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BMJ Open  Proinsulin in the identification and risk stratification of gestational diabetes mellitus: study protocol for a prospective, longitudinal cohort study

Rajesh Peter, Dominic Bright, Wai-Yee Cheung, Stephen D Luzio, Gareth J Dunseath

ABSTRACT

Introduction Gestational diabetes mellitus (GDM) is a common metabolic disorder occurring in up to 10% of pregnancies in the western world. Most women with GDM are asymptomatic; therefore, it is important to screen, diagnose and manage the condition as it is associated with an increased risk of maternal and perinatal complications. Diagnosis of GDM is made in the late second trimester or early third trimester because accurate diagnosis or risk stratification in the first trimester is still lacking. An increase in serum proinsulin may be seen earlier in pregnancy and before a change in glycaemic control can be identified. This study will aim to establish if fasting proinsulin concentrations at 16–18 weeks gestation will help to identify or stratify high-risk pregnant women with GDM.

Methods and analysis This is a prospective, longitudinal cohort study. Two oral glucose tolerance tests will be carried out at 16–18 and 24–28 weeks gestation in 200 pregnant women with at least one risk factor for GDM (body mass index >30 kg/m², previous macrosomic baby (>4.5 kg), previous gestational diabetes, first degree relative with type 2 diabetes mellitus) recruited from antenatal clinics. Blood samples will be taken fasting and at 30 min, 1 and 2 hours following the 75 g glucose load. In addition, a fasting blood sample will be taken 6-weeks post delivery. All samples will be analysed for glucose, insulin, C peptide and proinsulin. Recruitment began in November 2017. Optimal cut-off points for proinsulin to diagnose gestational diabetes according to National Institute for Health and Care Excellence (2015) criteria will be established by the receiver operating characteristic plot and sensitivity and specificity will be calculated to assess the diagnostic accuracy of proinsulin at 16–18 weeks gestation.

Ethics and dissemination This study received ethical approval from the Wales Research Ethics Committee (Panel 6) (Ref. 17/WA/0194). Data will be presented at international conferences and published in peer-reviewed journals.

Trial registration number ISRCTN16416602; Pre-results.

INTRODUCTION

Gestational diabetes mellitus (GDM) is a common metabolic disorder occurring in up to 10% of pregnancies in the Western world.¹ GDM is defined as any degree of carbohydrate intolerance with onset or first recognition during pregnancy. Most women with GDM are asymptomatic; hence, it is important to screen, diagnose and manage the condition as it is associated with an increased risk of maternal and perinatal complications such as pre-eclampsia, macrosomia, shoulder dystocia and neonatal hypoglycaemia. Diagnosis of GDM is made in the late second trimester or early third trimester because accurate diagnosis or risk stratification in the first trimester is still lacking, and in the UK women with a high risk of GDM are currently offered an oral glucose tolerance test (OGTT) at 24–28 weeks gestation.²

The risk factors that predispose to the development of GDM include body mass index (BMI) >30 kg/m², previous macrosomic baby
(>4.5 kg), previous gestational diabetes, family history of type 2 diabetes mellitus (T2DM) (first degree relative with diabetes) and certain ethnic groups. Current diagnostic criteria are a fasting plasma glucose concentration of ≥7.8 mmol/L or a 2 hour value of ≥11.1 mmol/L. Glycosylated haemoglobin (HbA1c) cannot be used to diagnose GDM as HbA1c is insufficiently sensitive to substitute for OGTT as a screening test.

Women who develop GDM have a high risk of future-compromised glycaemic control. A review of published studies indicated a 7.43-fold increase risk of postpartum diabetes in women with GDM compared with women with healthy glycaemic control during pregnancy and the incidence of postpartum diabetes in North America and Europe has indicated a prevalence rate of between 30% and 50% up to 15 years’ follow-up.

Normal pregnancy is accompanied by a progressive increase in insulin resistance that begins midway through pregnancy and progresses through the second and third trimesters with resultant increase in insulin secretion to compensate for the acquired resistance. The levels of insulin resistance are not too dissimilar to that seen in individuals with T2DM. GDM then develops when the insulin supply is no longer adequate to maintain normal blood glucose regulation.

Proinsulin is a precursor molecule for insulin and is synthesised by the pancreatic beta cells. Proinsulin is an 86 amino acid peptide, incorporating the A and B chains of insulin in addition to C peptide between amino acid residues 31 and 65. Under normal circumstances, virtually all proinsulin is cleaved at residues 32–33 and 65–66 to produce C peptide and insulin, although a small amount of intact proinsulin may also be released into the circulation along with des 31–32 split proinsulin and 32–33 split proinsulin. In the presence of insulin resistance, pancreatic beta-cell function is affected with disproportionately more proinsulin (both intact and split) being secreted compared with insulin as seen in subjects with T2DM.

Previous studies investigating serum proinsulin measurements in pregnancy have studied pregnant women irrespective of individual risk which may account for varied findings. However, studies comparing the proinsulin concentrations of healthy pregnant women and those who have gestational diabetes have found that proinsulin concentrations at fasting are significantly elevated in GDM compared with control subjects. These studies however did not assess the use of proinsulin as a biomarker for GDM; rather they compare women already diagnosed with GDM with proinsulin measured only at 24–28 weeks. Therefore it is possible that an increase in serum proinsulin may be seen earlier in pregnancy and before a change in glycaemic control can be identified. Specifically targeting pregnant women that are high risk may make serum proinsulin measurements more sensitive and specific to identify those that will develop GDM at an earlier stage in their pregnancy and subsequently suitable for earlier intervention.

AIMS AND OBJECTIVES

This study will aim to establish if fasting proinsulin concentrations at 16–18 weeks gestation will help to identify or risk stratify high-risk pregnant women with GDM diagnosed according to National Institute for Health and Care Excellence (2015) criteria. It will also seek to establish if 30 min and/or 1 and 2 hour post oral glucose load proinsulin measurements at 16–18 weeks gestation can discriminate women with gestational diabetes and predict which women will need insulin to control hyperglycaemia. The relationship of the various risk factors to plasma proinsulin levels will also be evaluated.

The primary objective of the study is to test the hypothesis that fasting intact proinsulin measurements at 16–18 weeks gestation will discriminate or risk stratify gestational diabetes (diagnosed from an OGTT at 24–28 weeks) from women with normal glucose tolerance.

The secondary objectives are:

- To test the hypothesis that 30 min, 1 and 2 hour post 75 g oral glucose load, proinsulin measurements at 16–18 weeks gestation can predict those women subsequently diagnosed with gestational diabetes at 24–28 weeks.
- To test the hypothesis that fasting, 30 min and/or 1 and 2 hour post oral glucose load, proinsulin measurements at 16–18 weeks gestation can predict those women with gestational diabetes that will need insulin during pregnancy.
- To study the relationship of various risk factors to plasma proinsulin concentrations and gestational diabetes in the second and third trimesters of pregnancy.

METHODS AND ANALYSIS

Study design

This is a prospective, longitudinal cohort study (table 1). Study recruitment started on 14 November 2017 and the study is expected to last until December 2019.

Two OGTTs will be carried out at 16–18 and 24–28 weeks gestation in 200 pregnant women with at least one risk factor for GDM (BMI >30 kg/m², previous macroscopic baby (>4.5 kg), previous gestational diabetes, first degree relative with T2DM) recruited from antenatal clinics within Abertawe Bro Morgannwg University Health Board, Wales. Women of ethnic origin considered to be high risk will need to have another mentioned risk factor to be eligible for inclusion in the study. Blood samples will be taken fasting and at 30 min, 1 and 2 hours following the 75 g glucose load. In addition, a fasting blood sample will be taken at 6-weeks post delivery. All samples will be analysed for glucose, insulin, C-peptide and proinsulin. Both intact proinsulin and total proinsulin (the sum of...

intact proinsulin, 32–33 split proinsulin and des 31–32 split proinsulin) will be assayed.

**Setting and site selection**
Recruitment of pregnant women will be via a study poster placed at antenatal clinics within Abertawe Bro Morgannwg University Health Board, Wales. Study procedures will be carried out at a single site (Joint Clinical Research Facility, Swansea University).

**Informed consent**
Informed consent for each subject will be obtained prior to initiating any trial procedures. Potential participants eligible to take part in the study will receive an invitation letter from their hospital consultant or midwife, along with a participant information leaflet and be given an oral explanation about the study from a research professional (usually a research nurse). Written informed consent is given by the participant by signing and dating a consent form which will be countersigned and dated by either a study nurse or principal investigator to confirm that the participant has the opportunity to ask questions and fully understands the nature of the study. Thereafter, a copy of the consent form will be given to each of the participants. Research professionals can facilitate the consent process for the study if authorised to do so on the site delegation log following appropriate training including good clinical practice (GCP). The consent process makes clear that the participant can withdraw from the trial whenever they wish without giving a reason and without affecting their future care in any way. The reasons for withdrawal will be documented, if known, and site staff will be encouraged to trace participants lost to follow-up and document the reasons for their loss whenever possible. Each participant will receive an identification number to ensure confidentiality and any samples will be identified by only using only the identification number. Data will be recorded in a case record form.

**Patient population**
Pregnant women at 16–18 weeks gestation with at least one of the following risk factors for GDM will be studied. Those recruited will meet the following inclusion/exclusion criteria:

<table>
<thead>
<tr>
<th>Inclusion criteria</th>
</tr>
</thead>
<tbody>
<tr>
<td>► BMI &gt;30 kg/m².</td>
</tr>
<tr>
<td>► Previous macrosomic baby (&gt;4.5 kg).</td>
</tr>
<tr>
<td>► Previous GDM.</td>
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<tr>
<td>► Family history of T2DM (first degree relative with diabetes).</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Exclusion criteria</th>
</tr>
</thead>
<tbody>
<tr>
<td>► Subjects unable or unwilling to sign informed consent.</td>
</tr>
<tr>
<td>► Known previous diabetes mellitus or on treatment with metformin.</td>
</tr>
<tr>
<td>► Known chronic infection like hepatitis or HIV or chronic kidney, liver or heart disease.</td>
</tr>
<tr>
<td>► Previous bariatric surgery.</td>
</tr>
</tbody>
</table>

**Table 1**

<table>
<thead>
<tr>
<th>Timepoint</th>
<th>Enrolment</th>
<th>Visit 1</th>
<th>Visit 2</th>
<th>Visit 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Enrolment</td>
<td></td>
<td>16–18 weeks gestation</td>
<td>24–28 weeks gestation</td>
<td>6 weeks post partum</td>
</tr>
<tr>
<td>Eligibility screen</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>Informed consent</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>Intervention</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OGTT (0, 30, 60 120 min)</td>
<td>x*</td>
<td>x</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fasting blood sample</td>
<td></td>
<td></td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>Assessments</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OGTT glucose</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>OGTT proinsulin</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>OGTT C peptide</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>OGTT insulin</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>Fasting blood for glucose, proinsulin, C peptide, insulin</td>
<td></td>
<td></td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>Birth weight of baby</td>
<td></td>
<td></td>
<td>x†</td>
<td></td>
</tr>
<tr>
<td>Apgar score at birth</td>
<td></td>
<td></td>
<td>x†</td>
<td></td>
</tr>
<tr>
<td>Medication</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
</tbody>
</table>

*Participants exceeding the National Institute for Health and Care Excellence (2015) gestational diabetes mellitus diagnostic thresholds at this visit will be withdrawn from the study and immediately referred back to their antenatal team for follow-up.
†Obtained from medical notes.
OGTT, oral glucose tolerance test.
Women who will be diagnosed with gestational diabetes at the first visit (16–18 weeks) will be withdrawn from the study and referred back to routine antenatal care.

**Study visits**
Participants will be seen on three different occasions.

The first two visits will be at 16–18 weeks and 24–28 weeks gestation. Participants will be asked to attend having fasted for 10 hours. On arrival at the clinical unit, a fasting blood sample will be taken. Patients will then be given a drink containing 75g glucose. At 30 min, 1 and 2 hours following the drink further blood samples will be taken. The 30 min sample has been included to allow capture of the peak plasma insulin response and for robust estimation of insulin sensitivity.

The third visit will be 6-week post delivery where participants will be asked again to attend having fasted for 10 hours. On this occasion, a fasting blood sample only will be taken. Additional data including birth weight of the baby, Apgar score at birth and medications including insulin doses just prior to birth will be collected.

**Laboratory measurements**
Laboratory measurements for glucose, insulin, C peptide and proinsulin (total and intact) will be carried out in the Good Clinical and Laboratory Practice Accredited Diabetes Research Unit Cymru Laboratory based at Swansea University.

Glucose samples will be taken into fluoride oxalate tubes and will be measured using a glucose oxidase assay (YSI 2300 Stat Plus, Fleet, Hampshire, UK). Insulin, C peptide, total and intact proinsulin samples will be taken into EDTA tubes and measured using specific immunoassays using chemiluminescent labels (Invitron, Mmmouth, UK).

**Safety evaluations and data monitoring**
The data monitoring committee (DMC) will monitor the overall conduct of the trial, safeguarding the interests of the trial participants and assessing the safety and efficacy of the intervention. Patients identified as glucose intolerant at 16 weeks will be considered to have had pre-existing glucose intolerance and will be referred back to their antenatal team and excluded from further participation in the study. All serious adverse events (AEs) that occur during the study will be recorded and reported in accordance with local requirements and will be reported to the DMC. All AEs will be recorded on a case report form and reviewed as part of central data monitoring.

**Statistical analysis plan**
To address the primary objective and the first secondary objective, glucose measurements following an OGTT at 24–28 weeks will be used as the reference standard to classify GDM status.

Optimal cut-off points for proinsulin will be established by the receiver operating characteristic plot which plotted the proportion of true positives against the proportion of false positives. 95% CIs for the area under curve, sensitivity and specificity will be calculated to assess the diagnostic accuracy of total and intact proinsulin at 16–18 weeks gestation.

The same approach will be used to address the second secondary objective, but with the use of insulin during pregnancy as the reference standard.

Logistic regression will be used to analyse the relationship of proinsulin concentrations at 16–18 weeks gestation and other individual risk factors for the development of GDM.

All the analysis and data preparation will be done using statistical analyses which will be performed by a statistician using SPSS V.22 which is validated statistical software for clinical trial studies.

Values will be checked for normality, applying suitable transformations as necessary. All statistical hypothesis tests will be performed at a 5% significance level. All available data from withdrawn subjects will be included in the analysis up to the time of withdrawal where possible.

**Sample size**
Sample size was estimated with Buderer’s formula\(^1^5\) which used prevalence, level of clinically acceptable precision and a hypothesised level of sensitivity and specificity as the parameters for estimating sample size.

There were no direct estimates of GDM prevalence in the high-risk population. Prevalence of GDM in the Western world was estimated to be \(\pm 10\%\)\(^6\) and the relative risk of developing GDM in women with one of the risk factors (BMI \(>30\) kg/m\(^2\)) was 2.74.\(^1^6\) There are also reports of women with a family history of diabetes having a \(>6\)-fold risk of developing gestational diabetes.\(^1^7\) Therefore, we have estimated the prevalence of GDM in our target population to be \(\pm 30\%\). We took the conservative approach of basing our sample size estimation on hypothesised sensitivity rather than specificity. Since the estimated prevalence was lower than 50%, this approach would give a higher estimate of the number to be included in the study.

As a meaningful screening test, a sensitivity of 90% for the circulating concentration of total and intact proinsulin at 16–18 weeks gestation for GDM is expected as established by glucose measurements following an OGTT at 24–28 weeks. The minimal clinically acceptable precision for estimate of sensitivity is 6%. We also expect loss to follow-up to be less than 10%.

Adjusted for possible loss to follow-up, a sample size of 200 will be required to estimate with precision of \(\pm 0.06\) for an expected sensitivity of 0.9 with the proinsulin test, at the significance level of 0.05, if the prevalence of GDM is not lower than 30%.

**Patient and public involvement**
In preparation for this study, the Public Reference Group of the Diabetes Research Unit Cymru was consulted and their opinions sought on the concept of the study and the study design.
ETHICS AND DISSEMINATION

Research governance
The study conforms with the Research Governance Frameworks for England and Wales, and the principles of GCP outlined by the International Conference on Harmonisation (http://www.ich.org/). The participating National Health Service (NHS) Health Board has given NHS permission and will be responsible for auditing the study.

Publication
In accordance with good practice, we have registered the GDM study in a public registry at International Standard Randomised Control Trial Number (ISRCTN 16416602). This is a diagnostic accuracy study and findings will be reported according to the Standards for Reporting of Diagnostic Accuracy Studies (STARD) guideline.18 We shall present study findings at national and international conferences and publish as widely as possible in open access, peer-reviewed journals. Any published results will be made available to study participants from their study nurse on request (this is made clear in the patient information sheet) and any results will also be made available on the Diabetes Research Unit Cymru website (www.diabeteswales.org.uk) with a direct link to any publication and with a summary in lay language.

DISCUSSION

A recent systematic review and meta-analysis19 have shown that there is no good evidence for any of the diagnostic criteria for early onset GDM and have suggested the use of a fasting glucose of 6.1–6.9 mmol/L in the first trimester. Others20 have tried to develop a prediction model for obese women at high risk of GDM to facilitate targeted interventions.

Current practice is that pregnant women with at least one risk factor for GDM have an OGTT at 24–28 weeks and a fasting glucose measurement 6 weeks post delivery. It is not unusual for some patients having an OGTT at 28 weeks to have glucose levels above 10 mmol/L indicating that they have been hyperglycaemic for some time and therefore it would be useful if we could reasonably identify at an earlier stage those women who will develop GDM and also those that would require insulin to control hyperglycaemia. This would help manage scarce resources, as those that are unlikely to require insulin could be followed up by experienced allied healthcare professionals like specialist midwives while those that require insulin could be followed up by diabetologists and/or diabetes specialist nurses earlier in their pregnancy. It will also be useful to revisit the pathophysiology of GDM with respect to the onset of insulin resistance and corresponding insulin response at the various stages of pregnancy. Moreover, proinsulin assays have become more specific; thus, proinsulin can now be reliably measured by well-characterised immunoassays. This paper summarises the current approved protocol.

REFERENCES


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