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1   **Predictable gene expression related to behavioral**  
2   **variation in parenting**

3

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15   Running title: “Predicting genes related to variable behavior”

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20 Differential gene expression has been associated with transitions between  
21 behavioral states for a wide variety of organisms and behaviors. Heterochrony,  
22 genetic toolkits, and predictable pathways underlying behavioral transitions have  
23 been hypothesized to explain the relationship between transcription and  
24 behavioral changes. Less studied is how variation in transcription is related to  
25 variation within a behavior, and if the genes that are associated with this variation  
26 are predictable. Here we adopt an evolutionary systems biology perspective to  
27 address two hypotheses relating differential expression to changes within and  
28 between behavior. We predicted fewer genes will be associated with variation  
29 within a behavior than with transitions between states, and the genes underlying  
30 variation within a behavior will represent a narrower set of biological functions.  
31 We tested for associations with parenting variation within a state with a set of  
32 genes known *a priori* to be differentially expressed between parenting states in  
33 the burying beetle *Nicrophorus vespilloides*. As predicted, we found that far fewer  
34 genes are differentially expressed related to variation within parenting. Moreover,  
35 these were not randomly distributed among categories or pathways in the gene  
36 set we tested and primarily involved genes associated with neurotransmission.  
37 We suggest that this means candidate genes will be easier to identify for  
38 associations within a behavior, as descriptions of behavioral state may include  
39 more than a single phenotype.

40

41 **Key words:** **behavior genetics, evolutionary systems biology, gene**  
42 **expression, *Nicrophorus vespilloides*, parental care**

43 **INTRODUCTION**

44 Ever since the description of the “phenotypic gambit” (Grafen 1984), evolutionary  
45 biologists in general (Travisano and Shaw 2013) and behavioral ecologists specifically  
46 (Stamps 1991; Zuk and Balenger 2014) have struggled with the questions of if and when  
47 it is necessary to understand the mechanisms underlying behavior. The development of  
48 transcriptomic, metabolomic, and proteomic approaches have provided us with new tools  
49 to address mechanisms (Moore et al. 2010), and advocates of a genomic approach to  
50 behavior have argued that finding the genes underlying behavior can lead us to important  
51 insights into the nature of pleiotropy and constraint, selection, and evolutionary  
52 convergence (Fitzpatrick et al. 2005; Rittschof and Robinson 2014; Bengston et al. 2018).  
53 Furthermore, understanding details of behavioral mechanisms may provide insight into  
54 whether behavior plays a unique role in evolution (Bailey et al. 2018). However, what  
55 precisely does it mean for a gene to underlie a behavior? Most studies have asked what  
56 genes are responsible for producing discrete changes in behavior and have focused on  
57 gene expression changes between two behavioral states (Harris and Hofmann 2014;  
58 Rittschof and Robinson 2016), particularly on a genome wide scale (Calisi and  
59 MacManes 2015). These studies have led to predictions of the types of genes that will be  
60 differentially expressed (DE). Two hypotheses have been generally supported to explain  
61 change in behavior. First, changes in timing of gene expression (heterochrony) will be  
62 associated with behavioral evolution (Linksvayer and Wade 2005). Second, the genes DE  
63 will be associated with a “genetic toolkit” (Rittschof and Robinson 2016; Toth and Rehan  
64 2017). The toolkit hypothesis states that specific genes may differ between organisms,  
65 but the underlying pathway or category altered will be shared across animals displaying

66 similar behavior. These hypotheses are applied to the generation of novel behavioral  
67 states and are therefore well-suited to answer questions about the evolutionary origins of  
68 behavior and the tradeoffs involved with its production.

69 Behavior also displays abundant variation within a state, which also must have  
70 some mechanistic basis. The genes related to this type of behavioral variation should be  
71 more important for understanding responses to contemporary selection; if selection acts  
72 to refine behaviors, it should similarly refine levels of transcription associated with  
73 specific behavioral values. Less attention has been paid to mechanisms responsible for  
74 this type of variation. For instance, the extent that differential gene expression within and  
75 between behavioral states overlaps is unknown (Bengston et al. 2018). Many genes  
76 undergo massive expression changes when behavior shifts, indicating that behavioral  
77 transitions also involve concurrent suites of major physiological changes. We do not  
78 know to what degree the same genes influence variation within state, possibly with less  
79 extreme differences in expression. Some have argued that the mechanisms controlling  
80 the maintenance of specific behavioral phenotypes are likely to be different than those  
81 controlling transitions between behaviors (Cardoso et al. 2015). Alternatively, others  
82 have predicted more overlap between these mechanisms (Duckworth 2015), perhaps  
83 reflecting the expectation that behavioral plasticity and behavioral evolution are  
84 mechanistically similar (Pfennig et al. 2010). However, existing data explicitly  
85 comparing transitions with variation for a single behavior are sparse. One example comes  
86 from Bell et al. (2016), who found notable but limited overlap between mechanisms of  
87 induction of aggression and variation in aggression in sticklebacks. They proceed to

88 suggest that the magnitude of such overlap may be less important than the identity of the  
89 common genes.

90 Theory from developmental biology may bear on our expectations for the  
91 distinction between transitions between behavior states and variation within a behavior  
92 and provide an addition to the toolkit and heterochrony hypotheses. Wagner's (2014)  
93 makes a distinction between the origin of novel types and modification of existing types.  
94 Wagner predicts that the former evolves via sweeping changes in transcriptional  
95 programs leading to novel regulatory networks, whereas the latter evolves within existing  
96 networks through the modification of expression of “realizer” genes, which are directly  
97 related to phenotypic variation. Adapting Wagner's hypothesis to behavior, we make two  
98 predictions. First, we propose that only a narrow subset of genes DE in changes between  
99 states will be associated with variation within that state. Changing behavioral states  
100 involves complex and multifaceted environmental changes, and therefore the need for  
101 changes in multiple coordinated sensory, physiological, and neurological processes, while  
102 variation within a state occurs in a single social and more uniform biotic and abiotic  
103 environment where only a single phenotype may be different among individuals.  
104 Therefore, fewer transcriptional changes should be needed. Second, we suggest that this  
105 subset of genes influencing variation within a state will fall into functional categories  
106 typically displaying causal links to behavior; i.e., neuropeptides (Chandrasekaran et al.  
107 2011). Presumably, if the behavior in question is heritable, *cis*-regulatory variants in  
108 such causal genes will allow them to vary transcriptionally without generating broad  
109 rewiring of transcriptional networks as occurs during transitions (Chandrasekaran et al.  
110 2011).

111 Parental care in the burying beetle *Nicrophorus vespilloides* presents an  
112 opportunity with which to test these predictions. Adult beetles rear their offspring on  
113 prepared vertebrate carcasses (Eggert and Müller 1997; Scott 1998), and the transition  
114 from a non-parenting to a parenting state is phenotypically complex and multifaceted.  
115 Parents undergo changes in immune function (Cotter and Kilner 2010; Palmer et al. 2016;  
116 Ziadie et al. 2019), physiology (Benowitz et al. 2017a), chemical status (Steiger et al.  
117 2008; 2009) and multiple behaviors (reviewed in Scott 1998; Royle and Hopwood 2017).  
118 Reflecting this, an RNA-seq study comparing non-parenting and parenting individuals  
119 identified broad transcriptional differences (> 700 genes) between the two states (Parker  
120 et al. 2015). Transcriptional differences have also been defined for responses to social  
121 context and for plasticity in male parenting (Cunningham et al. 2019). Thus, we have a  
122 clear phenotype with well characterized DE associations.

123 Within the burying beetle parenting state, the primary individual social behavior  
124 is provisioning, which consists of direct regurgitation of partially digested food from  
125 parents into the mouths of begging offspring (Pukowski 1933; Milne and Milne 1976).  
126 The amount of parental provisioning exhibited in *N. vespilloides* is highly variable  
127 (Benowitz et al. 2016a), heritable (Walling et al. 2008) and important for offspring  
128 development and fitness (Eggert et al. 1998; Lock et al. 2004). Variation in the extent of  
129 provisioning has been investigated in an RNA-seq study comparing genome wide  
130 transcriptional variation between ten high-caring and ten low-caring parents (Benowitz et  
131 al. 2017b). This study broadly supported the expectation of overlap between genes  
132 involved in transitions and within-state variation but could not specify the number or the  
133 identity of the genes affecting the variability of parental provisioning (Benowitz et al.

134 2017b). This likely reflects a basic limitation of using RNA-seq to investigate subtle  
135 behavioral differences; if small phenotypic changes are accompanied by small expression  
136 changes, important signals may be swamped by noise and costly multiple corrections.

137 Here, to further probe our hypotheses in *N. vespilloides*, we take a complementary  
138 approach to Benowitz et al. (2017b) but with an *a priori* candidate gene method rather  
139 than RNA-seq, creating a more focused and therefore more powerful test. We examine  
140 specific candidates shown to be significantly DE when switching from a non-parenting to  
141 a parenting state in earlier studies (Table 1, S1), and ask which of these genes also  
142 display differential expression between very high and very low caring mothers.  
143 Furthermore, we specifically chose to examine genes spanning a range of well-  
144 documented functions, including several neurotransmitters, in order to ask whether the  
145 genes associated with behavioral variation are more likely to display certain  
146 functionalities. There are certainly more genes than these involved; however, most DE  
147 genes identified in transcriptomic studies are unannotated and therefore the function is  
148 unknown (Parker et al. 2015).

149

## 150 **METHODS**

151 *Nicrophorus vespilloides* were collected and maintained as an outbred colony as  
152 described by Cunningham et al. (2014). For this study, our sample consisted of 57 female  
153 adult beetles on 19-21g mouse carcasses in a uniparental context. We observed parental  
154 provisioning, defined as mouthpart-to-mouthpart contact by parents and offspring, on the  
155 first day after larvae hatched. Following Benowitz et al. (2016) we made 80 scan samples  
156 for provisioning behavior over the course of an 8-hour period. Within the entire dataset,

157 parental care was roughly uniformly distributed (Benowitz et al. 2016a) with a mean and  
158 standard deviation of  $47.53 \pm 21.52$  observations of feeding (out of 80 scans) (Benowitz  
159 et al. 2016b). Immediately after observations we removed the heads, which were flash-  
160 frozen in liquid nitrogen and stored at -80°C.

161 Following the method of Benowitz et al. (2017b) we selected the 12 highest and  
162 12 lowest caring female parents for RNA extraction. Mean and standard deviation of  
163 parenting was  $11.67 \pm 8.33$  observations of feeding for the low group and  $72.42 \pm 2.18$   
164 observations of feeding for the high group, indicating substantial quantitative behavioral  
165 differences (Benowitz et al. 2016b). We analyzed head tissue, which contains both brain  
166 and fat body, following previous studies that identified differential expression of genes  
167 associated with parenting in females (Parker et al. 2015; Cunningham et al. 2014, 2016,  
168 2019; Roy-Zokan et al. 2015; Benowitz et al. 2017a,b). We performed phenol-  
169 chloroform extractions using Qiagen RNeasy Lipid Kits (Qiagen, Venlo, Netherlands)  
170 and synthesized cDNA using qScript (Quantabio, Beverly, MA) reverse transcriptase  
171 (Parker et al. 2015; Roy-Zokan et al. 2015). We designed quantitative real-time PCR  
172 (qRT-PCR) primers for 23 genes (Table S1) identified in other independent experiments  
173 to be DE in transition from non-parenting to parenting *N. vespilloides* females. Seventeen  
174 of these genes were identified in an RNA-seq experiment comparing parenting and non-  
175 parenting *N. vespilloides*, while the other six were identified in qRT-PCR experiments  
176 making the identical comparison (Cunningham et al. 2016; 2017; unpub. data). These  
177 genes have functional annotation and can be categorized into neurotransmission, energy  
178 acquisition and usage, immunity, and hormones (Table 1). This is not an exhaustive  
179 identification of any genes differing between parental states in *N. vespilloides*, but rather

180 those that are annotated, DE when comparing parenting and non-parenting states, and fall  
181 into different known functional categories. Therefore, this set is coarsely representative  
182 of the types of genes DE between states, and most importantly allows us to test our  
183 hypotheses. We performed qRT-PCR using each primer pair on each of the 24 samples  
184 with a Roche LightCycler 480 (Roche, Basel, Switzerland) with *alpha-tubulin* (Table S1)  
185 used as an endogenous reference gene. We calculated gene expression for each sample  
186 as  $2^{-(Cp_{exp} - Cp_{ref})}$ , where  $Cp_{exp}$  is the average cycle number of three technical  
187 replicates of each experimental gene and  $Cp_{ref}$  is the average cycle number of three  
188 technical replicates for *alpha-tubulin*. Following this, DE was calculated between high  
189 and low care using separate one-way ANOVAs for each experimental gene. Because  
190 each gene represents a distinct, *a priori* specified hypothesis, conservative multiple  
191 corrections are inappropriate (Rice 1989). We used a Fisher's Exact Test to test whether  
192 genes *a priori* classified as having roles in neurotransmission were more likely to be DE  
193 than genes with other functional roles. The data are publicly available in Dryad  
194 (Benowitz et al. 2018).

195

## 196 RESULTS

197 Four of seven neurotransmission genes (*serotonin receptor 2, octopamine/tyramine*  
198 *receptor 2, tachykinin*, and *glutamate receptor*) show statistically different expression  
199 between high vs. low caring *N. vespilloides* mothers (Table 1). Furthermore, the  
200 direction of differential expression followed *a priori* expectations based on our previous  
201 studies; genes that were upregulated during the transition into parental care were also  
202 upregulated in high caring mothers, and vice versa. However, of the other genes we

203 examined, which included hormones and genes related to energy, immunity, and general  
204 behavior, only one (*fatty acyl-CoA synthetase*) of 16 was DE between high and low  
205 caring mothers (Table 1), and in the opposite direction as the change with a transition to  
206 parenting. Neurotransmission genes were statistically more likely to be associated with  
207 provisioning variation than other genes (p = 0.017).

208

## 209 **DISCUSSION**

210 Differences in gene expression are suggested to be fundamental contributors to the  
211 evolution of social behavior (Calisi and McManes 2015; Rittschof and Robinson 2016;  
212 Toth and Rehan 2017; Kronauer and Libbrecht 2018). Despite this, we currently lack a  
213 nuanced understanding of how transcription is related to selectable variation in social  
214 behavior. Here, we examined how gene expression relates to quantitative variation in  
215 parental provisioning behavior involving direct regurgitation of food in the burying beetle  
216 *Nicrophorus vespilloides*. Rather than examining transcriptional differences associated  
217 with transitions between behavioral states (e.g., Harris and Hoffmann 2014; Rittschof and  
218 Robinson 2016), we determined which of the genes previously associated with this  
219 transition in *N. vespilloides* parenting were also related to variation in parenting within a  
220 state. Transitions often involve large changes in physiology, feeding, aggression and  
221 reproduction (Kronauer and Libbrecht 2018), and so many different pathways and genes  
222 should be involved. We specifically tested our predictions that far fewer genes will be  
223 involved in variation within a behavioral state, and for behavior they will be involved in  
224 neurotransmission rather than other physiological processes. Focusing on those genes  
225 that we know are associated with changes in parenting state provides a powerful test of

226 the hypothesis. That is, we bias our examination toward genes known to change, rather  
227 than looking for associations of genes and behavior, thereby reducing extraneous  
228 correlations and irrelevant tests. What we cannot determine from this approach is the  
229 contribution to variation of genes that do not see transcriptional changes across  
230 behavioral transitions. Behavioral syndromes theory (Sih et al. 2004) predicts the  
231 existence of large suites of genetically correlated behaviors, suggesting that common  
232 regulators may control behavior in a non-specific fashion, and therefore might not be  
233 involved in behavioral transitions. If such genes exist and are important, they may be  
234 difficult to identify by either *a priori* or genome-wide approaches.

235 Four of seven neurotransmitter genes associated with changes in parenting state  
236 were also associated with variation in parenting, while only one of 16 not associated with  
237 neurotransmission also influenced parental variation. This result suggests that many  
238 immune, energetic, and hormonal processes altered upon induction of parental behavior  
239 are not further changed in association with behavioral variation. We suggest this pattern  
240 is generalizable, and that this methodology eliminates many of the confounding  
241 transcriptional effects produced when comparing between different behaviors. Here, we  
242 predicted fewer transcriptional differences within parenting because the number of  
243 changes required to transition from a non-parenting to parenting state include those  
244 associated with changing biotic and abiotic conditions as well as changes in parenting.  
245 This highlights a further problem studying behavior: terminology. The labels widely used  
246 to group behaviors into categories (e.g. parenting) actually describe composites that are  
247 necessarily more complex than the individual phenotypes that are contained within them  
248 (Rittschof and Robinson 2016). However, it is these individual phenotypes that are

249 actually quantified, and often considered to be proxies for the entire category, which may  
250 lead to an oversimplification (Wenzel 1992).

251 The neurotransmission genes related to provisioning variation included serotonin,  
252 octopamine, glutamate, and tachykinin signaling genes. These pathways are all known to  
253 influence behavior, and expression of an octopamine/tyramine receptor has previously  
254 been shown to affect behavior quantitatively in *C. elegans* (Bendesky et al. 2011).

255 Furthermore, modification of neural signaling pathways leading to variable brain function  
256 is predicted to be a general mechanism for the evolution of social behavior (Baran et al.  
257 2017). Interestingly, the single other gene found to be differentially expressed was the  
258 lipid metabolism gene *fatty acyl CoA synthetase*, which was upregulated in high caring  
259 parents. This raises the possibility that body condition could be linked to variation in  
260 parenting, presenting precisely the sort of confound our study was designed to avoid.  
261 Because plastic changes of this nature likely require additional gene expression changes  
262 in order to incorporate environmental inputs into behavior, it will be interesting to see in  
263 the future whether plastic variation is more transcriptionally complex than genetic  
264 variation. We hypothesize that this will be the case, and therefore that more heritable  
265 behaviors will be associated with fewer DE genes.

266 There have been several attempts to provide a framework for predicting specific  
267 genes or pathways underlying behavior given the accessibility of modern molecular  
268 approaches, from heterochrony (Linksvayer and Wade 2005) to genetic toolkits (Toth and  
269 Rehan 2017), to hypotheses derived from ethological principles (Cunningham et al. 2016,  
270 2017; Kronauer and Libbrecht 2018). We suggest that molecules predicted from an  
271 evolutionary systems biology approach (Chandrasekaran et al. 2011; Wagner 2014)

272 coupled with predictions from ethology, allowing targeted transcriptional comparison  
273 between individuals with quantitatively different behavioral phenotypes, presents a  
274 promising methodology for finding genes that may be related to behavioral variation.  
275 The identities of these molecules, and their regulation in other behavioral contexts, will  
276 help inform important questions on the constraints and possibilities of behavioral  
277 evolution.

278

279

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286 presented here.

287

## 288 **AUTHORS' CONTRIBUTIONS**

289 K.M.B., E.C.M., and A.J.M. conceived the study and analyzed the data, E.C.M. and  
290 C.B.C. collected the data, and K.M.B. and A.J.M wrote the paper. All authors approved  
291 the version to be published and agree to be accountable for all aspects of the research.

292

## 293 **DATA ACCESSIBILITY**

294 Analyses reported in this article can be reproduced using the data provided by Benowitz  
295 et al. (2018).

296

297 **COMPETING INTERESTS.**

298 We declare no competing interests.

299

300 **ETHICAL STATEMENT**

301 All institutional, national, and international guidelines for the care and use of insects for  
302 research were followed.

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**Table 1.** Classification of genes and their patterns of differential expression between high and low caring mothers.

Gene	Functional Role	Evidence for DE in transition between parenting states	Expression in high (mean ± sd)	Expression in low (mean ± sd)	Comp of exp in high/low car
Thaumatin	Immune	Parker et al. 2015	1.177 ± 1.051	1.143 ± 1.787	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Peptidoglycan recognition protein SC2-like	Immune	Parker et al. 2015	3.447 ± 2.743	2.926 ± 3.519	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Peptidoglycan recognition protein A	Immune	Parker et al. 2015	2.578 ± 1.136	2.742 ± 0.995	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Defensin	Immune	Parker et al. 2015	9.564 ± 4.923	9.219 ± 5.237	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Prophenoloxidase	Immune	Parker et al. 2015	4.293 ± 2.098	3.942 ± 1.585	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Serine protease 93	Immune	Parker et al. 2015	0.885 ± 0.663	0.660 ± 0.638	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Beta glucosidase	Digestion/Immune	Parker et al. 2015	4.294 ± 2.822	3.881 ± 5.626	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Fatty acyl-CoA synthetase	Lipid synthesis	Parker et al. 2015	0.00927 ± 0.00401	0.00495 ± 0.00273	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Vitellogenin 1	Lipid transport/Hormone	Parker et al. 2015; Roy-Zokan et al. 2015	135.800 ± 85.387	113.003 ± 38.542	(F <sub>1,22</sub> = 0.0001, p = 0.999)
Hexamerin 3	Energy storage/Hormone Transport	Parker et al. 2015	0.0201 ± 0.00863	0.0253 ± 0.014	(F <sub>1,22</sub> = 0.0001, p = 0.999)

Hexamerin 4	Energy storage/Hormone Transport	Parker et al. 2015	$0.0677 \pm 0.104$	$0.0238 \pm 0.0240$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Apolipophorin-III	Lipid transport	Parker et al. 2015; Benowitz et al. 2017b	$135.430 \pm 55.438$	$117.294 \pm 25.408$	(F <sub>1,22</sub> = 0.3, p = 0.6)
Yellow 3	Hormone/Pigment/Immune	Parker et al. 2015	$0.802 \pm 0.277$	$0.628 \pm 0.302$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Insulin like peptide 3	Hormone	Parker et al. 2015	$0.0598 \pm 0.0136$	$0.0589 \pm 0.0175$	(F <sub>1,22</sub> = 0.8, p = 0.3)
Neuropeptide F	Neurotransmission	Cunningham et al. 2016	$0.0735 \pm 0.0212$	$0.0916 \pm 0.0601$	(F <sub>1,22</sub> = 0.3, p = 0.6)
Serotonin receptor 7	Neurotransmission	Unpub. data	$0.400 \pm 0.108$	$0.454 \pm 0.0941$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Serotonin receptor 2	Neurotransmission	Unpub. data	$0.0233 \pm 0.00336$	$0.0286 \pm 0.00742$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Glutamate receptor	Neurotransmission	Parker et al. 2015	$0.179 \pm 0.0261$	$0.0257 \pm 0.0679$	(F <sub>1,22</sub> = 13.97, p = 0.001)
Natalisin	Neurotransmission	Cunningham et al. 2017	$0.00964 \pm 0.00509$	$0.0208 \pm 0.0259$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Octopamine/Tyramine receptor 2	Neurotransmission	Unpub. data	$0.0250 \pm 0.00637$	$0.0326 \pm 0.00931$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Tachykinin	Neurotransmission	Unpub. data	$0.431 \pm 0.183$	$0.292 \pm 0.136$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Takeout	Circadian behavior/Feeding	Parker et al. 2015	$0.0499 \pm 0.0321$	$0.0435 \pm 0.0338$	(F <sub>1,22</sub> = 0.1, p = 0.9)
Pickpocket	Sodium channels/Olfaction	Parker et al. 2015	$0.0834 \pm 0.0655$	$0.0559 \pm 0.0176$	(F <sub>1,22</sub> = 0.1, p = 0.9)

**Table S1.** Primers and accession numbers of the genes used in this study.