1	The response of steroid estrogens bioavailability to various						
2	sorption mechanisms by soil organic matter extracted with						
3	sequential alkaline-extraction method from an agriculture soil						
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ABSTRACT: The long-term groundwater contamination risks posed by steroidal 16 estrogens (SEs) in animal-manured agricultural soils are closely associated with the soil 17 organic matter (SOM) content and composition. In this study, the bioavailability of 18 estrone (E1) and 17β -estradiol (17β -E2) under different sorption mechanism in humic 19 acids (HA1 and HA2) and humin (HM) extracted with sequential alkaline-extraction 20 technique (SAET) were examined. These SOMs extracted by SAET showed various 21 properties and sorption characteristics for SEs. The alkyl carbon and condensed SOM 22 23 increased during SAET, but aromatic carbon decreased and the same trend for polarity. Quick sorption was the major SEs sorption mechanism on HA1 and HA2, which 24 contributed more than 69%; whilst slow sorption rate was about 50% in soil and HM. 25 The $\log K_{oc}$ values were proportional to the TOC of SOM according to Freundlich fitting, 26 and the sorption capacity of sorbent for E1 and 17β -E2 was related to the log K_{ow} values, 27 indicating that the main mechanism controlling the SEs sorption was hydrophobic 28 interaction. The larger micropore volume of HM and soil was more conducive to the 29 micropore filling of SEs. Meanwhile, the specific sorption of SEs on condensed domain 30 31 of SOM was the main reason for the strong desorption hysteresis and slow sorption in HM and soil. The SEs degradation rate was positively correlated with the contribution 32 rate of quick adsorption and negatively correlated with the contribution rate of slow 33 adsorption, indicating that the bioavailability of SEs sorbed by hydrophobic interaction 34 was higher than that of micropore filling or specific sorption, which was also the reason 35 for the low bioavailability of SEs in HM and soil. This work confirms the regulation of 36 on-site SOM compositions and their properties on SEs sorption and bioavailability. 37 Characterization of these details is crucial for the improved prediction of long-term 38 39 risks to groundwater.

- Keywords: Steroid Estrogens, Sorption, Bioavailability, Soil Organic Matter, Risk
- 41

42 1. Introduction

Steroidal estrogens (SEs) such as E1 (estrone) and 17β-E2 (estradiol) are 43 commonly found in agricultural soils and groundwater after fertilization with livestock 44 and poultry manure (Gall et al., 2015; Fang et al., 2016; Zhao et al., 2019). Their high 45 bioactivity may cause endocrine disruption effects on the agroecological system, even 46 following exposure to low SEs concentrations (Adeel et al., 2017; Zhao et al., 2019; 47 Wei et al., 2020). Although gains have been made through organic green agriculture in 48 reducing chemical fertilizer application risks, these can be offset by greater 49 environmental risks from SEs posed by increased manure application. As a result of the 50 incomplete sorption and biodegradation in the vadose zone (Song et al., 2018; Zhao et 51 al., 2019), underlying groundwater resources are at risk of SEs contamination through 52 leaching from soils (Citulski et al., 2010; Yang et al., 2021). Improved understanding 53 of sorption and bioavailability controls on the accumulation and decline of legacy SEs 54 sources in agricultural soils and their regulation of groundwater risks is crucial. 55

The sorption mechanism of SEs in soil is complex due to the heterogeneity of soil 56 components. Especially the origin, type, and the distribution and structure of the 57 functional groups of soil organic matter (SOM), including humic acid (HA) and humin 58 (HM), play a central role in the sorption of SEs, which have a significant impact on the 59 bioavailability of SEs (Sangster et al; 2015; Alizadeh et al., 2018; Yu et al., 2020; Sun 60 et al., 2010). A few studies have confirmed that SEs sorption affinity has a positive 61 correlation with SOM content and the octanol-water partition coefficient (K_{ow}) of SEs, 62 dominance of 63 supporting the hydrophobic interaction mechanisms (Karnjanapiboonwong et al., 2010; Mashtare et al., 2011). However, further sorption 64 mechanisms must exist because of the presence of polar functional groups both in SEs 65 and humic substances (Sun et al., 2012; Gall et al., 2016). For instance, the dominant 66 role of aromatic carbon versus aliphatic carbon in SOM controlling the sorption of SEs 67 remains unresolved (Sun et al., 2008; Jiang et al., 2017). The sorption of SEs may be 68 significantly influenced by their polar hydroxyl function on the benzene ring of SEs 69 molecules or their π - π binding (Takigami et al., 2011; Bedard et al., 2014; Zhang et al., 70

2018; Zhao et al., 2020). The feature of SEs sorption on HA or HM is often nonlinear, 71 which is attributed for the specific polar interaction (H-bonding) or pore-filling (Lima 72 et al., 2012; Zhang et al., 2017). Soil minerals and its heterogeneity also affect sorption 73 (Tong et al., 2019; Xu et al., 2020). The sorption capacity of SEs on iron oxides and 74 clay minerals is governed by the content of their ion-exchange materials 75 (Thanhmingliana et al., 2016; Shi et al., 2017), whereas the sorption on 76 montmorillonites displayed interlayer spacing decrease, thereby supporting 77 78 incorporation of SEs into the clay interlayer space and producing slow sorption (Sun et al., 2017). Therefore, further investigation of the sorption behavior of SEs in a real 79 agricultural soil (in preference to artificial, commercial products) is needed to evaluate 80 SEs bioavailability and transport in the agricultural system. To our knowledge, related 81 82 research remains surprisingly limited.

The bioavailability of SEs in agricultural soil that are closely related to 83 biodegradation is another important factor affecting their environmental risk because 84 of their nonpersistent properties (Stumpe et al., 2010; Zheng et al., 2012). The 85 86 bioavailability of contaminants refers to the accessibility of microorganisms or extracellular enzymes in soil and groundwater, which is controlled by the concentration 87 of directly available contaminants and the rate of conversion from potentially available 88 contaminants (Wang et al., 2021). As a result of the heterogeneity of soil components 89 90 and the complexity of the hydrophobic organic contaminant (HOCs) sorption mechanism on SOM, there may be considerable differences in related bioavailability, 91 which leads to the different opportunities for the utilization of sorbed and free HOCs 92 by microorganisms (Zhang et al., 2018; Gámiz et al., 2018). It is generally believed that 93 94 only free SEs can be effectively degraded by soil microorganisms, while sorbed SEs need to be desorbed from the solid phase before utilization (Fan et al., 2007). In 95 particular, soil with high SOM content may reduce the bioavailability and 96 biodegradation rates due to irreversible sorption (Lee et al., 2011). However, 97 Combalbert et al. (2010) suggested that soils with different sorption capacities have 98 99 little influence on the biodegradation of SEs, and the sorbed SEs could be freely desorbed to the liquid phase and degraded. In addition, the degree of isolation of HOCs 100

increases significantly with the time spent in contact with the soil, a process known as
aging, which may increase desorption resistance and reduce the bioavailability of HOCs
(Xu et al., 2018). The response of SEs bioavailability to sorption mechanism and to
sorption aging with different SOM remains unclear, and is thus the key scientific
problem to be solved in the current study.

106 Thus, we hypothesized that the differences in composition and structure of SOM fractions can result in different controls on the SEs sorption, thereby different 107 bioavailability of sorbed SEs. The main objectives of the present research were to (1) 108 investigate controls on the sorption and desorption of E1 and 17β -E2 in a real 109 agricultural soil and in its component SOM fractions, (2) characterize the conformation 110 and functional groups of SOMs to gain insights into the mechanism of nonlinear and 111 slow sorption of SEs, and (3) explore the effect of sorption behavior on SEs 112 bioavailability in soil and clarify the possible mechanisms related to differences in 113 SOM properties and aging processes. The findings of this work could provide 114 theoretical support for guiding the sustainable development of agriculture and 115 116 controlling the risk of SEs contamination of groundwater from agricultural sources.

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2. Materials and methods

118 2.1. SEs solutions, agricultural soil, and soil bacterial suspension

The selected sorbates, E1 and 17β-E2, are commonly detected in farmyard manure and agricultural soil (Adeel et al., 2017), and their physicochemical properties are listed in Table S1. E1 (\geq 99.5%) and 17β-E2 (\geq 98.4%) were purchased from Sigma-Aldrich (USA). The stock solutions of individual SEs were prepared by dissolving each compound in methanol at a concentration of 1000 mg/L before being stored at -20 °C prior to use.

An agricultural silt loam (9.71% clay, 51.91% silt, and 38.38% sand) was collected from a rural farm in Shenyang, Northeastern China, where maize, vegetables, and fruits are predominantly grown. In our previous investigation (Song et al., 2018), the total concentrations of both SEs in the farm soil were 39.3–112.7 ng/g, compared with

manure at 85.6–357.3 ng/g. For this study, the soil samples without SEs used for SAET 129 of SOM and batch experiments were collected from 0-10 cm soil depth around the 130 edges of the farm where manure had not been applied; however, the soil microbial 131 suspension was prepared using the SEs-containing soil for bioavailability experiment. 132 The fresh soil was air-dried, passed through a 2-mm sieve, and stored at 4 °C before the 133 sorption and bioavailability experiments. The soil samples were treated by sterilization 134 and non-sterilization. For the sterile treatment, soils were autoclaved at 120 °C for 30 135 min three successive times. 136

The soil microbial suspension was prepared as follows: 10.0 g non-sterile soil and 20 mL sterile water were mixed well in a 200 mL amber reagent bottle and sealed, followed by incubating at 30 °C in the dark for 3 days. After 80 mL of sterile water was added, the slurry was stirred at 30 °C, 180 rpm for 1 h. Finally, the slurry was centrifuged at 1000 rpm for 10 min to collect the supernatant as a soil microbial suspension.

143 2.2. Sequential alkaline-extraction technique (SAET)

The SAET method, a modification of the standard method of the International 144 Humic Substances Society, was conducted for the isolation of HA and HM fractions 145 from the agricultural soil (Swift et al., 1996); in brief, (1) soil equilibration was set to 146 pH 1–2 and adjusted with 0.1 M HCl to a ratio of 10 mL liquid to 1 g dry sample, then 147 the suspension was shaken for 1 h at 70 °C; (2) residue was separated by centrifugation 148 and neutralized to pH 7 under a N_2 atmosphere; (3) the alkaline suspension was shaken 149 at 70 °C for 24 h and each supernatant was collected by centrifugation. The above steps 150 151 were repeated until the supernatant became light yellow or colorless, allowing the HM to be obtained as a precipitate. The supernatant was acidified with 6 M HCl to pH 1-152 1.5 and left to stand for 12 h, followed by centrifugation to separate the HA (precipitate). 153 The HA was suspended in 0.1 M HCl/0.3 M HF solution and shaken overnight. The 154 precipitate was collected by centrifugation and the HA was transferred to a dialysis tube 155 until the dialysis water gave a negative Cl⁻ test with AgNO₃, and then the HA was 156 freeze-dried. Next, 46% HF/2M HCl (1:1) was added to the HM, and the suspension 157

was shaken at 70-80 °C for 2-3 min. The precipitate was centrifuged for collection, 158 and the HM residue was washed with deionized water and then freeze-dried to obtain 159 the HM. The SAET for HA was conducted five times in this study. As the amount of 160 HA decreased with the sequential extractions, HAs obtained during the first and second 161 extraction were combined to give the sorbent HA1, and the HA fractions obtained 162 during the third to fifth extractions were combined to give the sorbent HA2. This was 163 required to fulfil the requirements for sorbent dosage in the experiments investigating 164 SEs sorption to different HA fractions. 165

166 *2.3. SEs detection and sorbent characterization*

The solid phase extraction (SPE) and gas chromatography-mass spectrometry 167 168 (GC-MS) approach previously developed by our team (Song et al., 2018) was applied for quantification of the E1 and 17β-E2 concentrations, and details on SEs detection 169 were provided in the Supplementary Material (S1). The C, H, O, N, and S contents of 170 the sorbent were analyzed using an Elementar Vario ELIII elemental analyzer 171 (Germany). The solid-state cross-polarization magic angle-spinning ¹³C NMR spectra 172 of all sorbents were obtained with a Bruker Avance III 600 MHz NMR Spectrometer 173 (Germany) operated at a ¹³C frequency of 100 MHz. Surface elemental composition 174 and carbon-based functionalities of the sorbent were determined using XPS (Thermo 175 ESCALAB 250Xi, USA) with a monochromatic Al KR radiation source operated at 176 225 W, 15 mA, and 15kV. Spectrum processing was accomplished with the Xpspeak41 177 software to identify the functional group corresponding to the binding energy (Nguyen 178 et al., 2009; Doskočil et al., 2015; Mylotte et al., 2015). FTIR (Perkin-Elmer 1725X) 179 180 was used to analyze the functional groups in soil and its organic fractions, with a resolution of 4 cm⁻¹ and measurement range of 4000–400 cme⁻¹. The surface area, 181 micropore area, and micropore volume were used to examine the level of pore filling 182 and calculated from the adsorption-desorption isotherm of N2 at 77 K by the multipoint 183 Brunauer-Emment-Teller (BET) method using a surface area and porosimetry analyzer 184 (Micromeritics ASAP 2460, USA) (Xin et al., 2012). 185

187 *2.4. Batch sorption experiments*

The batch sorption experiments of E1 and 17β-E2 were conducted on soil 188 component fractions in screw-cap vials with aluminum foil and Teflon liners. 189 Background solutions consisted of 0.01 M CaCl₂ in deionized water to maintain a 190 constant ionic strength. The optimal liquid/solid phase ratio (25:1 for soil and 40:1 for 191 HA/HM) and initial SEs concentration range were determined through preliminary 192 experiments to obtain 20%-80% uptake of sorbate at equilibrium. For kinetic 193 experiments, the vials were placed in a thermostatic oscillator (150 rpm) for shaking 194 over 7 d at 25 ± 1 °C with sampling throughout. Preliminary tests indicated that 195 apparent sorption equilibrium was attained within 3 d. The equilibrium sorption tests 196 197 were hence equilibrated for 3 d with initial concentration of SEs within the range of 100–3000 μ g/L. The supernatant was collected for SEs analysis after standing for 2 d. 198

Following the sorption experiments, the vials with initial concentrations of 500, 1000, and 2000 μ g/L were used to determine the SEs desorption. Background solution was added to each vial to begin the desorption study, and these desorption steps were repeated three times. All samples, including blanks, were run in duplicate. Because of the negligible mass loss of sorbates confirmed in the blanks (<2%), SEs sorption uptake by all samples was calculated based on mass difference.

205 2.5. Bioavailability experiment

To explore the bioavailability of SEs under different sorption mechanisms and 206 aging conditions simultaneously, soil, HA1, and HM were pre-equilibrated with SEs 207 solutions (E1 or 17 β -E2 1000 µg/L, 0.01 M CaCl₂) at 25 ± 1 °C for 3, 15, and 30 d in 208 dark, respectively. The soil, SOM and background solution were all autoclaved at 209 120 °C for 30 min three successive times in the aging processes to prevent SEs 210 biodegradation. Before the bioavailability experiment, the suspension was centrifuged 211 at 5000 rpm for 5 min, and then the supernatant was collected and an equal amount of 212 soil microbial suspension was added to the residues. The SEs content in supernatant 213 was detected to calculate the concentration of Ses sorbed on soil or SOM, which was 214 used as the initial concentration of bioavailability experiment. All treatments were 215

incubated in dark at 25 ± 1 °C, 150 rpm for 14 d. Samples containing both sorbent and aqueous phases were collected at different periods and analyzed using GC-MS to determine the biodegradation of SEs. Each treatment included two replicates.

219 2.6. Modelling of sorption kinetics and isotherms

The sorption kinetic data were fitted with a two-compartment first-order kinetic model (Johnson et al., 2001):

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$$q_t = q_e [f_1 (1 - e^{-k_1 t}) + f_2 (1 - e^{-k_2 t})], \qquad (1)$$

where t[h] is the sorption reaction time; $q_t [\mu g/g]$ is the SEs concentration in the sorbent at time t; $q_e [\mu g/g]$ is the equilibrium concentration of SEs in the sorbent; k_1 and $k_2 [1/h]$ are apparent first-order rate constants for the quick and slow sorption fractions, respectively; and f_1 and f_2 are the contribution rate of quick and slow sorption, respectively, where $f_1 + f_2 = 1$. The sorption equilibrium data were fitted with the logarithmic form of the Freundlich model:

$$\log q_e = \log K_f + n \log C_e, \tag{2}$$

where C_e [µg/L] is the equilibrium aqueous concentration; K_f [(µg/g)/(µg/L)ⁿ] is the sorption coefficient; and *n* is a site energy heterogeneity factor, typically used as an isotherm nonlinearity indicator. The concentration dependence of the affinity of the SEs to the sorbent was analyzed by calculating the solid/liquid distribution coefficients (K_D , L/g) at concentrations (C_e) equal to the aqueous solubility (S_w) of solute (at 0.001 S_w , 0.01 S_w , and 0.1 S_w). The organic carbon content-normalized distribution coefficient (K_{oc}) was calculated as follows:

$$K_D = q_e/C_e, \tag{3}$$

 $K_{oc} = (K_D / \% \text{ organic carbon}) \times 100.$ (4)

The desorption hysteresis index (HI) was obtained by calculating the ratio of linear index of the sorption–desorption isotherm, which was fitted with the Freundlich model:

$$HI_i = N_{Di}/N_S, \tag{5}$$

where N_S and N_{Di} are the linear indexes of the sorption isotherm and desorption

243 isotherm, respectively.

The degradation of SEs was fitted to a first-order kinetic equation:

245 $\ln C/C_0 = -kt$, (6)

where C_0 and C [µg/g] are concentration of SEs at the initial time and time *t*, respectively, and *k* is the degradation rate constant. The half-life ($t_{1/2}$) of SEs was computed from the rate constant using the following equation: $t_{1/2} = ln2/k$.

249 2.7 Statistical analysis

A one-way analysis of variation (ANOVA) and a least significant difference (LSD) multiple comparisons (p < 0.05) were used to assess the significant difference among different treatments (sorbents, SEs and so on). The Pearson correlation analysis was performed to measure the pairwise relationship between different variables (i.e., TOC, quick sorption, slow sorption, micropore volume and others).

255 **3. Results and discussion**

256 *3.1. Characteristics of SOM fractions*

The FTIR spectra of soil and its sequential alkaline-extraction fractions showed 257 some differences (Fig. 1). Observed functionality included (Pérez et al., 2004; Smidt et 258 al., 2007) the broad band at 3620 cm⁻¹ ascribed to O-H stretching vibration; the peaks 259 at 2950 and 2860 cm⁻¹ corresponding to aliphatic C-H stretching; the absorption peak 260 at 1710 cm⁻¹ attributed to C=O stretching of COOH and ketones; and the peaks at 1450, 261 1510, and 1645 cm⁻¹ attributed to structural vibrations of C=C within the aromatic ring 262 skeleton. In addition, the strong peak at 1030 cm⁻¹ was associated with stretching 263 vibration of C-O in aromatic ethers, carbohydrates, or polysaccharides, and that at 1110 264 cm⁻¹ was attributed to the stretching vibrations of C-O in aliphatic ethers. Thus, soil 265 and SOM fractions all contained prominent aliphatic carbon (3000-2800 cm⁻¹) 266 alongside aromatic carbon (1640 cm⁻¹). 267

The aliphatic carbon and aromatic carbon accounted for 31.2%–43.9% of the total organic carbon (TOC) in bulk SOM measured by the ¹³C NMR (Table S2; Fig. 1), which

was low than their surface content of 54.8%–62.1% determined using XPS (Table S2; 270 Fig. S1). These results indicated that more other O-containing polar functionalities were 271 distributed within the SOM interior, which was demonstrated by the higher bulk 272 polarity (O+N/C) compared with the surface polarity (Table S2). The total aliphatic 273 carbon content of bulk (0-109 ppm) and surface both increased with the sequential 274 extraction of HA (Table S2; Fig. 1), but aromatic carbon and O-containing polar 275 functionalities contents declined. Together these results indicate that aliphatic carbon 276 277 is more strongly bound to soil minerals than aromatic carbon (Zhang et al., 2014; Yang et al., 2012; Wang et al. 2011). The increasing C/N values (taken to be indicative of 278 higher stability of SOM) indicated that stability increased with HA isolation (HA1 <279 HA2 < HM; Table 1) (Zhu et al., 2005). Our work is consistent with previous studies 280 that showed that stability, compacted structure, condensation degree, and 281 intermolecular intersections of SOM increase in the following order: HA < HM < 282 kerogen < black carbon (Yang et al., 2004; Cornelissen et al., 2005). 283



HM HA2 HA1 250 200 150 100 50 0 13C Chemical shift (ppm)

Fig. 1. FTIR spectra of soil, HAs (HA1 and HA2), and HM.

Fig. 2. Solid-state ¹³C NMR spectra of HAs (HA1, HA2) and HM.

284 *3.2.* Slow sorption, sorption nonlinearity, and desorption hysteresis

Rates of E1 and 17β -E2 sorption on soil components were relatively high, with 80% of the equilibrium sorption capacity reached within 24 h and the apparent sorption equilibrium attained in 72 h (Table 1, Fig. 3). A typical two-stage process of quick sorption was evident, followed by a slow sorption process (Fig. 3). Quick sorption accounted for over 70% of sorbed mass occurring within 12 h (except for HM). 290 Thereafter, the contribution rate of slow sorption gradually increased until the sorption equilibrium was achieved. The quick sorption rate constant k_1 exceeded the slow 291 sorption rate constant k_2 by 1–2 orders of magnitude (Table 1). For the same sorbent, 292 the contribution rate of slow sorption (f_2) of 17 β -E2 was slightly higher than that of E1. 293 Comparison of the f_2 value of SEs in different sorbents showed that the f_2 of HM and 294 soil were highest, but quick sorption dominated in HA1 (Table 1). This also supports 295 the influence of progressive SOM fractionation on SEs sorption; with the sequential 296 297 extraction of SOM from HA1, HA2, and then HM, SEs sorption shifted from quick sorption (f_1) dominance to increasing slow sorption (f_2) contributions reaching similar 298 levels. 299

300 **Table 1**

301 Two-compartment first-order kinetics model parameters for E1 and 17β -E2 sorption in

soil and SOM.

C - ul - ut	E1				17β-E2					
Sorbent	<i>k</i> 1	<i>k</i> ₂	f_l	f_2	R ²	k_1	<i>k</i> ₂	f_l	f_2	R ²
Soil	3.93	0.09	0.51	0.49	0.999	2.81	0.05	0.49	0.51	0.998
HA1	4.92	0.10	0.74	0.26	0.996	9.30	0.11	0.69	0.31	0.998
HA2	0.88	0.05	0.69	0.31	0.986	0.60	0.03	0.46	0.54	0.995
HM	13.61	0.09	0.49	0.51	0.980	15.35	0.16	0.48	0.52	0.982



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311 The sorption and desorption processes fitted well to the Freundlich equation (all $R^2 > 0.96$) (Fig. 4 and S2; Tables S3 and S4), which is consistent with previous SEs 312

quick and slow sorption rate contributions.

Fig. 3. The batch data and simulated two-compartment first-order kinetic model fittings

of E1 and 17β -E2 sorbed on soil and SOM, with inset graphs showing the simulated

sorption studies. Nonlinear sorption isotherms with n < 1 were obtained for both SEs, 313 especially in soil and HM. The *n* value of E1 was greater than that of 17β -E2 (except 314 for HA2) with the same sorbent (Table S3), and hence nonlinear sorption of 17β -E2 315 was more evident. The sorption nonlinearity of humic substances also increased with 316 the sequential extraction of HA, consistent with the increased contribution rate of slow 317 sorption (f_2) . It is reasonable to suggest that slow sorption was primarily responsible 318 for the observed nonlinear sorption. The sorption capacity also increased with 319 sequential HA extraction (Table S3). The $\log K_{oc}$ values of 17 β -E2 were higher than 320 those for E1, consistent with its greater hydrophobicity. The desorption hysteresis index 321 (HI) was positively correlated with initial SEs concentration (Table S4). A consistently 322 slightly lower HI of 17 β -E2, when compared to E1 for the same sorbent, indicates its 323 greater desorption hysteresis, and hence its more difficult desorption with a higher 324 proportion of irreversible sorption sites. The HI values of sorbents increased with 325 sequential HA extraction, indicating that desorption became increasingly difficult and 326 the proportion of irreversible sorption sites also increased. 327



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Fig. 4. Freundlich sorption isotherms of E1 and 17 β -E2 by soil, HAs and HM, respectively.

330 3.3. Impact of sorbent elemental composition, polarity, functionality, and spatial 331 arrangement

The $\log K_{oc}$ of E1 and 17 β -E2 by soil and humic substances was positively correlated with TOC (p < 0.05) (Fig. 5a and 5b), and the $\log K_{oc}$ of 17 β -E2 was higher than that of E1, which was consistent with a higher $\log K_{ow}$ value of 17 β -E2 compared

with E1 (Fig. 5c). The hydrophobic interaction of SEs in SOM hence played a critical 335 role in their sorption. With the sequential extraction of SOM, the $\log K_{ov}/\log K_{ow}$ value 336 of 17β -E2 was slightly higher than that of E1 in soil and HA1, but with the opposing 337 trend found in HA2 and HM (Fig. 5d). This was probably related to the functionality 338 composition of hydrophobic organic compounds generally (including SEs) and their 339 interaction with the specific sorption sites (such as hydrogen bonds and π - π bonds) of 340 condensed organic matter in soil (Zhu et al., 2004, 2005; Sun et al., 2010; Takigami et 341 al., 2011). 342

Linear and quick sorption of HOCs is generally expected in the SOM amorphous 343 organic matter domain, and nonlinear, slow sorption to the crystalline SOM (Huang et 344 al., 1997; Johnson et al., 2001). The ¹³C NMR spectra of humic substances showed that 345 the crystalline carbon (33 ppm) became more prominent compared with amorphous 346 carbon (30 ppm) with the increased sequential extraction of humic substances (Fig. 2) 347 (Chen et al., 2017). The nonlinear sorption can hence be ascribed to the increase in 348 condensed organic matter content and the influence of these sorption sites. However, 349 350 the nonlinearity (n) of SEs in the bulk soil was also relatively high, which can be reasonably ascribed to the complicated composition and structure and increased 351 heterogeneity. The chemical π - π interactions and hydrogen bonding also contributed to 352 the nonlinear sorption due to the aromatic ring, hydroxyl, and ketone groups for both 353 E1 and 17β-E2 molecules (Kim et al., 2016). However, hydroxyl groups may form 354 stronger hydrogen bonds than their ketone counterparts, and the two hydroxyls within 355 17β -E2, versus E1's single hydroxyl and single ketone group, may cause increased 356 hydrogen bonding of the former to the SOM and explain the greater nonlinearity of 357 17β-E2 sorption observed. Therefore, the contents of SOM and its hydrophobic 358 distribution are the decisive factors controlling SEs adsorption capacity; however, the 359 difference in molecular structure between E1 and 17β-E2 and their specific sorption on 360 SOM via the hydrogen bond and π - π bond resulted in the different sorption nonlinearity. 361





Micropore area and volume of sorbents can be used to examine the sorption 368 mechanism of pore filling, which is the main process responsible for slow sorption and 369 subsequent desorption hysteresis due to accessibility constraints (Pan et al., 2006). The 370 contribution rate of the slow sorption of SEs in soil and SOM were positively related to 371 the micropore volumes of sorbents ($R^2 > 0.80$, p < 0.05), but poorly correlated with 372 TOC content ($R^2 < 0.1$, p > 0.05) (Fig. S3a–d). It can be concluded that the slow sorption 373 was mainly induced by pore filling. The micropore volumes of sorbents was negatively 374 correlated with HI (p < 0.05) (Fig. S3d). It is speculated that more SEs can enter the 375 sorption sites in micropores with larger micropore volumes, which cannot desorb, 376 leading to enhanced desorption hysteresis (Dai et al., 2022). The sorption capacity 377

 $(\log K_{oc})$ of SEs was negatively correlated with the surface and bulk polarity of sorbents 378 (p < 0.05) (Fig. S3e, f). Sorbents of higher polarity generally contain abundant 379 hydrophilic oxygen-containing moieties, which can provide sites for water cluster 380 formation at their surface through H-bonding (Wang et al., 2011). Water clusters may 381 further reduce the surface hydrophobicity of sorbents and the accessibility of those 382 383 HOCs to sorption sites, as well as compete with them; thus, the sorption of both E1 and 17β -E2 may be blocked by the high polar components on the sorbent surface. The 384 $\log K_{oc}$ of E1 and 17β-E2 was positively correlated with bulk alkyl carbon in SOM (p < 1385 0.05) (Fig. S3g), which demonstrated that the aliphatic carbon content of SOM was a 386 key factor regulating the sorption of SEs. Although the soil had the highest bulk H/C, 387 its TOC content was the lowest of the studied sorbents, resulting in the lowest $\log K_{oc}$ 388 value. This further indicated that SOM was the main factor controlling SEs sorption in 389 soil. The ratio of surface aliphatic carbon was lower than the bulk, indicating that the 390 aliphatic carbon was mainly distributed in the interior parts of sorbent rather than at the 391 surface. The sorption capacity ($\log K_{oc}$) of E1 and 17B-E2 was also positively correlated 392 393 with the content of aliphatic carbon of the sorbent surface and negatively correlated with the content of surface aromatic carbon (Fig. S3h), which further demonstrates that 394 the content of aliphatic carbon in SOM and the conformation of SOM were the key 395 factors governing SEs sorption. The overall impact of sorbent elemental composition, 396 polarity, functionalities, and their spatial arrangement on SEs sorption was significant 397 and complex. Especially, the spatial arrangement characteristics of functionalities by 398 the increase of surface hydrophobic alkyl carbon, and the decrease of surface polarity 399 with sequential alkaline-extraction of SOM contributed to more SEs sorption. 400

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3.4. Response of SEs bioavailability to sorption and aging processes with SOM

The biodegradation of E1 and 17 β -E2 in soil, HA1, and HM were well fitted with the first-order degradation kinetics model ($R^2 > 0.85$) (Fig. 6; Table S5). The degradation rate constant (k) and degradation ratio of SEs showed no significant correlation with the TOC of the sorbent (p > 0.05) (Fig. S4), indicating that the difference in SE biodegradation in soil and SOM could not be attributed to the TOC

content. The HOCs are first sorbed on the soil surface, into the large pores, or through 407 hydrophobic interaction into amorphous organic matter of SOM. This is followed by 408 the slow and gradual adsorption of SEs into micropores or condensed organic matter of 409 SOM that cannot be entered by microorganisms. Therefore, it could be inferred that the 410 bioavailability of SEs by quick sorption is higher than that of slow sorption. The content 411 of condensed organic matter increases with sequential alkaline-extraction, and the k412 value and degradation ratio of E1 and 17β-E2 were both positively correlated with the 413 414 contribution rate of fast sorption, but negatively correlated with the contribution rate of slow adsorption (Table 2). This indicates that the rapid degradation in the early stage is 415 associated with the SEs in the dissolved phase or in fast sorption, as both gradually 416 reduce as the degradation rate decreases. Subsequently, the SEs of slow sorption are 417 used by microorganisms until desorption stops, giving a stable degradation ratio. This 418 further indicates that the irreversible sorption and strong desorption hysteresis of 419 estrogen on HM with the maximum content of condensed matter organic carbon and 420 micropore volume contribute to the low bioavailability, which is the key controlling 421 422 factor led to the differences in SEs bioavailability among these sorbents. Overall, these results suggested that the hydrophobic interaction was the key mechanism of SEs 423 sorption and was characterized with quick sorption and high bioavailability. However, 424 micropore filling and its specific sorption in condensed SOM were likely the main cause 425 of the slow or irreversible sorption and desorption hysteresis, which also contributed to 426 the low bioavailability. 427



Fig. 6. Effect of the aging time (3 d, 15 d, and 30 d) on E1 and 17β -E2 degradation in soil and SOM fractions (HA1 and HM).

With the increase of aging time, the bioavailability and biodegradation rate 434 constant decreased, and the degradation half-life increased (Table S5). In the aging 435 process, the bioavailability of SEs was greatly reduced because the SEs have more time 436 to diffuse into soil micropores and condensed organic matter, leading to an increase in 437 the proportion of irreversible sorption and the blocking of the exposure pathway of SEs 438 to microorganisms. This is confirmed by the well-developed micropores and high 439 content of condensed organic matter in HM. For the same sorbent, the degradation rate 440 of 17β -E2 was lower than that of E1 at different aging times. This may be related to the 441 strong hydrophobicity of 17β-E2 and its strong affinity with soil and SOM. In addition, 442 the 17β-E2 molecule contains two hydroxyl groups and has a stronger specific binding 443

ability (such as hydrogen bonding) to sorbents than E1, which may be the reason for the lower bioavailability of 17β -E2.

446 **Table 2**

447 Relationship between sorption kinetic parameters and degradation kinetic parameters

448 after aging 3 days.

SEs	sorbents	k (h ⁻¹)	degradation ratio (%)	contribution rate of quick sorption	contribution rate of slow sorption
	HM	0.007	88.0	0.49	0.51
E1	Soil	0.009	96.3	0.51	0.49
	HA1	0.012	97.2	0.74	0.26
	HM	0.008	81.5	0.48	0.52
17β-E2	Soil	0.012	89.8	0.49	0.51
	HA1	0.021	91.7	0.69	0.31

449 **4.** Conclusions

Application of soil sequential extraction techniques provide an important means 450 of assessing the contributions of the various SOM components to the sorption, 451 desorption, and bioavailability of SEs in real agricultural soils. These findings provide 452 an improved understanding of the risks posed to agricultural environment by SEs. The 453 sorption of E1 and 17β-E2 was dominated by hydrophobic partitioning and quick 454 455 sorption. However, this does not negate the importance of other sorption mechanisms. Specific sorption (hydrogen bond and π - π bond) interactions of SEs with the SOM and 456 micropore filling were responsible for nonlinear and slow sorption, as well as 457 desorption hysteresis. There was a significant effect of sorption on the bioavailability 458 of SEs. In particular, the irreversible sorption in microporous and condensed organic 459 matter improved the environmental persistence of SEs. This effect was significantly 460 enhanced with an increase in aging time. 461

The gradual accumulation and distribution of SEs in agricultural soils receiving manures, and the potentially greatly delayed release of SEs, are each influenced by the balance between sorption on the SOM composition and the degradation of microorganisms. Even with assumptions of simple linear sorption control, lag times of

decades have been predicted by Gall et al. (2016) for the release of SEs from 466 agricultural soils, with attendant risks posed over these timeframes to groundwater. The 467 results of the current study suggest that these timeframes will be further extended 468 because of the need to additionally consider the hysteretic, slow, non-linear desorption 469 of SEs from the soil matrix. In particular, a fraction of SEs is subjected to pore filling 470 sorption within any crystalline carbon present, which was characterized by high 471 persistence due to the low bioavailability. Under these conditions, establishing the SEs 472 473 sorption on different SOM fractions associated with the response of SEs bioavailability is necessary to predict the long-term risks of SEs in agricultural environment. 474

475 CRediT authorship contribution statement

476 Xiaoming Song: Experiment, Data curation, Writing - original draft, Writing 477 review & editing. Zhipeng Zhang: Experiment, Data curation, Writing - review &
478 editing. Yujuan Wen: Experiment & SEs detection. Wei Zhang: Writing - review &
479 editing. Yi Xie: Modeling. Nan Cao: Data curation, Formal analysis. Dong Sun: Data
480 curation. Yuesuo Yang: Writing - review & editing, Supervision.

481

Declaration of competing interest

We declare that the research was conducted in the absence of any commercial orfinancial relationships that could be constructed as a potential conflict of interest.

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- 489 Appendix A. Supplementary data
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