MG, Walsh TJ.** 2002. Testing
693 Conditions for Determination of Minimum Fungicidal Concentrations of New and
694 Established Antifungal Agents for *Aspergillus* spp.: NCCLS Collaborative Study. *J Clin*
695 *Microbiol* **40**:3204-3208.
- 696 35. **Meletiadis J, Antachopoulos C, Stergiopoulou T, Pournaras S, Roilides E, Walsh**
697 **TJ.** 2007. Differential Fungicidal Activities of Amphotericin B and Voriconazole against
698 *Aspergillus* Species Determined by Microbroth Methodology ν . *Antimicrob Agents*
699 *Chemother* **51**:3329-3337.
- 700 36. **Southam DS, Dolovich M, O'Byrne PM, Inman MD.** 2002. Distribution of intranasal
701 instillations in mice: effects of volume, time, body position, and anesthesia. *Am J Physiol*
702 *Lung Cell Mol Physiol* **282**:L833-839.
- 703 37. **Miller MA, Stabenow JM, Parvathareddy J, Wodowski AJ, Fabrizio TP, Bina XR,**
704 **Zalduondo L, Bina JE.** 2012. Visualization of murine intranasal dosing efficiency using
705 luminescent *Francisella tularensis*: effect of instillation volume and form of anesthesia.
706 *PLoS One* **7**:e31359.
- 707 38. **Warrilow AGS, Melo N, Martel CM, Parker JE, Nes WD, Kelly SL, Kelly DE.** 2010.
708 Expression, Purification, and Characterization of *Aspergillus fumigatus* Sterol 14- α
709 Demethylase (CYP51) Isoenzymes A and B ν . *Antimicrob Agents Chemother* **54**:4225-
710 4234.
- 711 39. **Lutz JD, Dixit V, Yeung CK, Dickmann LJ, Zelter A, Thatcher JE, Nelson WL,**
712 **Isoherranen N.** 2009. Expression and functional characterization of cytochrome P450
713 26A1, a retinoic acid hydroxylase. *Biochem Pharmacol* **77**:258-268.
- 714 40. **Warrilow AG, Parker JE, Price CL, Nes WD, Kelly SL, Kelly DE.** 2015. In Vitro
715 Biochemical Study of CYP51-Mediated Azole Resistance in *Aspergillus fumigatus*.
716 *Antimicrob Agents Chemother* **59**:7771-7778.
- 717 41. **Parker JE, Warrilow AG, Cools HJ, Fraaije BA, Lucas JA, Rigdova K, Griffiths**
718 **WJ, Kelly DE, Kelly SL.** 2013. Prothioconazole and prothioconazole-desthio activities
719 against *Candida albicans* sterol 14-alpha-demethylase. *Appl Environ Microbiol* **79**:1639-
720 1645.
- 721 42. **Arendrup MC, Cuenca-Estrella M, Lass-Flörl C, Hope W.** 2012. EUCAST technical
722 note on the EUCAST definitive document EDef 7.2: method for the determination of
723 broth dilution minimum inhibitory concentrations of antifungal agents for yeasts EDef
724 7.2 (EUCAST-AFST). *Clin Microbiol Infect* **18**:E246-247.
- 725 43. **Arendrup MC, Cuenca-Estrella M, Lass-Flörl C, Hope W, Howard SJ.** 2014.
726 EUCAST Definitive document EDef 9.2 Method for the determination of broth

AAC (Research Article, T.Colley/G.Sehra)

- 727 microdilution minimum inhibitory concentrations of antifungal agents for conidia
728 forming moulds. EUCAST.
- 729 44. **Institute CaLS**. 2008. M27-A2; Reference method for broth dilution antifungal
730 susceptibility testing of yeasts; approved standard-second edition.
- 731 45. **Zaas AK, Liao G, Chien JW, Weinberg C, Shore D, Giles SS, Marr KA, Usuka J,**
732 **Burch LH, Perera L, Perfect JR, Peltz G, Schwartz DA**. 2008. Plasminogen Alleles
733 Influence Susceptibility to Invasive Aspergillosis. *PLoS Genet* **4**.
- 734 46. **Kimura G, Ueda K, Eto S, Watanabe Y, Masuko T, Kusama T, Barnes PJ, Ito K,**
735 **Kizawa Y**. 2013. Toll-like receptor 3 stimulation causes corticosteroid-refractory airway
736 neutrophilia and hyperresponsiveness in mice. *Chest* **144**:99-105.
737
- 738

AAC (Research Article, T.Colley/G.Sehra)

739 **FIGURE LEGENDS**

740 **FIG 1** Structure of PC1244

741 **FIG 2** Inhibitory activity of PC1244 against 96 clinical isolates of *A. fumigatus* in France and
742 UK. Each horizontal bar was presented as Geometric mean with 95% confidence interval.

743

744 **FIG 3** Efficacy of PC1244 on sterol 14 α -demethylase (CYP51) activity. (A, B) Type II azole
745 binding spectra for *A. fumigatus* CYP51A (A) and CYP51B (B), each experiment was performed
746 4-6 times although only one replicate is shown, (C, D) Azole IC₅₀ determinations of
747 posaconazole (●) and PC1244 (○), mean relative velocity values are shown with standard
748 deviations for *A. fumigatus* CYP51A (C) and CYP51B (D).

749

750 **FIG 4** Duration of action of PC1244 against *A. fumigatus*. (A, B) Persistence of action of
751 PC1244 (A) and voriconazole (B) on *A. fumigatus* hyphae, mean values and SEM of 3
752 independent experiments (each experiment was conducted in quadruplicate), (C, D) Persistence of
753 action of PC1244 (C) and voriconazole (D) on human bronchial cell lines (BEAS2B) infected
754 with *A. fumigatus*, mean values and SEM of 3 independent experiments (each experiment was
755 conducted in triplicate).

756

757 **FIG 5** Colorimetric microbroth assessment of fungicidal activity of PC1244 against *A. fumigatus*
758 [NCPF2010] *in vitro*. Mean values and SEM of 3 independent experiments.

759

760 **FIG 6** Antifungal activity of PC1244 against *A. fumigatus in vivo*. PC1244 (0.0032, 0.0016 and
761 0.08 mg/ml aqueous suspension) and posaconazole (0.08, 0.4 and 2 mg/ml aqueous suspension)

AAC (Research Article, T.Colley/G.Sehra)

762 were intranasally given on days 1, 2 and 3 post infection of *A. fumigatus* in temporary
763 neutropenic immunocompromised mice, Fungal load (CFU/g lung tissue) in lung (A),
764 galactomannan (GM) in BALF (B) galactomannan (GM) in serum (C), CXCL1 in BALF (D),
765 IL-6 in serum (E) and TNF α in serum (F) were evaluated on day 3 post infection (N=5~6).
766 (N=6). Each horizontal bar was presented as mean \pm SD from 5~6 mice per group. * $P<0.05$,
767 ** $P<0.01$, *** $p<0.001$ vs. infected control. “+” dead before sample collection. Serum could not
768 collected from dead mice.

769

770 **FIG 7** Antifungal activity of extended prophylaxis treatment of PC1244 against *A. fumigatus* in
771 *vivo*. Effects of 7 days extended prophylaxis with intranasal PC1244 was compared with that of 1
772 day prophylaxis treatment only on lung fungal load (CFU/g tissue) (A), GM (COI) in serum (B)
773 and malondialdehyde (MDA) in BALF (C) of *A. fumigatus* infected immunocompromised mice
774 (N=4~5). Each horizontal bar was presented as mean \pm SD from 4~6 mice per group. * $P<0.05$.

775

776 **FIG 8** Single prophylactic treatment of PC1244 and posaconazole against *A. fumigatus* in *vivo*.
777 PC1244 at 0.4 mg/ml (14 μ g/mouse) and posaconazole at 0.8 mg/ml (28 μ g/mouse) were
778 intranasally administered 16 h before *A. fumigatus* inoculation, and the lungs were collected for
779 galactomannan (GM, COI) (A) and fungal load (CFU/g tissue) (B) assessment at 8 h after *A.*
780 *fumigatus* inoculation. Each bar was presented as mean \pm SD from 3~4 mice per group. * $P<0.05$.

781

782

783

784

785

AAC (Research Article, T.Colley/G.Sehra)

786

787

788

789

790

791

792

793

794

795

796

797

AAC (Research Article, T.Colley/G.Sehra)

798 **TABLE 1** Antifungal effects of PC1244 and known antifungal agents in azole susceptible and azole resistant strains of *A. fumigatus*^a.

IC₅₀ (IC₉₀) (µg/ml) of indicated agent						
Strain	PC1244	Voriconazole	Posaconazole	Itraconazole	Amphotericin B	Caspofungin
NCPF2010	0.0017 (0.0022)	0.15 (0.21)	0.0070 (0.0084)	0.037 (0.054)	0.20 (0.62)	0.065 (>1)
AF294	0.0021 (0.0041)	0.083 (0.27)	0.0056 (0.011)	0.041 (0.052)	0.21 (0.79)	>1 (>1)
AF293	0.0026 (0.012)	0.25 (0.74)	0.010 (0.028)	0.032 (0.23)	0.24 (0.85)	>1 (>1)
AF72	0.0024 (0.026)	0.025 (0.066)	0.042 (0.30)	0.31 (>1)	0.12 (0.42)	0.065 (>1)
AF91	0.0037 (0.024)	0.14 (0.28)	0.038 (0.049)	0.22 (>1)	0.28 (0.75)	0.11 (>1)

799 ^a IC₅₀ and IC₉₀ values were determined from optical density measurements.

800 All compounds have been tested in a range of concentrations (0.002 ~ 1 µg/ml). The data are from 3 independent experiments and
801 each test was performed in quadruplicate.

802

803

804

805

806

807

808

AAC (Research article, Kimura et al)

809 **TABLE 2** *In vitro* activities of PC1244, posaconazole and voriconazole against 96 clinically
810 isolated *A. fumigatus* strains ^a.

Test Agent	MIC ($\mu\text{g/ml}$) ^b				
	Range	Geometric mean	Mode	MIC ₅₀	MIC ₉₀
PC1244	0.008 – 2	0.067 ^{****}	0.016	0.032	0.50
Voriconazole	0.06 – 4	0.42	0.50	0.50	1.0
Posaconazole	0.016 – 2	0.10 ^{****}	0.032	0.063	0.50

811 ^aAll MIC were determined visually; MIC₅₀ and MIC₉₀ values represent the concentrations
812 required to inhibit 50 and 90% of the strains tested.

813 ^{b****}, P < 0.0001; versus the results for voriconazole (One way ANOVA with Tukey's test).

814

815

816

817

818

819

820

821

822

823

824

825

826

827

828

AAC (Research article, Kimura et al)

829 **TABLE 3** Mean percentage sterol composition of *A. fumigatus* treated with either posaconazole
830 (A) or PC1244 (B) of three biological replicates (\pm standard deviation from the mean).

(A) Sterol	Sterol compositions (posaconazole-treated [$\mu\text{g/ml}$])					
	DMSO	0.0001	0.001	0.01	0.1	1
Ergosterol	100 (\pm 0)	94.5 (\pm 0.1)	87.2 (\pm 0.7)	74.7 (\pm 0.8)	67.8 (\pm 0.3)	67.4 (\pm 1.5)
Ergost-5,7- dienol	0	3.3 (\pm 1.9)	3.9 (\pm 0.6)	0	0	0
Lanosterol	0	0	3.0 (\pm 0.9)	7.0 (\pm 0.3)	8.8 (\pm 0.5)	8.8 (\pm 0.4)
Eburicol	0	2.2 (\pm 1.2)	5.9 (\pm 0.5)	18.3 (\pm 1.1)	23.4 (\pm 0.8)	23.8 (\pm 1.3)

831

(B) Sterol	Sterol compositions (PC1244-treated [$\mu\text{g/ml}$])					
	DMSO	0.0001	0.001	0.01	0.1	1
Ergosterol	100 (\pm 0)	91.3 (\pm 0.1)	89.2 (\pm 0.2)	76.8 (\pm 2.6)	61.0 (\pm 1.6)	58.7 (\pm 2.2)
Ergost-5,7- dienol	0	4.6 (\pm 1.8)	4.6 (\pm 0.8)	0	0	0
Lanosterol	0	1.7 (\pm 0.7)	2.8 (\pm 0.6)	8.5 (\pm 1.1)	12.3 (\pm 0.8)	13.1 (\pm 0.9)
Eburicol	0	2.5 (\pm 1.1)	3.4 (\pm 1.2)	14.7 (\pm 1.5)	26.7 (\pm 0.8)	28.2 (\pm 1.3)

832

833

834

835

836

837

838

839

840

841

842

843

844

AAC (Research article, Kimura et al)

845 **TABLE 4** Potencies and persistence of action of PC1244, posaconazole and voriconazole in *A.*
846 *fumigatus* hyphae and in BEAS2B cells infected with *A. fumigatus*.

Test Agent	Hyphae			BEAS2B		
	IC ₅₀ (µg/ml) ^a			IC ₅₀ (µg/ml)		
	No washout	Washout	Fold change	No washout	Washout	Fold change
PC1244	0.00011 [*]	0.00025	2.41	0.0034 [*]	0.018	5.40
Voriconazole	0.011	>1	>93	0.054	>1	>18.6
Posaconazole	0.00045	0.0022	4.90	0.0031	0.046	14.7

847 ^a*, P < 0.05 for PC1244 versus the results for voriconazole (Kruskal-Wallis one-way ANOVA
848 with Dunn's test).

849 Data are from 3 independent experiments, and each assay was conducted in quadruplicate for
850 hyphae assay and in triplicate for BEAS2B assay.

851

852

853

854

855

856

857

858

859

860

861

862

AAC (Research article, Kimura et al)

863 **TABLE 5** Mean fungicidal activity of PC1244, posaconazole and voriconazole against *A.*
864 *fumigatus* (NCPF2010) of three biological replicates (\pm standard deviation from the mean)..

Test Agent	MIC/MFC ($\mu\text{g/ml}$) ^b [MFC/MIC ratio]		
	MIC	CFU-MFC	XTT-MFC
PC1244	0.063 \pm 0	2 [32]	0.14 [2.2]
Voriconazole	1.67 \pm 0.58	16 [9.6]	>32 [>19]
Posaconazole	0.125 \pm 0	4 [32]	0.42 [3.4]

865 Data are from 3 independent experiments, and each assay was conducted in duplicate.

866

867

868

869

AAC (Research article, Kimura et al)

870 **TABLE 6** Conversion of units of dose give to mice *in vivo* study

mg/ml aqueous suspension	mg/mouse	approx.mg/kg ¹
0.0032	0.00011	0.0056
0.016	0.00056	0.028
0.08	0.0028	0.14
0.4	0.014	0.70
0.8	0.028	1.4
2	0.07	3.5

871 1. 20g is used for calculation as average body weight of used mice

872

873

874

875

876

877

878

879

880

881

882

883

884

885

886

887

AAC (Research article, Kimura et al)

888 **TABLE 7** Antifungal effects of PC1244, voriconazole and posaconazole on other fungal species.

Species (Strain[s])	Strains tested	Culture method	MIC ($\mu\text{g/ml}$) ^a		
			PC1244	Voriconazole	Posaconazole
<i>Aspergillus carbonarius</i> (ATCC8740) ^d	1	CLSI	0.063	0.5	0.063
<i>Aspergillus flavus</i> (ATCC204304) ^d	1	CLSI	0.25	2	0.13
<i>Aspergillus flavus</i> (AFL8; NRRC3357)	2	EUCAST	0.38	0.63	0.16
<i>Aspergillus niger</i> (ATCC1015)	1	EUCAST	0.5	1	0.20
<i>Aspergillus terreus</i> (AT49; AT7130)	2	EUCAST	0.38	1	0.093
<i>Penicillium chrysogenum</i> (ATCC9480) ^d	1	CLSI	0.13	2	0.13
<i>Penicillium citrinum</i> (ATCC9849) ^d	1	CLSI	0.5	>8	0.5
<i>Trichophyton rubrum</i> (ATCC10218) ^d	1	CLSI	0.031	0.063	0.031
<i>Aureobasidium pullulans</i> (ATCC9348) ^d	1	CLSI	1	>8	1
<i>Cladosporium argillaceum</i> (ATCC38013) ^d	1	CLSI	0.25	0.5	0.25
<i>Candida albicans</i> (20240.047; ATCC 10231) ^d	2	CLSI	<0.0078 ^b	0.14 ^b	0.081 ^b
<i>Candida albicans-AR</i> ^c (20183.073; 20186.025) ^d	2	CLSI	(0.25, <0.0078) ^b	10 ^b	8.13 ^b
<i>Candida glabrata</i> (ATCC 36583; R363) ^d	2	CLSI	(<0.0078, 0.13) ^b	8.13 ^b	0.5 ^b
<i>Candida krusei</i> (ATCC6258) ^d	1	CLSI	0.13	0.25	0.125
<i>Candida parapsilosis</i> (ATCC22019) ^d	1	CLSI	0.25 ^b	NT	0.25 ^b
<i>Chaetomium globosum</i> (ATCC44699) ^d	1	CLSI	0.063	1	0.25
<i>Gibberella zeae</i> (<i>Fusarium graminearum</i>) (ATCC16106) ^d	1	CLSI	1	>8	>8
<i>Cryptococcus gattii</i> (Clinical isolate)	1	EUCAST	0.5	0.125	0.5

42

AAC (Research article, Kimura et al)

<i>Cryptococcus neoformans</i> (ATCC24067) ^d	1	CLSI	0.016	0.016	0.016
<i>Lichtheimia corymbifera</i> (ATCC 7909) ^d	1	CLSI	1	>8	>8
<i>Mucor circinelloides</i> (ATCC8542) ^d	1	CLSI	2	>8	>8
<i>Rhizomucor pusillus</i> (ATCC16458) ^d	1	CLSI	2	>8	>8
<i>Rhizopus oryzae</i> (ATCC11145) ^d	1	CLSI	0.25	>8	>8

889 ^a Due to the limited number of strains tested, the mean of isolate MICs is presented.

890 ^b MIC indicates 50% inhibition of fungal growth as azole readout.

891 ^cAR, azole resistant (fluconazole and voriconazole).

892 ^d All details of isolate and assay protocol are described in <https://www.eurofinspanlabs.com>

893 (Anti-infective assay/Fungi)

894 NT = not tested.

895

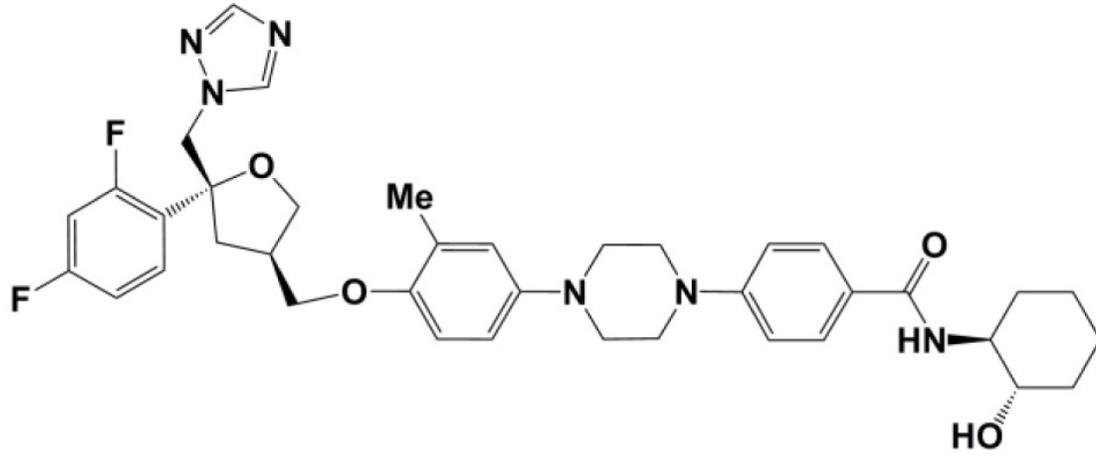
FIGURE 1

FIGURE 2

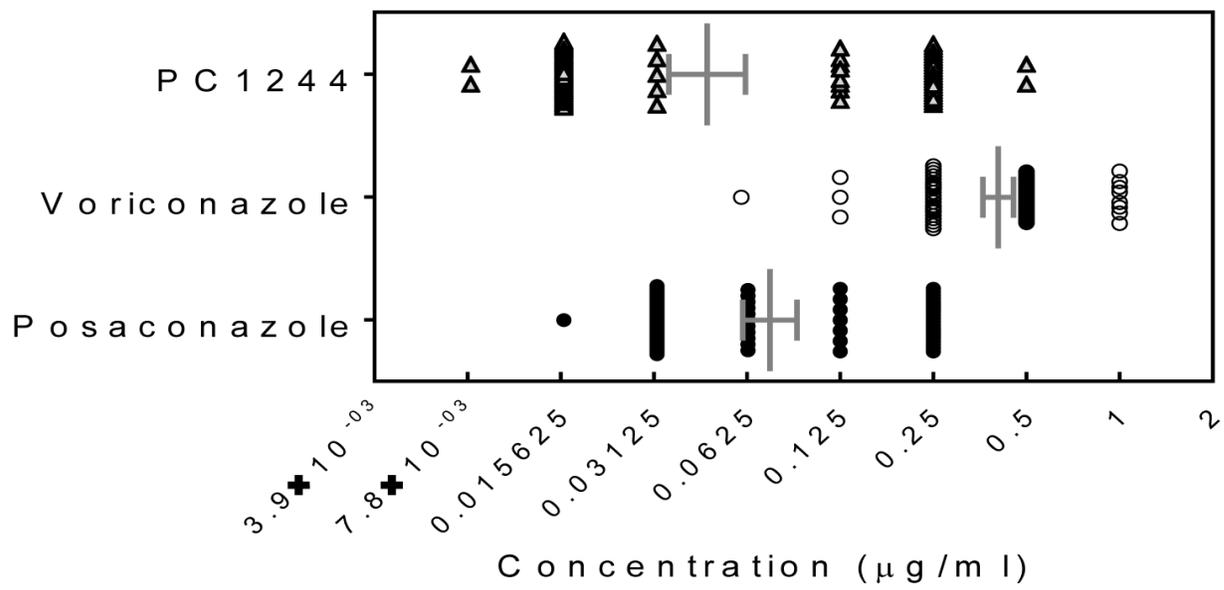


FIGURE 3

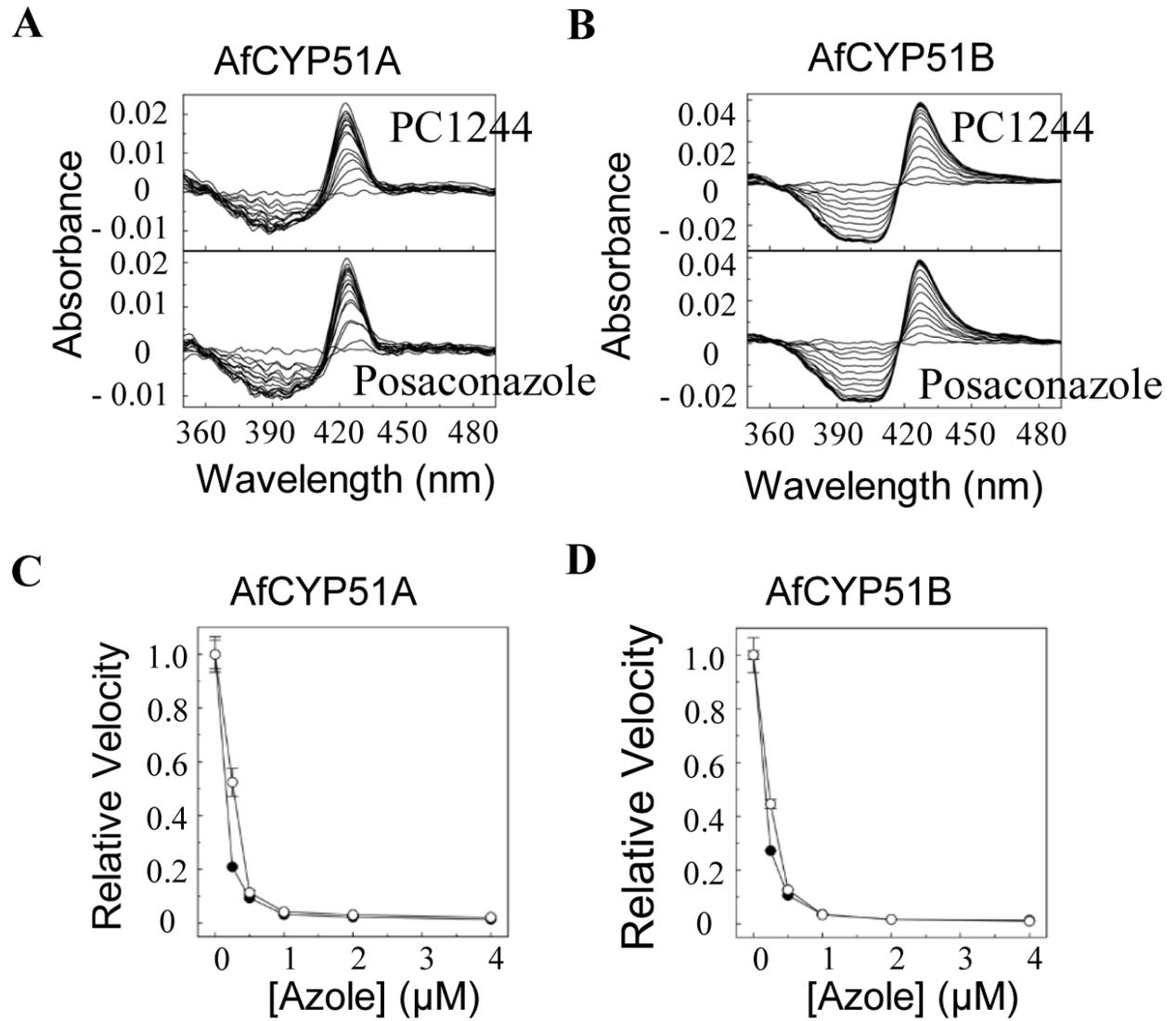


FIGURE 4

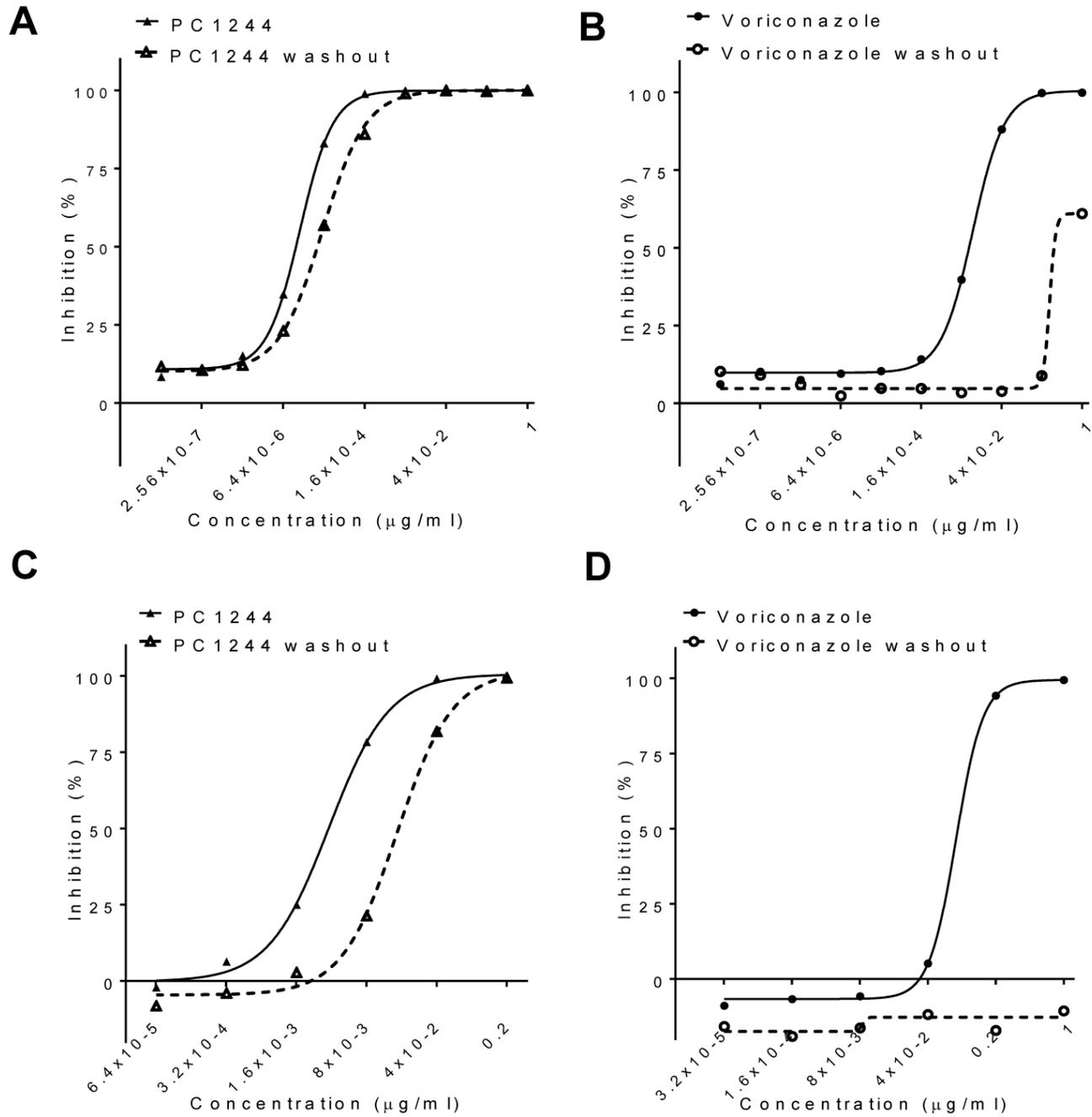


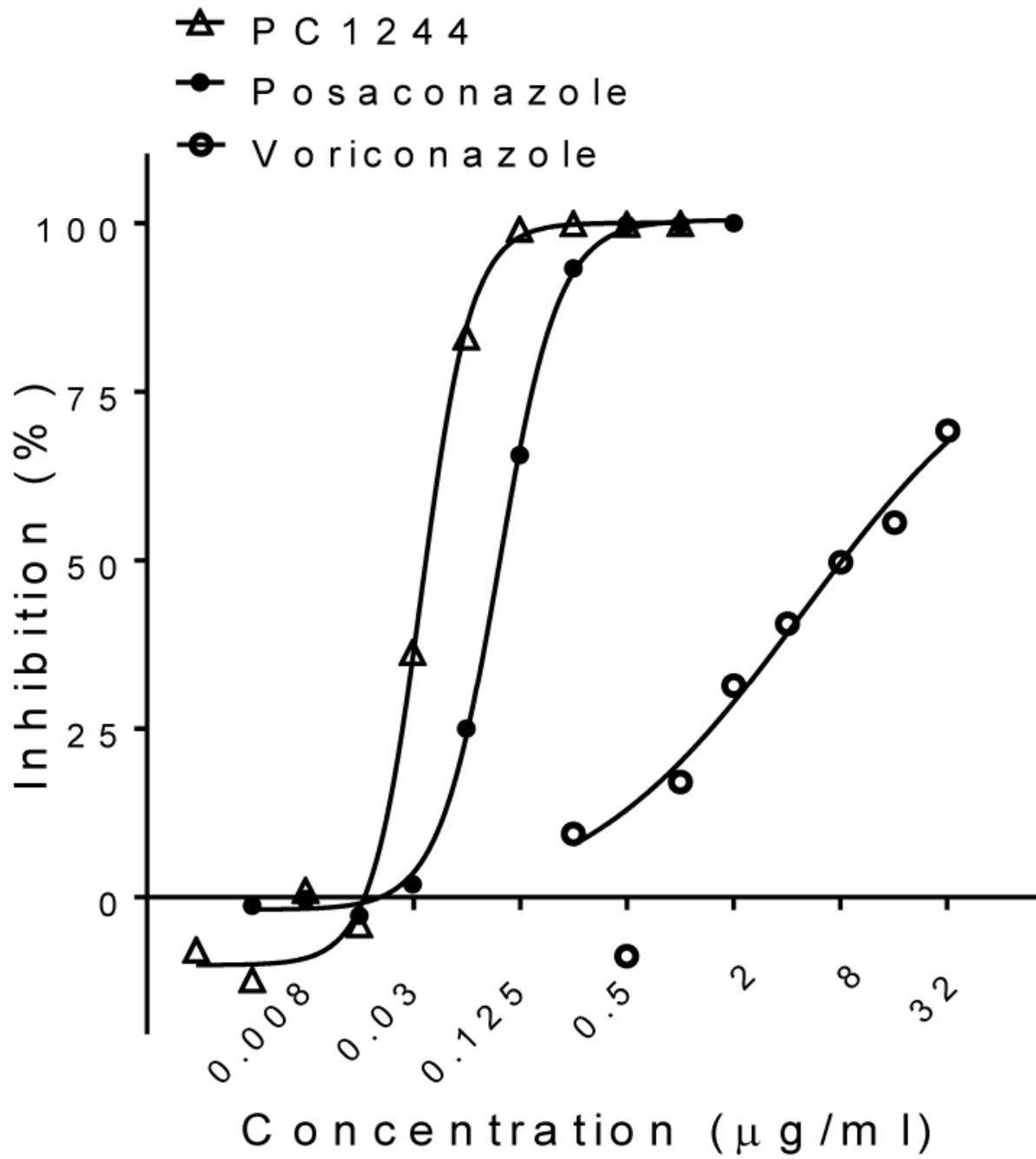
FIGURE 5

FIGURE 6

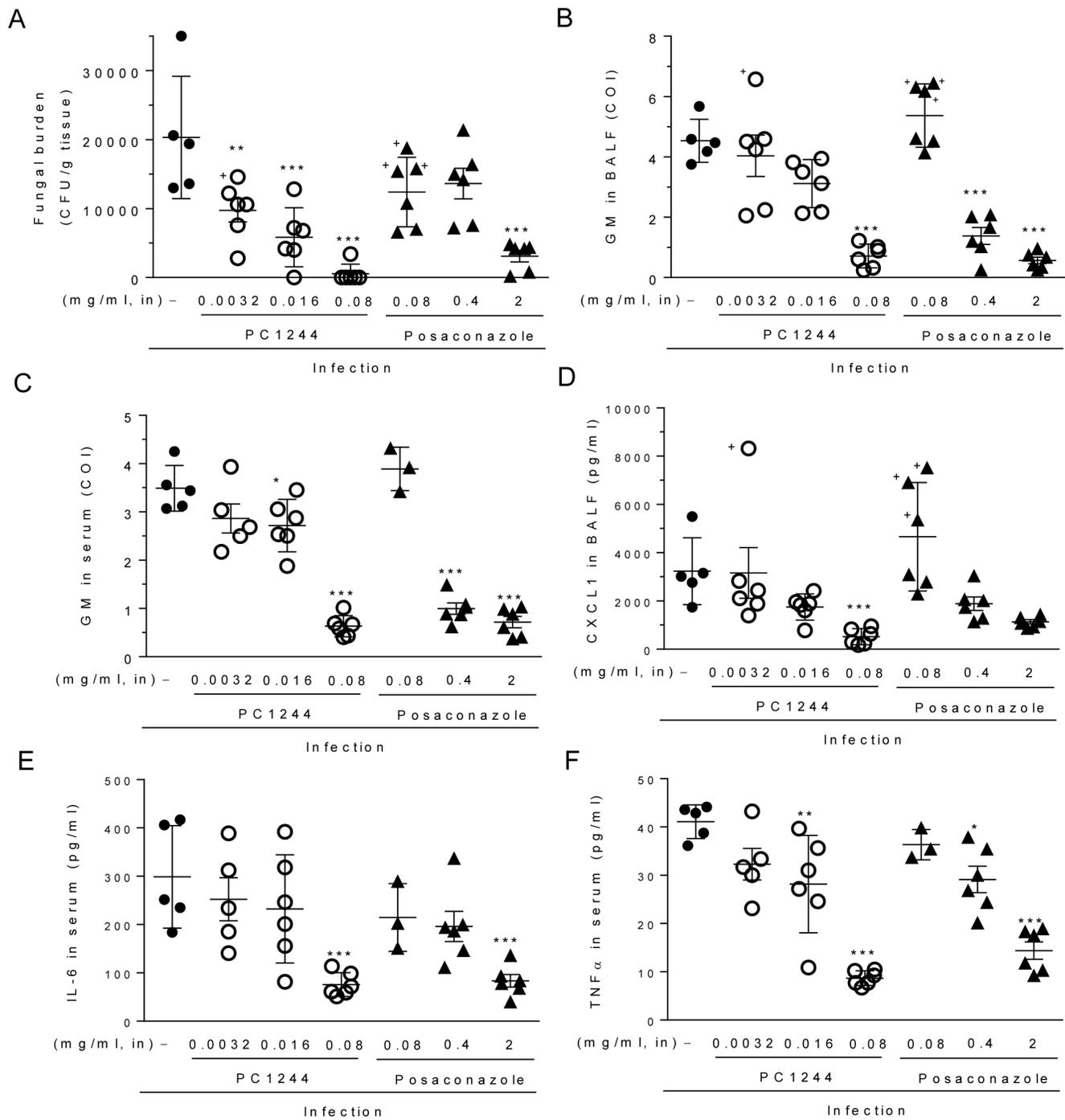


FIGURE 8

