

1 **Title:** The effect of dietary supplements on core temperature and sweating responses in hot environmental
2 conditions: a meta-analysis and meta-regression.

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4 **Running heading:** Dietary supplements for thermoregulation in the heat.

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6 **Authors:** ¹Jennifer S Peel*, ¹Melitta A McNarry, ¹Shane M Heffernan, ^{1,2}Venturino R Nevola, ^{1,3}Liam P Kilduff,
7 ^{1,3,4}Mark Waldron

8

9 * = corresponding author

10 ¹A-STEM Centre, Faculty of Science and Engineering, Swansea University, Swansea, UK.

11 ²Defence Science and Technology Laboratory (Dstl), Fareham, Hampshire, UK.

12 ³Welsh Institute of Performance Science, Swansea University, Swansea, UK.

13 ⁴School of Health and Behavioural Sciences, University of the Sunshine Coast, Sippy Downs, QLD, Australia.

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15 **Details for the Corresponding Author:**

16 Jennifer S Peel

17 856558@swansea.ac.uk

18 ORCID: 0000-0002-7651-8979

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29 **Abstract**

30 Dietary supplements are widely used among individuals exposed to hot environments, but whether their
31 consumption confers any thermoregulatory effect is unclear. Therefore, we systematically evaluated the effect of
32 dietary supplementation on key aspects of thermoregulation (core temperature [T_{core}] and sweating responses) in
33 the heat. Three databases were searched in April 2024. After screening, 124 peer-reviewed articles were identified
34 for inclusion within three separate meta-analyses: (1) peak T_{core} ; (2) whole-body sweat rate (WBSR); (3) local
35 sweat rate (LSR). The moderating effect of several variables (e.g. training and heat acclimation status), known to
36 influence thermoregulatory function, were assessed via sub-analysis and meta-regression. There was no overall
37 effect of the differing supplement types on WBSR ($p = 0.405$) and LSR ($p = 0.769$), despite taurine significantly
38 increasing WBSR ($n = 3$, Hedges' $g = 0.79$, $p = 0.006$). Peak T_{core} was significantly affected by supplement type
39 ($p = 0.011$), primarily due to caffeine's *small* significant positive effect ($n = 30$; Hedges' $g = 0.44$, $p < 0.001$) and
40 taurine's ($n = 3$, Hedges' $g = -0.66$, $p = 0.043$) and oligonol's ($n = 3$; Hedges' $g = -0.50$, $p = 0.014$) *medium*
41 significant negative effects. Dietary supplements, such as amino acids (e.g. taurine), some anti-oxidants and anti-
42 inflammatory (e.g. oligonol) conferred the greatest thermoregulatory benefits during heat exposure. Taurine
43 ingestion in such conditions may lower heat strain, which is likely through its augmentation of thermal sweating.
44 Conversely, caffeine intake may potentially pose the greatest risk in the heat due to its effect on T_{core} .

45 **Key words:** Dietary supplements; thermoregulation; core temperature; sweating; evaporative cooling; heat

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57 **1. Introduction**

58 Adult humans rely on eccrine sweat production to facilitate evaporative cooling and maintain thermal balance,
59 particularly in hot and/or humid environments (high wet-bulb globe temperature [WBGT]; (1, 2)). In hot
60 conditions, evaporation typically represents the primary heat transfer avenue (2), which offsets heat production at
61 rest and during exercise or occupational work (3, 4). If heat is not sufficiently dissipated from the body to the
62 environment, positive heat storage ensues, leading to rises in core temperature (T_{core} ; uncompensable heat stress)
63 (5). If left uncorrected, heat strain can cause heat exhaustion, heat stroke and even death in extreme scenarios (6,
64 7). Thus, understanding factors that control thermal sweating is of great importance for the health and wellbeing
65 of many people.

66 Thermoregulatory capacity is largely determined by three primary modifiable factors: metabolic heat production,
67 vasodilation (i.e. dry heat loss) and sweating (i.e. evaporative heat loss; (1, 8, 9)). Consequently, the ability to
68 activate thermoregulatory defenses (i.e. thermoeffectors) and to tolerate exposure to hot environmental conditions
69 can be improved, with sweating being the primary manipulable pathway (2, 10, 11). For example, endurance
70 training and heat acclimation regimes are capable of lowering resting T_{core} , and the oxygen cost of exercise at a
71 given intensity, but are notable in their capacity to accelerate sweating onset and increase sweat rate, plasma
72 volume and skin blood flow (SkBF) (10-14). In various ways, these physiological adaptations augment avenues
73 of heat transfer, control heat production and, ultimately, aid in maintaining thermal equilibrium during heat
74 exposure. Given the importance of thermal sweating in achieving this, further understanding of the capacity for
75 adaptation in sweating variables in response to various interventions is required.

76 More recently, the notion that dietary supplementation may offer thermoregulatory benefits or, alternatively,
77 heighten the risk of heat illness when ingested in hot conditions has been considered (15-17). There are a number
78 of motivations for individuals to consider dietary supplementation, such as ensuring adequate intake of certain
79 nutrients, improving health, or supporting specific physiological functions (18, 19). Approximately 50% of US
80 adults (20) and between 15 to 41% of UK adults (21) report dietary supplement use, with only a quarter of users
81 taking supplements that have been recommended by a healthcare professional (22). Whilst such dietary
82 supplements are not commonly consumed for the purpose of influencing thermoregulation, they may inadvertently
83 affect it (16). As the popularity of dietary supplements continues to rise, in a world which is likely to experience
84 more frequent, prolonged and intense heatwaves (23), research is needed to better understand the potential
85 thermoregulatory effects upon human health and performance. For example, the amino acid taurine, often ingested

86 for its anti-oxidative and anti-hypertensive effects (24, 25), has more recently been reported to increase sweating
87 rate/loss (a key modifiable heat dissipation pathway) by approximately 13% (26) and 27% (27) as well as reducing
88 T_{core} compared to placebo in the heat (26, 28). Furthermore, another dietary supplement, creatine, is not typically
89 considered to help in offsetting hyperthermia, but is commonly taken to improve high-intensity exercise
90 performance (29). However, a review of its thermoregulatory effects highlighted that supplementation may be
91 beneficial during exercise in high ambient temperatures due to its effects on fluid balance (17). Additionally, a
92 recent meta-analysis established that pre-exercise hyperhydration with glycerol and/or creatine supplementation
93 decreased the rate of rise in T_{core} after constant work exercise in both thermoneutral and hot conditions, compared
94 to placebo (15).

95 Other commonly used supplements, such as dietary nitrate (NO_3^-), which has a key role in blood pressure
96 regulation and endurance exercise enhancement (30, 31), does not appear to maintain all of these effects when
97 humans are exposed to the heat (16). This is surprising, as there is a plausible mechanistic basis for
98 thermoregulatory enhancement following ingestion of dietary nitrate and L-arginine, as both are known to improve
99 NO (nitric oxide) bioavailability (32, 33). Specifically, NO bioavailability could have direct and indirect effects
100 on eccrine sweat gland and microvascular function (34, 35). Indeed, other supplements, such as anti-oxidants (i.e.
101 polyphenols), may support thermoregulation through protection of NO against oxidative destruction, thereby
102 improving its bioavailability (36) and enhancing or preserving peripheral vasodilation. However, given that body
103 fluid loss, and secondary hypovolemia, is accelerated in the heat (37), the reported reductions in blood pressure
104 following supplementation with NO donors (38) could increase the risk of acute hypotension, particularly in the
105 post-exercising state (39).

106 Branched-chain amino acids (BCAAs) have been extensively researched for their potential ergogenic role amongst
107 athletes (40), yet have several health-related applications (41) and can be supplemented to account for age-related
108 decline in lean muscle mass (42). Whilst less commonly supplemented for such reasons among the general
109 population (43), BCAAs and other amino acids have a wide variety of biological roles. For example, tyrosine is
110 used to enhance cognitive function (44, 45) and BCAAs have been reported to alleviate skeletal muscle damage
111 and soreness following exhaustive and resistance exercise (46, 47). Given that both tyrosine and BCAAs may
112 compete for the same blood-brain-barrier transporters, coupled with their wider roles in neurotransmitter
113 biosynthesis pathways (48-50), sufficient balance of both supplements may be important during heat exposure. In
114 a previous meta-analysis, the use of orally administered tyrosine or BCAAs (used separately), were capable of
115 enhancing endurance exercise performance in the heat, but there was no effect on sub-maximal or maximal T_{core}

116 responses (16), thereby questioning their thermoregulatory role. Whilst many of the above-mentioned
117 supplements are used more modestly across the population (43, 51), caffeine features in the daily intake of
118 approximately 80 to 85% of people globally (52, 53) and is a prominent dietary supplement among athletes (54).
119 However, caffeine has been reported to increase T_{core} when ingested before or during exercise in the heat (16), but
120 its effects in the resting state have not been evaluated meta-analytically. Given the high prevalence of caffeine
121 consumption, mixed with the understanding of its cardiometabolic side-effects (55, 56), this perhaps places one
122 of the greatest risks to the general population when consumed in the heat. Collectively, it is apparent that
123 supplementing the diet with some substances, could have implications for thermoregulatory capacity, and further
124 research is required to understand the consistency and magnitude of effects reported across the empirical literature.

125 Based on the evidence, to date, a systematic evaluation of the effect of all dietary supplements on the primary
126 modifiable thermoregulatory process of sweating, and subsequent T_{core} responses, is warranted. This is necessary
127 to provide clarity on the magnitude and consistency of the effect of supplements on thermal balance during rest
128 and exercise. This impact has not previously been fully considered, and there remains limited official guidance on
129 dietary supplement intake for those exposed to thermally stressful conditions, such as athletes (57-59), and
130 military personnel (60) or, indeed, the general public (61). Given the range of effects that different supplements
131 appear to have on T_{core} , at least in the exercising state in the heat (16), coupled with the clear lack of specific
132 guidance on this topic, a comprehensive evaluation of the collective evidence is an important step in developing
133 an evidence-based understanding of the benefits or risks associated with using dietary supplements in hot
134 conditions.

135 The aims of the current meta-analysis were to investigate the effects of all known orally administered dietary
136 supplements on T_{core} and sweating responses in the heat. The effect of rehydration solutions, such as electrolytes,
137 on thermal sweating have been thoroughly evaluated (37, 62, 63) and this was not replicated here; however, a
138 number of factors were considered as moderators of T_{core} and sweating responses, such as hydration status among
139 participants in studies evaluating dietary supplementation in the heat (64, 65). Likewise, training and
140 acclimatization/acclimation status (11), protocol (rest vs exercise) and exercise intensity were considered to
141 potentially impact thermoregulatory sweating and T_{core} (66-68). Environmental conditions, such as WBGT (and/or
142 heat stress index; and vapor pressure) will also influence the ability to evaporatively cool (69, 70). Therefore, to
143 evaluate the effects of dietary supplements on thermoregulation in the heat, these factors were considered as
144 potential moderating variables and formed part of a secondary meta-regression analysis.

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146 **2. Methods**

147 **2.1 Search strategy**

148 All of the available literature was searched and obtained according to the PRISMA guidelines, with a
149 predetermined search strategy (71). Medical subject heading (MeSH) terms were active during the searches. There
150 was no limit on the status, date or language of the publication. The final Boolean searches were performed in
151 PubMed, SPORTDiscus (EBSCO) and Scopus on 9th April 2024. The search terms used were '(dietary
152 supplements OR dietary supplementation OR nutritional supplements OR nutritional supplementation OR
153 supplements OR supplementation OR ergogenic OR ergogenic aids OR nutraceuticals OR amino acids OR anti-
154 oxidants OR vitamins OR minerals OR stimulants OR herbs OR herbal) AND (heat OR temperature OR sweat
155 OR sweating OR sweat response OR sweating response OR sudomotor OR body temperature regulation OR
156 thermoregulation OR thermoregulatory OR heat loss OR cooling OR evaporative OR evaporation OR thermal
157 stress OR heat stress OR hyperthermia OR hyperthermic)'. As there is no *a-priori* list of dietary supplements that
158 effect thermal balance, no supplements were searched individually by name. Two authors (JP and MW) verified
159 the search terms and the accuracy of the returned results. 'Other sources' were also identified, such as through
160 social media (Twitter or 'X'), the reference lists of included papers and additional database (Google Scholar)
161 searching using various combinations of the above search terms.

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163 **2.2 Study selection**

164 Any duplicates were removed, and titles and abstracts were screened for inclusion by two investigators (JP and
165 MW), in accordance with agreed inclusion criteria. The single paper retrieved which had been published in a
166 language other than English was translated digitally using two separate translation software programs; Google
167 Translate and DeepL Translator (DeepL GmbH, Cologne, Germany). The reference lists of the initial papers were
168 reviewed independently by two authors (JP and MW). The remaining articles were then assessed separately (and
169 without influence) by JP and MW against the inclusion and exclusion criteria. There was 100% agreement in study
170 selection between the two reviewers. Papers were required to have been published in a peer-reviewed journal as
171 original research articles with a cross-over, randomized control trial, an intervention or an independent groups
172 design. They must also have included a control or placebo group, and participants were required to be healthy

173 adults (≥ 18 years). To be included in this analysis, the studies must have: (1) administered a dietary supplement
174 (by the definition below); (2) been conducted in an ambient dry-bulb temperature of ≥ 30 °C or WBGT ≥ 20 °C
175 or small ranges up to those temperatures in either a laboratory or field setting. A WBGT of ≥ 20 °C was considered
176 to provide sufficient heat stress, even when dry-bulb temperature was < 30 °C (72). Of the remaining papers, 71
177 were removed for the reasons outlined in Figure 1, which were primarily that they included supplements that were:
178 a drug; not orally administered; a macro-nutrient or a rehydration solution (e.g. electrolytes or a supplement with
179 a mechanism of action considered to be directly related to hydration). Other reasons were the absence of measures
180 of T_{core} and the sweating response, or environmental issues.

181 A dietary supplement was defined by adapting the IOC position statement (58) and the European Food Safety
182 Authority statement (19) as: a non-food, non-pharmacological, food component, nutrient or non-food compound
183 that is purposefully orally ingested in addition to the habitual diet, for its nutritional or physiological effects. This
184 may be to maintain sufficient intake of certain nutrients, correct deficiencies, or support physiological function,
185 including thermoregulatory responses to the heat. The supplement is not being consumed for its calorific value,
186 its effects on hydration (the mechanism of action is not through rehydration) and is not an energy drink. Ingestion
187 of the supplement is also recognized to be legal as per the Misuse of Drugs Act 1971 (73) and is not on the World
188 Anti-Doping Association's prohibited substances list (74).

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190 **2.3 Data extraction and quality assessment**

191 Data were manually extracted independently by two authors (JP and MW) and entered into a custom-designed
192 Microsoft Excel spreadsheet. Any discrepancies were verified by a third independent reviewer. Extracted data
193 included: (1) characteristics of the sample (sex, age, health, training and heat acclimation/acclimatization status);
194 (2) study design; (3) supplement, dose and timing of intake; (4) fluid intake before and during exercise (i.e.
195 hydration status); (5) environmental conditions (temperature and humidity); (6) trial type (i.e. exercise type or rest
196 and length); (7) peak T_{core} (rectal, gastrointestinal, esophageal or tympanic); and (8) bias. Risk of bias was assessed
197 independently by two authors (JP and MW) according to the Cochrane collaboration guidelines (75). Where details
198 of the study were unclear, the authors of the relevant papers were contacted for specific information or to clarify
199 the method that was used. There was 100% agreement between the investigators concerning the outcome of this
200 quality assurance procedure, hence, it was not considered necessary to include a third independent reviewer. There
201 were three outcome measures for this meta-analysis: (1) T_{core} reported at the end of the trial, the end of the

202 exercising portion of the trial or at the point of the highest thermal strain, hereafter referred to as ‘peak T_{core} ’; (2)
203 whole-body sweat rate (WBSR) across the trial or exercising portion of the trial; and (3) local sweat rate (LSR)
204 reported at the end of the trial or at the point of the highest thermal strain.

205

206 **2.4 Statistical analysis**

207 Data analysis was performed by one author (JP). Data were extracted from the qualifying papers in the form of a
208 mean, standard deviation (SD) and sample size (n) for the meta-analysis. Publicly available software
209 (WebPlotDigitizer, Version 4.3) was used to extrapolate any unreported values from the figures to mean and SD
210 data. Where data were expressed as mean and standard error (SE or SEM) or CI, they were converted to mean and
211 SD. Authors of the original research articles were contacted for any missing data; however, if mean data were not
212 accessible, these articles were excluded. If standard error or CI were missing, they were imputed using the sample
213 pooled SD from similar included studies in accordance with Cochrane guidelines (75, 76). There were 17 instances
214 (seven in the T_{core} meta-analysis and 10 in the WBSR meta-analysis) where no dispersion data (SD, SE, SEM or
215 CI) were provided. For selected study designs (i.e. intervention studies with pre-post supplementation), the post-
216 intervention values were extracted as the outcome measures for the ‘supplementation condition’ and the pre-
217 intervention values as the ‘placebo or control condition’ (75). For cross-over trials (within-subject) or independent
218 designs, the outcome measures for the supplementation condition were considered against the placebo or control
219 condition. Standardized mean difference (SMD) was used to compare the results between studies utilizing
220 different protocols and measures. Peak T_{core} outcome data were reported as peak T_{core} ($^{\circ}\text{C}$) or rate of rise ($^{\circ}\text{C}\cdot\text{h}^{-1}$)
221 of T_{core} . Mean, maximum, peak and mean body temperature were also included if peak T_{core} data were not
222 provided. Whole-body sweating response outcome data were reported as WBSR ($\text{mL}\cdot\text{min}^{-1}$) and body mass
223 change (%). Outcome data representing WBSL (i.e. body mass or sweat loss and body mass change), reported in
224 absolute L, mL, kg or g were converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$) using trial length data and WBSR reported in $\text{L}\cdot\text{h}^{-1}$
225 or $\text{mL}\cdot\text{h}^{-1}$ were directly converted to $\text{mL}\cdot\text{min}^{-1}$. LSR outcome data reported in $\text{nL}\cdot\text{min}^{-1}$, were converted to
226 $\text{mg}\cdot\text{cm}\cdot\text{min}^{-1}$ and reported as such.

227 Three meta-analyses were conducted, one for each outcome measure. These were performed in RStudio (R Core
228 Team; (77)) and included 135, 106 and 11 comparison groups for the peak T_{core} , WBSR and LSR meta-analyses,
229 respectively. Not all studies reported T_{core} or a sweating response data, hence, they were excluded from the
230 respective analyses. All data were analyzed with a random-effects model, with heterogeneity assessed using the

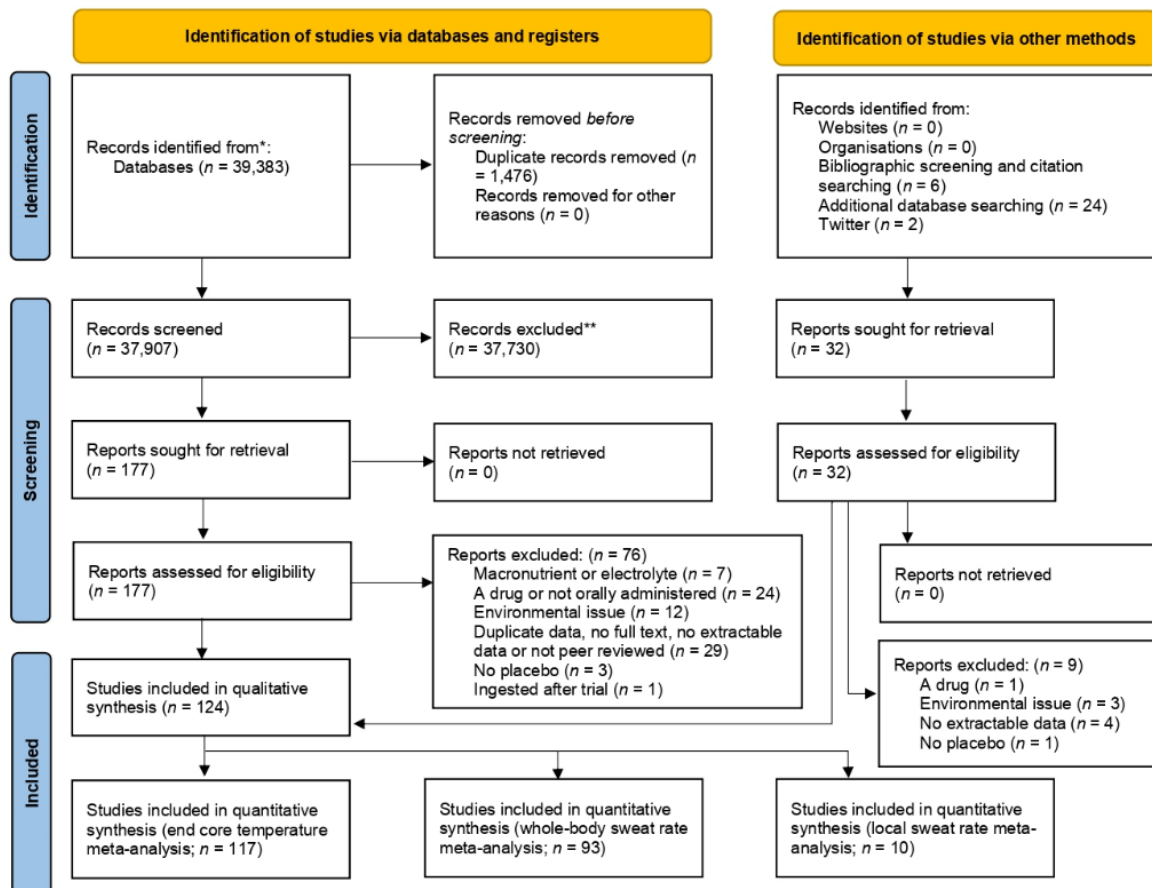
231 I^2 statistic. Outliers were detected using a function in RStudio and influence on analysis investigated. Publication
232 bias was accounted for by funnel plots and conducting Egger's test (78). Any adjustments to the effect sizes based
233 on this procedure are reported in the results. Hedges' g and 95% CIs were used to express SMD between dietary
234 supplementation and placebo groups across studies. Sub-analysis of the different dietary supplements included
235 were conducted for all three meta-analyses. Meta-regressions were also conducted to determine the effect of
236 candidate moderators on peak T_{core} , WBSR and LSR outcomes, as reported in each study: training status (highly
237 trained vs recreationally active); heat acclimation status (heat acclimated vs non-heat acclimated); hydration status
238 (euhydrated vs hypohydrated); fluid ingestion during exercise (fluid ingestion vs no fluid ingestion); duration of
239 trial; WBGT; trial type (exercise vs rest); supplement dose (where sufficient no. of studies) and duration of
240 supplementation (where applicable). The thresholds for the magnitude of effects were < 0.2 , 0.2 , 0.5 and 0.8 for
241 *trivial*, *small*, *medium* and *large* effects, respectively (79). Alpha (α) was set at $P \leq 0.05$ for all analyses.

242

243 3. Results

244 3.1 Study selection

245 The initial searches retrieved 39,383 articles, which were reduced to 37,907 after removal of duplicates. After
246 further screening and removal of reviews, animal studies and other irrelevant papers, 177 articles remained.
247 Searches of social media (Twitter or 'X'), additional databases and reference lists within the 177 papers provided
248 32 further papers. Of the 209 articles, 52 were removed based on their incomplete compliance with the inclusion
249 criteria and a further 33 were removed due to having: no full-text available, duplicate data with another paper or
250 no extractable data. This left 124 papers, of which 117, 93 and 10 papers were included in the peak T_{core} , WBSR
251 and LSR analyses, respectively (Figure 1). Sixteen papers had more than one comparison group and, therefore,
252 one or more additional data sets were added to the analysis for each study. As these additional comparison groups
253 shared participants, the sample size was reduced to mitigate any unit-of-analysis error, as per the Cochrane
254 guidelines (75). Four papers also included multiple comparison groups; however, as these did not share
255 participants, they were included without sample size adjustment. One paper was included without addition of the
256 duplicate peak T_{core} data.



257

258 **Figure 1.** The process of study selection.

259

260 **3.2 Study characteristics**

261 The characteristics of the 124 included studies are summarized in Table 1. The studies included a total of 1,553
 262 participants, comprising both males and females (males 90%; both males and females 9%; unreported 1%) of
 263 varying training (highly trained 43%; recreationally active 40%; unreported 18%) and heat acclimation statuses
 264 (heat acclimated 13%; non-heated acclimated 38%; unreported 49%). One hundred and six studies were cross-
 265 over designs, 12 studies were an independent groups design, and 6 studies were pre-to-post interventions. Thirty-
 266 nine different types of dietary supplements or supplement combinations were included in varying doses (Table 1).
 267 These were a combination of acute doses (single day; n = 81; 65%) and chronic administration (≥ 2 days; n = 43;
 268 35%). The trial types included were exercise (90%) and rest (10%). The measures of T_{core} were rectal (62%),
 269 tympanic (10%), esophageal (9%), gastrointestinal (14%), oral (1%) and unreported (4%). The measures of body
 270 mass or sweat loss or sweat rate, representing WBSR were reported in L (7%), mL (7%), kg (17%) or g (3%),

271 $\text{g}\cdot\text{m}^{-2}\cdot\text{h}^{-1}$ (1%), % change (11%), $\text{L}\cdot\text{h}^{-1}$ (16%), $\text{mL}\cdot\text{h}^{-1}$ (1%) and $\text{mL}\cdot\text{min}^{-1}$ (8%) or were unreported (29%).
272 Ambient dry-bulb temperature (mean 33.8°C; range 25 to 46.6°C), WBGT (mean 27.5°C; range 18.5 to 35.1°C)
273 and RH% (mean 47%; range 12 to 80%) are reported herein. There were no adverse health-related events noted
274 in any of the studies.

275

276 3.3 Meta-analysis

277 The results of the peak T_{core} meta-analysis ($n = 135$) are reported in Figure 2. Overall, the pooled analysis of all
278 supplements revealed that there was a *trivial* non-significant positive effect on peak T_{core} compared to placebo
279 (Hedges' $g = 0.004$, 95% CI -0.091 to 0.100, $p = 0.930$). The I^2 statistic demonstrated 20.6% heterogeneity. The
280 results of the WBSR ($n = 106$) and LSR ($n = 11$) meta-analyses are reported in Figure 3 and Figure 4 respectively.
281 Overall, WBSR (Hedges' $g = 0.041$, 95% CI -0.095 to 0.176, $p = 0.559$) and LSR (Hedges' $g = 0.021$, 95% CI -
282 0.224 to 0.266, $p = 0.869$) had a *trivial* non-significant increase with dietary supplementation compared to
283 placebo, with 1.7% and 0% heterogeneity (I^2), respectively

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Table 1. Summary of studies included in the meta-analyses ($n = 124$).

Study	Design	Sample	Supplement, dose and timing	Temperature and relative humidity	Core temperature method	Trial type	Sweating measure
Caffeine							
Anderson & Hickey (4) ²	Double-blind, counter-balanced, placebo-controlled, cross-over	Moderately trained males ($n = 8$). Age 24 ± 3 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (30 min pre-exercise)	28°C 50% RH WBGT 22.9°C	Rectal every 10 min (PCT)	60 min cycling @ 50% $\dot{V}O_{2\text{max}}$	No sweating response data reported
Beaumont & James (13)	Double-blind, randomized, repeated-measures, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated males ($n = 8$). Age 22 ± 1 years	Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	30°C 50% RH WBGT 24.6°C	Gastrointestinal every 5 min (PCT)	60 min cycling @ 55% W_{max} followed by 30 min TT	Sweat rate (L). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Chevront et al. (34) A and B	Double-blind, randomized, placebo-controlled, cross-over	Healthy, physically active, moderately fit, non-heat acclimated males ($n = 10$). Age 23 (18-37) years	Caffeine $9 \text{ mg}\cdot\text{kg}^{-1}$ (timing not mentioned) A Quercetin 2000 mg (timing not mentioned) B	40°C 20-30% RH WBGT 28-30.1°C	Rectal every 5 min (PCT)	30 min cycling @ 50% $\dot{V}O_{2\text{peak}}$ followed by 15 min TT	Sweat rate ($\text{L}\cdot\text{h}^{-1}$). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Cohen et al. (37) A and B	Double-blind, randomized, placebo-controlled, cross-over	Healthy, heat acclimatized, competitive male ($n = 5$) and female ($n = 2$) runners ($n = 7$). Age 33.3 ± 9.2 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) A Caffeine $9 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) B	WBGT 24-28°C	Tympanic pre and post exercise (PCT)	21 km running TT	Body mass change (%).

Del Coso et al. (49)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, heat acclimated, endurance trained males ($n = 7$). Age 27 ± 1 years.	Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (45 min pre-exercise)	36.0°C $29.0\% \text{ RH}$ WBGT 26.7°C	Rectal every 10 min (PCT)	120 min cycling @ $63\% \dot{V}O_{2\text{max}}$	Sweat loss (L). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Dias et al. (52) ¹ A and B	Double-blind, randomized, independent design	Healthy males ($n = 59$; 20 vs 20 vs 19) Age 21.6 ± 6.9 years	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (6 x 3 $\text{mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$; no timing mentioned) A Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (5 x 6 $\text{mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$; no timing mentioned) B	37.7°C $56.3\% \text{ RH}$ WBGT 32.4°C	Rectal (no T_{core} data reported)	90 min treadmill walking @ $5.6 \text{ km}\cdot\text{h}^{-1}$ with a 5% incline	Body-weight loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Ely et al. (57)	Double-blind, counter-balanced, placebo-controlled, cross-over	Healthy, moderately fit, non-heat acclimated males ($n = 10$). Age 23 (range 19) years	Caffeine $9 \text{ mg}\cdot\text{kg}^{-1}$ (90 min pre-exercise)	40°C $25\% \text{ RH}$ WBGT 29.1°C	Rectal every 5 min (reported mean body temperature; End mean body temperature)	30 min cycling @ $50\% \dot{V}O_{2\text{peak}}$	Sweat rate ($\text{L}\cdot\text{h}^{-1}$). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Falk et al. (58)	Double-blind, placebo-controlled, cross-over	Trained males ($n = 7$). Age 23.8 ± 2.4 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (120 min pre-exercise) and $2.5 \text{ mg}\cdot\text{kg}^{-1}$ (30 min pre-exercise)	25°C $50\% \text{ RH}$ WBGT 20.3°C	Rectal every 15 min (PCT)	Treadmill walking @ $70\text{-}75\% \dot{V}O_{2\text{max}}$ (speed 1.56 s^{-1} with a 22-kg backpack)	Water loss (mL). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Ferreira et al. (62) A and B	Double-blind, randomized, placebo-controlled, cross-over	Well-trained, heat acclimated, male cyclists ($n = 8$). Age 23.9 ± 8.6 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) A Caffeine $9 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) B	30°C average, ranged from $28.5\text{-}32^{\circ}\text{C}$ $71\text{-}78\% \text{ RH}$ WBGT $25.6\text{-}29.7^{\circ}\text{C}$	Tympanic pre and post exercise (PCT)	45 km cycling TT	Body mass loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)

Fujii et al. (65)	Single-blind, randomized, placebo-controlled, cross-over	Healthy, physically active, non-heat acclimatized males ($n = 12$). Age 23 ± 2 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (70 min pre-exercise)	37°C 50% RH WBGT 31°C	Esophageal continuously (T_{core} rate of rise °C/hr)	45 min cycling @ 55% $\dot{V}O_{2\text{peak}}$	Sweat loss (L). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Ganio et al. (74)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, trained, non-heat acclimated male cyclists ($n = 11$). Age 25 ± 6 years	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	33°C 41% RH WBGT 26.1°C	Rectal every 15 min (PCT)	90 min cycling @ 65% thermoneutral $\dot{V}O_{2\text{max}}$ followed by 15 min TT	Sweat rate ($\text{L}\cdot\text{h}^{-1}$). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Gordon et al. (78)	Double-blind, independent design	Healthy, fit males ($n = 10$; 5 vs 5). Age 19.4 ± 1.5 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	26.7°C average, ranged from 24.5-28.9°C 41-54% RH WBGT 18.9-24.1°C	Rectal pre- and post-exercise (PCT)	120 min running	Sweat loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Hanson et al. (83) ² A and B	Single-blind, randomized, placebo-controlled, cross-over	Trained male ($n = 6$) and female ($n = 4$) endurance runners ($n = 10$). Age 26 ± 9 years	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) A Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise) B	30.6°C 50% RH WBGT 25.2°C	Gastrointestinal every 1 km (PCT)	10 km running TT	No sweating response data reported
Hunt et al. (95) ³ A and B	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, non-heat acclimated, caffeine habituated males ($n = 10$) and females ($n = 4$; $n = 14$; A) and caffeine non-habituated males ($n = 8$) and females ($n = 6$; $n = 14$; B). Age	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	30.6°C 31% RH WBGT 22.6°C	Esophageal every 5 s (T_{core} rate of rise °C/hr)	60 min cycling @ 7 $\text{W}\cdot\text{kg}^{-1} \dot{H}_{\text{prod}}$	WBSL (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$). LSR at the back and arm ($\text{mg}\cdot\text{min}\cdot\text{cm}^{-1}$; ventilated)

		27 ± 5 vs 23 ± 3 years					capsule technique)
John et al. (102)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated, non caffeine-habituated males (<i>n</i> = 12). Age 23 ± 4 years	Caffeine 5 mg·kg ⁻¹ (60 min pre-exercise)	35°C 40% RH WBGT 27.6°C	Rectal continuously (PCT)	Cycling @ thermoneutral GET	WBSR (mL·min ⁻¹).
Kazman et al. (109)	Double-blind, randomized, placebo-controlled, cross-over	Healthy males and females (<i>n</i> = 32). Age 27 ± 8 years	Caffeine 7.5 mg·kg ⁻¹ (60 min pre-exercise)	40°C 40% RH WBGT 31.9°C	Rectal (no timing mentioned; PCT)	60 min treadmill walking @ 5 km·h ⁻¹ with 2% incline, followed by a 5 min stepping test (24 steps·min ⁻¹) and 15 deep knee bends	Sweat rate (L·h ⁻¹). Converted to WBSR (mL·min ⁻¹)
Kim & Lee (119) ¹	Randomized, cross-over	Healthy males (<i>n</i> = 9). Age 24.1 ± 3.5 years	Caffeine 3 mg·kg ⁻¹ (60 min pre-trial)	25°C 60% RH WBGT 21.3°C 42°C bath	No T _{core} data reported	30 min water immersion up to umbilical line	WBSL volume (mL). Converted to WBSR (mL·min ⁻¹)
MacNaughton et al. (144) ²	Double-blind, counter-balanced, placebo-controlled, cross-over	Healthy males (<i>n</i> = 6). Age 22 range 19-25 years	Caffeine 5 mg·kg ⁻¹ (no timing mentioned)	28°C 42% RH WBGT 22°C	Rectal (no timing mentioned; PCT)	120 min resting	No sweating response data reported
Millard-Stafford et al. (158)	Double-blind, randomized, repeated-measures, placebo-	Healthy, highly trained male cyclists (<i>n</i> = 16). Age 27.5 ± 7 years	Caffeine 1.2 mg·kg ⁻¹ (0 min pre-exercise) and 3.5 mg·kg ⁻¹ (at 60 min)	28°C 60% RH WBGT 24°C	Rectal every 5 min (PCT)	120 min cycling @ alternating 15 mins of 60 and 70% $\dot{V}O_{2max}$ followed by 15 min TT	Sweat rate (mL·h ⁻¹). Converted to WBSR (mL·min ⁻¹)

	controlled, cross-over						
Nakamura et al. (172)	Double-blind, randomized, placebo-controlled, cross-over	Trained male footballers ($n = 8$). Age 19.9 ± 0.3 years.	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-trial)	31.7°C $63.5\% \text{ RH}$ WBGT 27.9°C	Rectal every 30 s (PCT)	2 x 43 min bouts consisting of 21 cycling intermittent sprints	Sweat volume (L). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Ping et al. (201) ²	Double-blind, randomized, placebo-controlled, cross-over	Recreational, heat acclimated male runners ($n = 9$). Age 25.4 ± 6.9 years	Caffeine $5 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	31°C $70\% \text{ RH}$ WBGT 27.9°C	Rectal every 10 min (PCT)	Treadmill running @ $70\% \dot{V}O_{2\text{max}}$	No sweating response data reported
Pitchford et al. (202)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Highly trained, non-heat acclimated male cyclists ($n = 9$). Age range 22-42 years	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (90 min pre-exercise)	35°C $25\% \text{ RH}$ WBGT 25.2°C	Gastrointestinal continuously (PCT)	Total work cycling TT	Body weight loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Roelands et al. (213)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, trained, non-heat acclimated males ($n = 8$). Age 23 ± 5 years	Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (60 min pre-exercise)	30°C $50\text{-}60\% \text{ RH}$ WBGT $24.6\text{-}25.9^\circ\text{C}$	Rectal every 5 min (PCT)	60 min cycling @ $55\% W_{\text{max}}$ followed by total work TT	Sweat rate ($\text{mL}\cdot\text{min}^{-1}$).
Roti et al. (218) A and B	Double-blind, randomized, independent design	Healthy, active males ($n = 59$; 20 vs 20 vs 19). Age 21.6 ± 3.1 years	Caffeine $3 \text{ mg}\cdot\text{kg}^{-1}$ (6 x 3 $\text{mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$; no timing mentioned) A Caffeine $6 \text{ mg}\cdot\text{kg}^{-1}$ (6 x 6 $\text{mg}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$; no timing mentioned) B	37.7°C $56.3\% \text{ RH}$ WBGT 32.4°C	Rectal every 15 min (PCT)	90 min treadmill walking @ $1.56 \text{ m}\cdot\text{s}^{-1}$ with a 5% incline	Sweat rate ($\text{L}\cdot\text{h}^{-1}$). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)

Stebbins et al. (242) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, active males ($n = 11$). Age range 18-40 years	Caffeine 6 mg·kg ⁻¹ (0 min pre-trial and 45 min pre-exercise)	38°C 40% RH WBGT 30.2°C	Rectal periodically (Mean T _{core})	40 min resting followed by 35 min cycling @ 50% $\dot{V}O_{2max}$	No sweating response data reported
Suvi et al. (246) ¹	Double-blind, randomized, placebo-controlled, cross-over	Healthy, physically active, non-heat acclimated males ($n = 13$) and females ($n = 10$; $n = 23$). Age 24.9 ± 4.1 vs 22.5 ± 2 years	Caffeine 6 mg·kg ⁻¹ (4 mg·kg ⁻¹ 60 min and 2 mg·kg ⁻¹ 0 min pre-exercise)	42°C 20% RH WBGT 29.5°C	Rectal every 1 min (PCT)	50 min treadmill walking @ 60% thermoneutral $\dot{V}O_{2peak}$ followed by TTE	Sweat production (mL·min ⁻¹)
Creatine							
Branch et al. (23)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, competitive male cyclists and triathletes ($n = 7$). Age 38 ± 7 years	Creatine 100 g (5 x 20 g·d ⁻¹)	38°C 35% RH WBGT 29.3°C	Tympanic every 10 min (PCT)	60 min cycling @ 50% $\dot{V}O_{2max}$	Body mass loss (%)
Kern et al. (117)	Double-blind, randomized, independent design	Healthy moderately-highly active males ($n = 20$; 10 vs 10). Age 22.3 ± 3.6 years.	Creatine 335 g (5 x 21 g·d ⁻¹ followed by 23 x 10 g·d ⁻¹)	37°C 25% RH WBGT 26.7°C	Rectal every 15 min (PCT)	60 min cycling @ 60% $\dot{V}O_{2max}$	Body weight loss (%)
Kilduff et al. (118)	Double-blind, randomized, independent design	Endurance-trained, non-heat acclimated males ($n = 21$; 11 vs 10). Age 27 ± 5 vs 27 ± 4 years	Creatine 159.6 g (7 x 22.8 g·d ⁻¹)	30.3°C 70% RH WBGT 27.2°C	Rectal every 5 min (PCT)	Cycling @ incremental work rate at 60-90 rpm	Sweat rate (mL·min ⁻¹)
Mendel et al. (157)	Double-blind, independent design	Healthy, recreationally active, non-heat acclimated	Creatine 100 g (5 x 20 g·d ⁻¹)	39°C 26% RH	Rectal every 10 min (PCT)	40 min cycling @ 55% $\dot{V}O_{2max}$	Weight loss (kg). Converted to

		males ($n = 15$) and female ($n = 1$; $n = 16$ 8 vs 8). Age 26 ± 3.6 vs 26 ± 1.9 years		WBGT 28.5°C			WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Rosene et al. (215) ²	Double-blind, randomized, cross-over	Regularly exercising males ($n = 14$). Age 21.1 ± 1.4 years.	Creatine $0.3 \text{ g}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$ (3 x $0.3 \text{ g}\cdot\text{kg}^{-1}\cdot\text{d}^{-1}$)	32.6°C 18.5% RH	Rectal every 5 min (PCT)	60 min treadmill running @ 60-65% $\dot{V}\text{O}_{2\text{max}}$	No sweating response data reported
Vogel et al. (261) ¹	Randomized, independent design	Healthy, recreationally active, non-heat acclimated males ($n = 16$; 7 vs 9). Age 22 ± 1 years	Creatine 100 g (5 x $20 \text{ g}\cdot\text{d}^{-1}$)	32°C 50% RH WBGT 26.5°C	T_{core} not measured	2 x 75 min exercise (4 x 10 min cycling bouts @ 30% initial sprint resistance at 60 rpm)	Body weight change (%)
Volek et al. (263)	Double-blind, randomized, independent design	Healthy males ($n = 20$; 10 vs 10). Age 23 ± 1 years	Creatine $0.3 \text{ g}\cdot\text{kg}^{-1}$ 7 x $0.3 \text{ g}\cdot\text{kg}^{-1}$)	37°C 80% RH WBGT 34.8°C	Rectal every 5 min (PCT)	15 min cycling @ 70% $\dot{V}\text{O}_{2\text{peak}}$, followed by 15 min @ 60% $\dot{V}\text{O}_{2\text{peak}}$, followed by 3 x 10 s maximal sprints)	Sweat rate ($\text{mL}\cdot\text{min}^{-1}$)
Watson et al. (267)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, physically active, non-heat acclimated males ($n = 12$). Age 22 ± 1 years	Creatine 194 g (9 x $21.6 \text{ g}\cdot\text{d}^{-1}$)	33.5°C 41% RH WBGT 26.4°C	Rectal every 20 min (PCT)	80 min treadmill exercise (4 x 20 min sequences of 4 min resting, alternating 3 min walking, 1 min run x 3 and 4 min walk)	Sweat loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Weiss & Powers (270)	Double-blind, randomized, counter-balanced, independent design	Healthy, aerobically trained males ($n = 24$; 12 vs 12). Age 22.9 ± 3.0 years	Creatine 125 g (5 x $25 \text{ g}\cdot\text{d}^{-1}$)	37°C %RH – not mentioned	Gastrointestinal every 10 min (PCT)	60 min cycling @ 70% age predicted maximum HR	Sweat loss (kg). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)

Wright et al. (278)	Single-blind, intervention	Physically active, heat acclimatized males ($n = 10$). Age 25.7 ± 4.9 years	Creatine 120 g ($6 \times 20 \text{ g} \cdot \text{d}^{-1}$)	35°C 60% RH WBGT 30.4°C	Rectal continuously (PCT)	6 x 10 s maximal cycling sprints	Sweat loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Glycerol							
Anderson et al. (5) ²	Double-blind, randomized, placebo-controlled, cross-over	Endurance trained males ($n = 6$). Age 23.3 ± 6.6 years	Glycerol 1 $\text{g} \cdot \text{kg}^{-1}$ (120 min pre-exercise)	35°C 30% RH WBGT 26°C	Rectal every 15 min (PCT)	90 min cycling @ 98% L_T , followed by 15 min TT	No sweating response data reported
Coutts et al. (41) ¹	Randomized, placebo-controlled, cross-over	Well-trained heat acclimatized male ($n = 7$) and female ($n = 3$) triathletes ($n = 10$). Age 33.3 ± 7.3 years	Glycerol 1.2 $\text{g} \cdot \text{kg}^{-1}$ (130 min pre-exercise)	30.1-45.6°C 46.3-51.7% RH WBGT 23.9-32.9°C	No T_{core} data reported	Olympic distance triathlon	Sweat loss (%)
Desroches et al. (103)	Randomized, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 9$) and females ($n = 1$; $n = 10$). Age 24 ± 4 years	Glycerol 1.4 $\text{g} \cdot \text{kg}^{-1}$ FFM (120, 100, 80 and 60 min pre-exercise)	30°C 50% RH WBGT 24.6°C	Gastrointestinal (no timing mentioned)	5 km treadmill running TT	Sweat loss (mL).
Dini et al. (54) ¹ A and B	Randomized, independent design	High-level oarsmen ($n = 14$; 5 vs 5 vs 4). Age 26 ± 5 years	Glycerol 1 $\text{g} \cdot \text{kg}^{-1}$ (180 min pre-exercise) A Glycerol 1 $\text{g} \cdot \text{kg}^{-1}$ (1 $\text{g} \cdot \text{kg}^{-1}$ 180 min pre-exercise and 23 and 61 min during) B	36°C 30% RH WBGT 26.8°C	Rectal continuously (PCT)	89 min rowing	Fluid loss (mL). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)

Hillman et al. (91)	Randomized, placebo-controlled, cross-over	Healthy, non-heat acclimated trained male cyclists ($n = 7$). Age 28 ± 8 years	Glycerol $1.2 \text{ g}\cdot\text{kg}^{-1}$ (120 min pre-exercise)	35°C 40% RH WBGT 27.6°C	Rectal every 5 min (PCT)	90 min cycling TT	Body mass change (%)
Hitchins et al. (92)	Double-blind, counter-balanced, placebo-controlled, cross-over	Trained, non-heat acclimated male cyclists ($n = 8$). Age 27 ± 4.2 years	Glycerol $1 \text{ g}\cdot\text{kg}^{-1}$ (150 min pre-exercise)	33.2°C 57.8% RH WBGT 28.6°C	Rectal every 5 min (PCT)	60 min cycling (30 min @ fixed power output, followed by 30 min @ self-paced power output)	Sweat loss (%)
Kavouras et al. (107)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, endurance trained male cyclists ($n = 8$). Age 24 ± 3 years	Glycerol $1 \text{ g}\cdot\text{kg}^{-1}$ (30 min pre-exercise)	36.8°C 48.1% RH WBGT 30.5°C	Rectal every 4 min (PCT)	Cycling @ 74% $\dot{V}O_{2\text{peak}}$	Sweating (mL). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Latzka et al. (128) ³	Double-blind, randomized, placebo-controlled, cross-over	Healthy, heat acclimated males ($n = 8$). Age 23 ± 6 years	Glycerol $1.2 \text{ g}\cdot\text{kg}^{-1}$ (no timing mentioned)	34.9°C % RH (not reported) WBGT 30.3°C	Rectal and Esophageal (no timing mentioned)	120 min treadmill exercise @ 45% $\dot{V}O_{2\text{max}}$	Whole-body sweating rate ($\text{g}\cdot\text{m}^{-2}\cdot\text{h}^{-1}$). Local sweating rate of the upper arm ($\text{mg}\cdot\text{min}\cdot\text{cm}^{-1}$; ventilated capsule technique)
Lyons et al. (142)	Randomized, placebo-controlled, cross-over	Healthy, heat acclimatized males ($n = 4$) and females ($n = 2$; $n = 6$). Age 26.2 ± 3.7 years	Glycerol $1 \text{ g}\cdot\text{kg}^{-1}$ (150 min pre-exercise)	42°C 25% RH WBGT 30.6°C	Rectal every 15 min (PCT)	90 min treadmill exercise @ 60% $\dot{V}O_{2\text{max}}$	Sweat output (mL). Converted to WBSR ($\text{mL}\cdot\text{min}^{-1}$)
Marino et al. (146)	Double-blind, randomized, placebo-	Healthy, moderately-to-well trained males ($n = 6$) and females	Glycerol $1.2 \text{ g}\cdot\text{kg}^{-1}$ (150 min pre-exercise)	34.5°C 63.4% RH	Rectal every 5 min (PCT)	60 min cycling TT	Sweat rate ($\text{L}\cdot\text{h}^{-1}$). Converted to

	controlled, cross-over	($n = 1$; $n = 7$). Age 21.2 ± 2.4 years		WBGT 30.5°C			WBSR ($\text{mL} \cdot \text{min}^{-1}$)
McCullagh et al. (150) ¹	Double-blind, randomized, placebo-controlled, cross-over	Healthy, well-trained males ($n = 5$) and females ($n = 1$; $n = 6$). No age provided	Glycerol $1.2 \text{ g} \cdot \text{kg}^{-1}$ (120 min pre-exercise)	30°C % RH (not reported) WBGT 30.3°C	No T_{core} data reported	150 min exercise (10 km treadmill running and 40 km cycling @ a set load [$\sim 177 \text{ W}$] followed by 5 km treadmill running TT)	Body weight loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Scheidler (227)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, non-heat acclimatized, endurance trained males ($n = 6$). Age 27.8 ± 6 years	Glycerol $1.2 \text{ g} \cdot \text{kg}^{-1}$ (140 min pre-exercise)	30°C 50% RH WBGT 24.6°C	Gastrointestinal (no timing mentioned; PCT)	Set distance treadmill running @ $\sim 83\% \dot{V}\text{O}_{2\text{peak}}$	Sweat rate ($\text{L} \cdot \text{h}^{-1}$). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Wingo et al. (277)	Double-blind, randomized, repeated-measures, placebo-controlled, cross-over	Heat acclimatized male mountain bikers ($n = 12$). Age 24.5 ± 3.8 years	Glycerol $1 \text{ g} \cdot \text{kg}^{-1}$ (no timing mentioned)	WBGT 28.1°C	Rectal every 16 km (PCT)	48 km mountain-bicycle race	Sweat rate ($\text{L} \cdot \text{h}^{-1}$). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Sodium citrate							
Nelson et al. (177)	Blinded, randomized, placebo-controlled, cross-over	Healthy, non-heat acclimatized, moderately trained males ($n = 12$). Age 24.3 ± 4.2 years	Sodium citrate $0.2 \text{ g} \cdot \text{kg}^{-1}$ (100 min pre-exercise)	30.9°C 63.8% RH WBGT 27.2°C	Rectal every 5 min (PCT)	62 min cycling @ 15% below V_{T} ($\sim 60\% \dot{V}\text{O}_{2\text{peak}}$)	Sweat loss (L). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Sims et al. (238)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, non-heat acclimatized, endurance trained males ($n = 8$). Age 36 ± 11 years	Sodium citrate 7.72 g AND Sodium chloride 4.5 g (45 min pre-exercise)	32°C 50% RH WBGT 26.5°C	Rectal every 30 s (PCT)	Treadmill running @ $70\% \dot{V}\text{O}_{2\text{max}}$	Sweat loss rate ($\text{L} \cdot \text{h}^{-1}$). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)

Sims et al. (237)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, non-heat acclimatized, endurance trained females ($n = 13$). Age 26 ± 6 years	Sodium citrate 7.72 g AND Sodium chloride 4.5 g (20 min pre-exercise)	32°C 50% RH WBGT 26.5°C	Rectal every 1 min (PCT)	Treadmill running @ 70% $\dot{V}O_{2max}$	Sweat loss rate ($L \cdot h^{-1}$). Converted to WBSR ($mL \cdot min^{-1}$)
Suvi et al. (245)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, endurance trained males ($n = 20$). Age 30.8 ± 5.4 years	Sodium citrate 200 $mg \cdot kg^{-1}$ (3 x 200 $mg \cdot kg^{-1} \cdot d^{-1}$; consumed the day before, the evening before and 120 min pre-exercise)	32°C 46% RH WBGT 25.9°C	Rectal every 1 min (PCT)	40 km cycling TT	Sweat production ($L \cdot h^{-1}$). Converted to WBSR ($mL \cdot min^{-1}$)
Vaher et al. (259)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, endurance trained, non-heat acclimated males ($n = 16$). Age 25.8 ± 4.4 years	Sodium citrate 500 $mg \cdot kg^{-1}$ (120 min pre-exercise)	32°C 50% RH WBGT 26.5°C	Rectal every 1 min (PCT)	5 km treadmill running TT	Body mass loss (kg). Converted to WBSR ($mL \cdot min^{-1}$)
Sodium bicarbonate							
Katagiri et al. (106) ³	Blinded, placebo-controlled, cross-over	Healthy males ($n = 11$). Age 23 ± 4 years	Sodium bicarbonate 300 $mg \cdot kg^{-1}$ (90 min pre-exercise)	35°C 40% RH WBGT 27.6°C	Esophageal every 1 s (PCT)	60 min cycling @ 50% $\dot{V}O_{2peak}$	Body weight loss (kg). Converted to WBSR ($mL \cdot min^{-1}$).
Katagiri et al. (105) ³	Counter-balanced, placebo-controlled, cross-over	Healthy males ($n = 13$). Age 24 ± 2 years	Sodium bicarbonate 300 $mg \cdot kg^{-1}$ (95 min pre-exercise)	35°C 50% RH WBGT 29.1°C	Esophageal every 1 s (PCT)	60 min cycling @ 50% $\dot{V}O_{2peak}$	Body weight loss (%). Converted to WBSR ($mL \cdot min^{-1}$). LSR on the left forearm and chest ($mg \cdot min \cdot cm^{-1}$;

ventilated capsule technique)

Nitrate

Amano et al. (2) ^{2,3}	Double-blind, randomized, placebo-controlled, cross-over	Healthy, active males ($n = 5$) and females ($n = 3$; $n = 8$). Age 24 ± 4 years	Nitrate (NO_3^-) 8 mmol (2 x 8 mmol·d ⁻¹ and 8 mmol 120 min pre-exercise)	30°C 50% RH WBGT 24.6°C	Esophageal continuously (PCT)	30 min cycling @ 55% $\dot{V}\text{O}_{2\text{max}}$	LSR on the left ventral forearm and chest (mg·min·cm ⁻¹ ; ventilated capsule technique)
Cramer et al. (42)	Intervention	Healthy males ($n = 3$) and females ($n = 6$; $n = 9$). Age 67 ± 5 years	Nitrate (NO_3^-) 16.8 mmol (6 x 16.8 mmol·d ⁻¹ and 16.8 mmol 120 min pre-trial)	42.5°C 34.2% RH WBGT 33°C	Gastrointestinal continuously (PCT)	120 min resting in a reclining chair	WBSL (kg). Converted to WBSR (mL·min ⁻¹)
Fowler et al. (64) ³	Double-blind, randomized, placebo-controlled, cross-over	Healthy, physically inactive, non-heat acclimated males ($n = 11$). Age 25 ± 5 years	Nitrate (NO_3^-) 9.2 mmol (5 x 9.2 mmol·d ⁻¹)	35°C 28% RH WBGT 25.7°C	Rectal every 1 min (PCT)	Cycling @ thermoneutral GET at 70 rpm	Body mass change (%). LSR on the chest, forearm, thigh and calf (nL·min ⁻¹ ; ventilated capsule technique)
Kent et al. (115) ²	Double-blind, repeated-measures, counter-balanced, placebo-controlled, cross-over	Endurance-trained male cyclists ($n = 12$). Age 26.6 ± 4.4 years	Nitrate (NO_3^-) 13 mmol (2 x 6.5 mmol·d ⁻¹ and 13 mmol 120 min pre-exercise)	35°C 48% RH WBGT 28.9°C	Gastrointestinal every 20% work rate (PCT)	Total work cycling TT	No sweat response data extractable

Kent et al. (114)	Double-blind, counter-balanced, placebo-controlled, cross-over	Endurance trained male cyclists ($n = 12$). Age 27 ± 6 years.	Nitrate (NO_3^-) 13 mmol (2×6.5 mmol·d ⁻¹ and 13 mmol 120 min pre-trial)	33.3°C 48.8% RH WBGT 27.5°C	Gastrointestinal every 5 min (PCT)	60 min cycling @ 60% $\dot{V}\text{O}_{2\text{peak}}$	Sweat loss (L). Converted to WBSR (mL·min ⁻¹)
Kuennen et al. (125)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 9$). Age 24 ± 1 years	Nitrate (NO_3^-) 4.2 mmol (6×8.4 mmol·d ⁻¹ with 4.2 mmol 2.5 h pre-trial)	41.2°C 15% RH WBGT 27.8°C	Rectal every 5 s (PCT)	45 min treadmill walking @ 4.83 km·h ⁻¹ with a 1.5% incline	Sweat rate (mL·min ⁻¹)
McQuillan et al. (155)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, well-trained endurance male cyclists ($n = 8$). Age 25 ± 8 years	Nitrate (NO_3^-) 8 mmol (2×8 mmol·d ⁻¹ and 8 mmol 90 min pre-exercise)	35°C 60% RH WBGT 30.4°C	Rectal continuously (PCT)	20 min cycling @ 40-60% PPO, followed by 4 km TT	Sweat loss (L·h ⁻¹). Converted to WBSR (mL·min ⁻¹)
Smith et al. (239) ²	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Recreationally trained males ($n = 12$), Age 22 ± 4 years	Nitrate (NO_3^-) 6.2 mmol (180 min pre-exercise)	30°C 70% RH WBGT 26.9°C	Tympanic post IST (PCT)	20 x 6s cycling sprints (114s active recovery)	No sweating response data reported
L-glutamine							
Nava et al. (175) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, physically active, non-heat acclimated males ($n = 7$) and females ($n = 4$; $n = 11$). Age 28.3 ± 6.8 years	L-glutamine 0.15 g·kg ⁻¹ (60 min pre-exercise)	38°C 35% RH WBGT 29.3°C	Rectal (PCT)	87 min simulated fire-fighting exercise	No sweating response data reported
Ogden et al. (183)	Double-blind, randomized, counter-balanced, placebo-	Healthy, recreationally active, non-heat acclimated	L-glutamine 0.3 g·kg ⁻¹ FFM (60 min pre-exercise)	40.3°C 38% RH	Rectal every 10 min (PCT)	30 min treadmill running @ normothermic anaerobic LT	Sweat rate (L·h ⁻¹). Converted to

	controlled, cross-over	males ($n = 10$). Age 29 ± 7 years		WBGT 31.8°C			WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Ogden et al. (184)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated males ($n = 12$). Age 32 ± 6 years	L-glutamine $0.3 \text{ g} \cdot \text{kg}^{-1}$ FFM (60 min pre-exercise)	35.3°C $30.5\% \text{ RH}$ WBGT 26.3°C	Rectal every 20 min (PCT)	2 x 40 min bouts treadmill walking @ $6 \text{ km} \cdot \text{h}^{-1}$ with a 7% incline	WBSL ($\text{L} \cdot \text{h}^{-1}$). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Osborne et al. (186)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, trained, male cyclists ($n = 12$). Age 32 ± 6 years	L-glutamine $0.9 \text{ g} \cdot \text{kg}^{-1}$ FFM (60 min pre-exercise)	35.1°C $51\% \text{ RH}$ WBGT 29.4°C	Rectal every 2 s (Mean T_{core})	20 km cycling TT	Body mass loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Pugh et al. (207) ² A, B and C	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 10$). Age 24 ± 4 years	L-glutamine $0.25 \text{ g} \cdot \text{kg}^{-1}$ FFM (120 min pre-exercise) A L-glutamine $0.5 \text{ g} \cdot \text{kg}^{-1}$ FFM (120 min pre-exercise) B L-glutamine $0.9 \text{ g} \cdot \text{kg}^{-1}$ FFM (120 min pre-exercise) C	30°C $40\text{-}45\% \text{ RH}$ WBGT $23.3\text{-}24^{\circ}\text{C}$	Rectal continuously (Mean T_{core})	60 min treadmill running @ $70\% \dot{V}\text{O}_{2\text{max}}$	No sweating response data reported
Zheng et al. (285)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, untrained males ($n = 13$). Age 20.2 ± 1.1 years	L-glutamine $0.6 \text{ g} \cdot \text{kg}^{-1}$ (30 min pre-exercise)	38°C $60\% \text{ RH}$ WBGT 33.2°C	Gastrointestinal continuously (PCT)	Treadmill running @ $40\% \dot{V}\text{O}_{2\text{max}}$	Body weight loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Zuhl et al. (287) ²	Double-blind, counter-balanced, placebo-	Healthy, endurance trained males ($n = 8$). Age 25 ± 4 years	L-glutamine $0.9 \text{ g} \cdot \text{kg}^{-1}$ FFM (7 x $0.9 \text{ g} \cdot \text{kg}^{-1} \cdot \text{ffm} \cdot \text{d}^{-1}$;	30°C $12\text{-}20\% \text{ RH}$	Rectal (no timing mentioned; PCT)	60 min treadmill running @ $65\text{-}70\% \dot{V}\text{O}_{2\text{max}}$	No sweating response data reported

	controlled, cross-over		120 min pre-exercise)	WBGT 19.2-20.4°C			
Zuhl et al. (286) ²	Double-blind, placebo-controlled, cross-over	Healthy, endurance trained males ($n = 2$) and females ($n = 5$; $n = 7$). Age 26 ± 4 years	L-glutamine 0.9 g·kg ⁻¹ FFM (120 min pre-exercise)	30°C 12-20 % RH WBGT 19.2-20.4°C	Rectal (no timing mentioned; PCT)	60 min treadmill running @ 70% $\dot{V}O_{2max}$	No sweating response data reported
Bovine colostrum							
March et al. (145) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, regularly exercising males ($n = 12$). Age 26 ± 6 years	Bovine colostrum 20 g (14 x 20 g·d ⁻¹)	30°C 60% RH WBGT 25.9°C	Rectal every 10 min (PCT)	60 min treadmill running @ 70% $\dot{V}O_{2max}$ with a 1% incline	No sweating response data reported
McKenna et al. (152)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, active males ($n = 10$). Age 20 ± 2 years	Bovine colostrum 20 g (14 x 20 g·d ⁻¹)	40°C 50% RH WBGT 33.5°C	Gastrointestinal every 5 min (PCT)	~46 min treadmill running @ 95% V_T	Sweat rate (mL·min ⁻¹)
Morrison et al. (170) ² A and B	Double-blind, randomized, placebo-controlled, cross-over	Healthy, trained ($n = 7$; A) and untrained ($n = 8$; B; $n = 15$) males. Age 23 ± 4 vs 21 ± 2 years	Bovine colostrum 1.7 g·kg (7 x 1.7 g·kg ⁻¹ ·d ⁻¹)	30°C 50% RH WBGT 24.6°C	Esophageal every 1 min (PCT)	15 min cycling @ 50% HRR, followed by 30 min treadmill running @ 80% HRR, followed by 30 min TT, followed by 15 min cycling @ 50% HRR	No sweating response data reported
Probiotics							

Gill et al. (76)	Blinded, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, endurance trained, non-heat acclimated male runners ($n = 8$). Age 26 ± 6 years	Probiotic L.casei 100 billion ($7 \times 10^{11} \cdot d^{-1}$)	34°C 32% RH WBGT 25.5°C	Rectal every 10 min (Mean T_{core})	120 min treadmill running @ 60% $\dot{V}O_{2max}$	Body mass loss (%)
Shing et al. (234)	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Healthy, trained, non-heat acclimated, male runners ($n = 10$). Age 27 ± 2 years	Probiotics 28 capsules (28×1 capsule $\cdot kg^{-1} \cdot d^{-1} = 45$ billion colony forming units)	35°C 40% RH WBGT 27.6°C	Gastrointestinal every 1 min (PCT)	Running @ 80% V_T	Body mass loss (kg). Converted to WBSR ($mL \cdot min^{-1}$)
Blackcurrant extract							
Hiles et al. (90)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 12$) and females ($n = 6$; $n = 18$). Age 27 ± 6 years	Blackcurrant extract 600 mg (7×600 mg $\cdot d^{-1}$)	34.1°C 40.8% RH WBGT 27°C	Rectal every 10 min (Mean T_{core})	60 min treadmill running @ 65% $\dot{V}O_{2max}$ with a 1% incline	Whole-body sweat rate ($L \cdot h^{-1}$). Converted to WBSR ($mL \cdot min^{-1}$)
Lee et al. (129)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 12$). Age 28 ± 6 years	Blackcurrant extract 600 mg (7×600 mg $\cdot d^{-1}$)	34.1°C 40.8% RH WBGT 27°C	Rectal every 10 min (PCT)	60 min treadmill running @ 70% $\dot{V}O_{2max}$ with a 1% incline	Sweat rate ($L \cdot h^{-1}$). Converted to WBSR ($mL \cdot min^{-1}$)
Tyrosine							
Coull et al. (40)	Double-blind, counter-balanced, placebo-controlled, cross-over	Recreationally active, non-heat acclimated males ($n = 8$). Age 23 ± 1 years	Tyrosine 150 mg $\cdot kg^{-1}$ (60 min pre-exercise)	40°C 30% RH WBGT 30.1°C	Rectal every 5 min (PCT)	60 min treadmill walk followed by 2.4 km TT wearing a 25 kg backpack	Sweat loss (L). Converted to WBSR ($mL \cdot min^{-1}$)

Kishore et al. (120) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy males ($n = 10$). Age range 20-30 years	Tyrosine 6.5 g (90 min pre-trial)	45°C 30% RH WBGT 34.1°C	Oral temperature (Peak)	90 min resting	No sweating response data reported
Tumilty et al. (255)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, endurance exercising, non-heat acclimated males ($n = 8$). Age 32 ± 11 years	Tyrosine 150 mg·kg ⁻¹ (60 pre-exercise)	30°C 60% RH WBGT 25.9°C	Rectal every 10 min (PCT)	Cycling @ 68% $\dot{V}O_{2peak}$	Body mass loss (kg). Converted to WBSR (mL·min ⁻¹)
Tumilty et al. (254)	Double-blind, randomized, placebo-controlled, cross-over	Endurance exercising, non-heat acclimated males ($n = 7$). Age 20 (range 26) years	Tyrosine 150 mg·kg ⁻¹ (60 pre-exercise)	30°C 60% RH WBGT 25.9°C	Rectal every 5 min (PCT)	60 min cycling @ 57% $\dot{V}O_{2peak}$ followed by total work TT	Body mass loss rate (kg·h ⁻¹). Converted to WBSR (mL·min ⁻¹)
Tumilty et al. (256) A, B and C	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated males ($n = 8$). Age 23 ± 4 years	Tyrosine 150 mg·kg ⁻¹ (timing not mentioned) A Tyrosine 300 mg·kg ⁻¹ (timing not mentioned) B Tyrosine 400 mg·kg ⁻¹ (timing not mentioned) C	30°C 60% RH WBGT 25.9°C	Rectal continuously (PCT)	60 min cycling @ 10% delta of the $\dot{V}O_2$ at GET plus 10% of the difference between GET and $\dot{V}O_{2peak}$, followed by a individualized work target TT	Body mass change (%)
Watson et al. (268)	Randomized, counter-balanced, placebo-controlled, cross-over	Physically active, trained, non-heat acclimated males ($n = 8$). Age 23 ± 3 years	Tyrosine 150 mg·kg ⁻¹ (120 min, 60 min and during)	30°C 50% RH WBGT 24.6°C	Rectal every 5 min (PCT)	Cycling @ 70% $\dot{V}O_{2peak}$	Sweat rate (mL·min ⁻¹)

BCAAs

Cheuvront et al. (33)	Placebo-controlled, cross-over	Healthy, physically active, moderately fit, heat acclimated males ($n = 7$). Age 21 ± 2 years	BCAAs 14 g (0 min pre- and during exercise)	40°C 20% RH WBGT 28°C	Rectal every 10 min (PCT)	60 min cycling @ 50% $\dot{V}O_{2peak}$ followed by 30 min TT	Fluid loss (%)
Macedo et al. (143) ²	Double-blind, randomized, placebo-controlled, cross-over	Physically active males ($n = 9$). Age 25.4 ± 1.2 years	BCAAs 30 mg·kg ⁻¹ (120, 60 and 0 min pre-exercise and every 30 min during exercise)	35°C 60% RH WBGT 30.4°C	Rectal every 30 s (PCT)	Cycling @ 40% peak power at 50 rpm	No sweating response data reported
Mittleman et al. (161)	Double-blind, placebo-controlled, cross-over	Healthy, moderately trained males ($n = 7$) and females ($n = 6$; $n = 13$). Age 24 ± 2.9 vs 25.6 ± 7 years	BCAAs Females (9.4 g) and males (15.8 g; 5 mL·kg ⁻¹ of 5.88 g·L ⁻¹ (every 60 min at rest and 30 min during exercise)	34.4°C 39% RH WBGT 27°C	Esophageal every 5 min (PCT)	Cycling @ 40% $\dot{V}O_{2peak}$	Sweat loss (L). Converted to WBSR (mL·min ⁻¹)
Watson et al. (269) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, endurance exercising, non-heat acclimated males ($n = 8$). Age 28.5 ± 8.2 years	BCAAs 4 x 250 mL at 12 g·L ⁻¹ (30 min intervals pre-exercise and 150 mL every 15 min during exercise)	30°C 38% RH WBGT 23.1°C	Rectal every 10 min (PCT)	Cycling @ 50% $\dot{V}O_{2peak}$	No sweating response data reported
Taurine							
Page et al. (187) ³	Double-blind, randomized, placebo-controlled, cross-over	Healthy, non-heat acclimated males ($n = 11$). Age 23 ± 2 years.	Taurine 50 mg·kg ⁻¹ (120 min pre-exercise)	35°C 40% RH WBGT 27.6°C	Rectal every 1 min (PCT)	Cycling @ thermoneutral V_T at 80 rpm	Body mass change (g). Converted to WBSR (mL·min ⁻¹). LSR on the chest, upper-arm

							thigh and calf ($\text{nL} \cdot \text{min}^{-1}$; ventilated capsule technique)
Peel et al. (192) ³	Double-blind, randomized, placebo- controlled, cross- over	Healthy, active, non- heat acclimated males ($n = 12$) and females ($n = 3$; $n =$ 15). Age 27 ± 5 years	Taurine $50 \text{ mg} \cdot \text{kg}^{-1}$ (~60 min pre-exercise)	$37.5 \text{ }^\circ\text{C}$ $34.2\% \text{ RH}$ WBGT $28.9 \text{ }^\circ\text{C}$	Rectal every 5s (PCT)	45 min treadmill walking @ 200 $\text{W}/\text{m}^2 \dot{H}_{\text{prod}}$	WBSL (g; calculated from pre-post BM loss) LSR upper back (absorbent patch technique)
GABA							
Miyazawa et al. (163)	Randomized, placebo- controlled, cross- over	Healthy, exercise trained males ($n = 8$). Age 22.8 ± 3.7 years	GABA 1 g (20 min pre-trial)	35°C $50\% \text{ RH}$ WBGT 29.1°C	Esophageal (no timing mentioned; PCT)	30 min semi- recumbent cycling @ $65\% \dot{V}\text{O}_{2\text{peak}}$ at 60 rpm	Sweat loss (g). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Miyazawa et al. (162) ³	Double-blind, randomized, placebo- controlled, cross- over	Healthy, moderately active males ($n = 8$). Age 23.5 ± 3.6 years	GABA 1 g (0 min pre-trial)	33°C $50\% \text{ RH}$ WBGT 27.3°C	Esophageal (no timing mentioned; PCT)	30 min resting	Sweat loss (g). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$). LSR on the chest ($\text{mg} \cdot \text{min} \cdot \text{cm}^{-1}$; ventilated capsule technique)
Betaine							
Armstrong et al. (6) ²	Double-blind, randomized, placebo- controlled, cross- over	Healthy, well-trained male runners ($n =$ 10). Age 20 ± 2 years.	Betaine 5 g (45 min pre- exercise)	31.1°C $34.7\% \text{ RH}$ WBGT 23.6°C	Rectal periodically (PCT)	75 min treadmill running @ 65% $\dot{V}\text{O}_{2\text{max}}$ followed by TTE @ $84\% \dot{V}\text{O}_{2\text{max}}$	WBSR ($\text{L} \cdot \text{h}^{-1}$). No sweating data extractable

Willingham et al. (276)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, recreationally active males ($n = 11$). Age 29.1 ± 5.2 years	Betaine $50 \text{ mg} \cdot \text{kg}^{-1}$ ($7 \times 50 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$)	40°C 60% RH WBGT 35.1°C	Gastrointestinal every 15 min (PCT)	60 min resting	Sweat rate ($\text{L} \cdot \text{h}^{-1}$). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
L-arginine							
Tyler et al. (258)	Double-blind, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated males ($n = 8$). Age 27 ± 6 years	L-arginine 10 g (30 min pre-trial)	35°C 50% RH WBGT 29.1°C	Rectal every 5 min (PCT)	90 min resting, followed by 30 min cycling @ 60% W_{max} , followed by 30 min resting	Body mass loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Ascorbic acid							
Carrillo et al. (27) ²	Double-blind, randomized, independent design	Healthy, aerobically fit males ($n = 8$) and females ($n = 4$; $n = 12$; 6 vs 6). Age 23.4 ± 4.6 years.	Ascorbic acid (vitamin C) 1500 mg ($9 \times 1500 \text{ mg} \cdot \text{d}^{-1}$ and 8 mmol 120 min pre-exercise)	34.8°C 13% RH WBGT 22.8°C	Rectal pre and post exercise (PCT)	180 min cycling @ 55% $\dot{V}O_{2\text{max}}$	No sweating response data extractable
Kotze et al. (123) A and B	Placebo-controlled, independent design	Non-heat acclimated, males ($n = 13$; 4 vs 5 vs 4). Age 23 ± 3 vs 24 ± 2 vs 20 ± 2.9 years	Ascorbic acid (vitamin C) 250 mg (180-240 min pre-exercise) A Ascorbic acid (vitamin C) 500 mg (180-240 min pre-exercise) B	33.9°C %RH (did not mention)	Rectal every 60 min (PCT)	240 min block stepping @ 35 W workload	Sweat output (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Vitamin E							

Keong et al. (116) ²	Double-blind, randomized, placebo-controlled, cross-over	Recreational, heat acclimated male athletes ($n = 18$). Age 24.9 ± 1.4 years	Vitamin E No dose stated (6 weeks)	31°C 70% RH WBGT 27.9°C	Rectal every 10 min (PCT)	Treadmill running @ 70% $\dot{V}O_{2max}$	No sweating response data reported
Eurycoma longifolia Jack							
Muhamad et al. (171)	Double-blind, randomized, placebo-controlled, cross-over	Healthy, male recreational athletes ($n = 12$). Age 23.3 ± 3.7 years	Eurycoma longifolia Jack 150 mg (7 x 150 mg·d ⁻¹ and 150 mg 60 min pre-exercise)	31°C 70% RH WBGT 27.9°C	Tympanic every 10 min (PCT)	60 min treadmill running @ 60% $\dot{V}O_{2max}$ followed by 20 min TT	Sweat rate (L·h ⁻¹). Converted to SR (mL·min ⁻¹)
Oligonol							
Lee & Shin (132) ^{2,3}	Placebo-controlled, cross-over	Healthy males ($n = 19$). Age 23.7 ± 2.3 years	Oligonol 200 mg (7 x 200 mg)	26°C 60% RH WBGT 22.2°C 42°C bath	Tympanic (no timing mentioned; PCT)	30 min half body water immersion	LSR on the chest, back, abdomen and thigh (mg·min·cm ⁻¹ ; ventilated capsule technique)
Lee et al. (131) ¹	Placebo-controlled, cross-over	Healthy males ($n = 19$). Age 23.7 ± 2.3 years	Oligonol 200 mg (7 x 200 mg)	26°C 60% RH WBGT 22.2°C 42°C bath	Tympanic (no timing mentioned; PCT)	30 min half body water immersion	WBSL volume (mL·30 min). Converted to WBSR (mL·min ⁻¹)
Shin et al. (232) ²	Randomized, placebo-controlled, cross-over	Healthy males ($n = 13$). Age 21.8 ± 2.3 years.	Oligonol 100 mg (30 min pre-trial)	26°C 60% RH	Tympanic (no timing mentioned; PCT)	30 min lower leg water immersion	No sweating response data reported

				WBGT 22.2°C			
				43°C bath			
Shin et al. (233) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy males ($n = 17$). Age 21.6 ± 2.1 years	Oligonol 100 mg (60 min pre-trial)	26°C 60% RH WBGT 22.2°C 42°C bath	Tympanic (no timing mentioned; PCT)	30 min half body water immersion	No sweating response data reported
Polyphenols							
Trinity et al. (253) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, well-trained male cyclists ($n = 12$). Age 26.8 ± 5 years	Polyphenols 3600-ppm (7 x 3600-ppm·d ⁻¹)	31.5°C 55% RH WBGT 26.7°C	Rectal continuously (PCT)	20 min cycling @ 40, 50, 60 and 70% followed by 30 min cycling @ 5% above L _T , followed by 10 min TT @ 90% $\dot{V}O_{2max}$	Body mass loss (kg). Converted to WBSR (mL·min ⁻¹)
Curcumin							
Szymanski et al. (249)	Double-blind, counter-balanced, placebo-controlled, cross-over	Healthy, recreationally active, non-heat acclimated males ($n = 6$) and females ($n = 2$; $n = 8$). Age 19 ± 1 years	Curcumin 300 mg (3 x 500 mg·d ⁻¹ , with 300 mg 60 min pre-exercise)	37°C 25% RH WBGT 26.7°C	Esophageal every 5 min (PCT)	60 min treadmill running @ 65% $\dot{V}O_{2max}$	Sweat rate (L·h ⁻¹). Converted to WBSR (mL·min ⁻¹)
Quercetin							
Kuennen et al. (124)	Double-blind, counter-balanced, placebo-controlled, cross-over	Healthy, non-heat acclimated, physically active males ($n = 8$). Age 28 ± 4.8 years	Quercetin 2000 mg·d ⁻¹ (with breakfast)	46.6°C 21% RH WBGT 33.3°C	Rectal continuously (PCT)	45 min treadmill running @ 50% $\dot{V}O_{2max}$	WBSR (mL·min ⁻¹)

Menthol							
Bray et al. (24)	Double-blind, randomized, cross-over	Healthy, moderately trained, non-heat acclimated males ($n = 5$) and females ($n = 5$; $n = 10$). Age 23 ± 5 years	Menthol drink 85 mL (0.01% menthol; 0 min pre-exercise, every 10 min during and 1 min pre-TT)	35°C 54% RH WBGT 29.7°C	Rectal every 20 s (PCT)	40 min @ 50% $\dot{V}O_{2max}$, followed by 15 min TT	WBSL (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Riera et al. (211) ²	Randomized, placebo-controlled, cross-over	Healthy, heat acclimated, trained male cyclists and triathletes ($n = 12$). Age 42 ± 13 years	Menthol aroma 190 mL (0.5 g/L; 0.01% menthol; 15 and 0 min pre-trial and every 5 km)	30.7°C 78% RH WBGT 28.6°C	Gastrointestinal pre, post and every 5 km (PCT)	20 km cycling TT	No sweating response data reported
Vogel et al. (262) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy, non-heat acclimated, endurance trained male ($n = 8$) and female ($n = 6$) runners ($n = 14$). Age 31 ± 6 years	Menthol gel 16 g (0.5% menthol; 5 min pre-exercise and 50 and 40 min mid-trial)	33°C 49% RH WBGT 27.2°C	Gastrointestinal every 10 min (PCT)	40 min treadmill running @ 60% HR_{max} at $\dot{V}O_{2max}$, followed by 20 min TT with a 1% incline	Body mass loss (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$)
Folic acid							
Gagnon et al. (73) ³	Intervention	Healthy males ($n = 3$) and females ($n = 6$; $n = 9$). Age 68 ± 3 years	Folic acid 5 mg ($42 \times 5 \text{ mg} \cdot \text{d}^{-1}$)	42°C 30-70% RH WBGT 31.7-38.3°C	Esophageal continuously (PCT)	100 min resting	WBSL (kg). Converted to WBSR ($\text{mL} \cdot \text{min}^{-1}$). LSR on the forearm ($\text{mg} \cdot \text{min} \cdot \text{cm}^{-1}$; ventilated capsule technique)

Beta-glucan							
Zabriskie et al. (282)	Double-blind, randomized, counter-balanced, placebo-controlled. cross-over	Healthy, recreationally active males ($n = 16$) and females ($n = 15$; $n = 31$). Age 29.6 ± 6.7 vs 30.1 ± 8.9 years	Beta-glucan 250 mg (11 x 250 mg·d ⁻¹)	37.2°C 45.2% RH WBGT 30.3°C	Gastrointestinal every 10 min (PCT)	60 min treadmill walking @ 55% $\dot{V}O_{2peak}$	Body weight loss (kg). Converted to WBSR (mL·min ⁻¹)
Ginseng							
Ping et al. (200) ²	Double-blind, randomized, placebo-controlled, cross-over	Recreational, heat acclimated male runners ($n = 9$). Age 25.4 ± 6.9 years	Ginseng 200 mg (60 min pre-exercise)	31°C 70% RH WBGT 27.9°C	Rectal every 10 min (PCT)	Treadmill running @ 70% $\dot{V}O_{2max}$	No sweating response data reported
Catechin							
Nishimura et al. (180) ^{2,3}	Placebo-controlled, cross-over	Healthy males ($n = 8$). Age 26 ± 8 years	Catechin 121 mg/100 mL (4 mL·kg ⁻¹ 3 x (0, 30 and 60 min during)	35°C 75% RH WBGT 32.3°C 40°C bath	Tympanic (no timing mentioned; PCT)	90 min lower leg water immersion	LSR on the upper arm (mg·min·cm ⁻¹ ; ventilated capsule technique)
Effective microorganism X							
Taylor et al. (251) ²	Double-blind, randomized, cross-over	Males ($n = 6$). Age 22.0 ± 1.3 years	Effective microorganism X 70 mL (7 x 70 mL·d ⁻¹)	34.7°C 51.7% RH WBGT 29°C	Rectal every 5 min (PCT)	20 x 10 s IST @ maximal running velocity, with 80 s active recover @ 35% $\dot{V}O_{2max}$	No sweating response data reported
α-KG and 5-HMF							

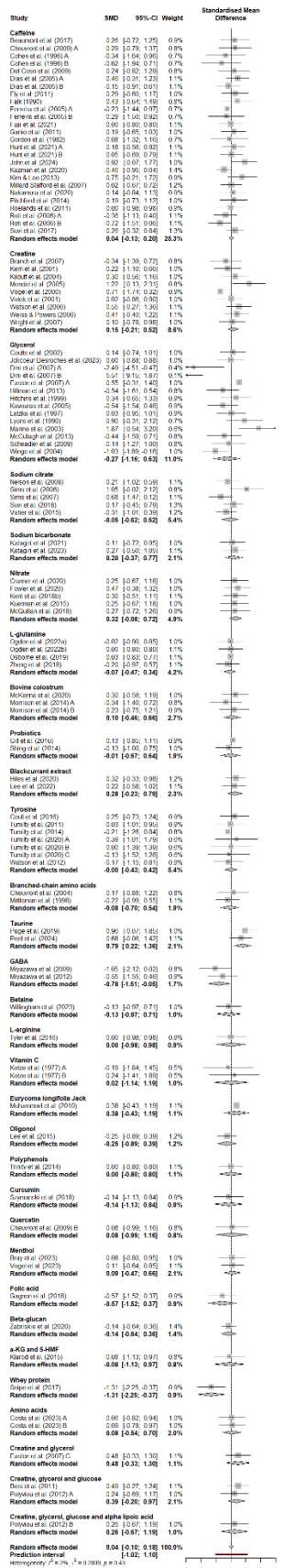
Klarod et al. (121)	Randomized, placebo-controlled, cross-over	Healthy, regularly active males ($n = 7$). Age 2.7 ± 2.6 years	α -KG 4.8 g AND 5-HMF 60 mg (48 h pre-trial)	33°C 40% RH WBGT 26°C	Tympanic pre and post exercise (PCT)	Treadmill running @ $1 \text{ km} \cdot \text{h}^{-1} \cdot \text{min}^{-1}$ increases	No sweating response data reported
Thermo Speed Extreme							
Pokora et al. (204) ²	Double-blind, randomized, placebo-controlled, cross-over	Healthy males ($n = 12$) and females ($n = 13$; $n = 25$). Age 23 ± 1.3 years	Thermo Speed Extreme (green tea extract $5.1 \text{ mg} \cdot \text{kg}^{-1}$, synephrine $0.3 \text{ mg} \cdot \text{kg}^{-1}$ and caffeine $3 \text{ mg} \cdot \text{kg}^{-1}$; 0 min pre-trial)	26°C 56% RH WBGT 21.8°C	Tympanic periodically (PCT)	6 h resting	No sweating response data reported
Whey protein							
Snipe et al. (240)	Randomized, cross-over	Healthy non heat acclimatized, endurance trained male ($n = 6$) and female ($n = 5$) runners ($n = 11$). Age 31 ± 5 years	Whey protein hydrolysate 15 g (0 min pre-exercise and every 20 min during)	35.5°C 27% RH WBGT 25.9°C	Rectal every 10 mins (PCT)	120 min treadmill running @ 60% $\dot{V}O_{2\text{max}}$	Body mass loss (%)
Amino acids							
Costa et al. (39) A and B	Double-blind, randomized, counter-balanced, placebo-controlled, cross-over	Non-heat acclimated, endurance running trained males ($n = 20$; 10 vs 10). Age 32 ± 8 years	Amino acid beverage VS001 $474 \text{ mL} \cdot \text{d}^{-1}$ (7 x 474 $\text{mL} \cdot \text{d}^{-1}$; 237 mL 0 min pre-exercise and every 20 min during) A	34.6°C 38% RH WBGT 27.1°C	Rectal (no timing mentioned; PCT)	120 min treadmill running @ 60% $\dot{V}O_{2\text{max}}$	Body mass loss (%)

			Amino acid beverage VS006 474 mL·d ⁻¹ (7 x 474 mL·d ⁻¹ ; 237 mL 0 min pre-exercise and every 20 min during) B				
Mixed supplements							
Bandyopadhyay et al. (10) ²	Double-blind, randomized, placebo-controlled, cross-over	Recreational, heat acclimated male runners (<i>n</i> = 9). Age 25.4 ± 6.9 years	Caffeine 5 mg·kg ⁻¹ AND Ginseng 200 mg (60 min pre-exercise)	31°C 70% RH WBGT 27.9°C	Rectal every 10 min (PCT)	Treadmill running @ 70% $\dot{V}O_{2max}$	No sweating response data reported
Yu et al. (281) ² A, B and C	Single-blind, randomized, placebo-controlled, cross-over	Healthy, non caffeine-habituated students (<i>n</i> = 12). Age 23.8 ± 2.4 years	Caffeine 5 mg·kg ⁻¹ (60 min pre-exercise) A Taurine 50 mg·kg ⁻¹ (60 min pre-exercise) B Taurine 5 mg·kg ⁻¹ AND Caffeine 50 mg·kg ⁻¹ (60 min pre-exercise) C	35°C 65% RH WBGT 31.1°C	Tympanic every 3 min (PCT)	Cycling @ thermoneutral V_T at 80 rpm	No sweating response data reported
Easton et al. (55) A, B ² and C	Double-blind, randomized, intervention	Healthy, endurance trained males (<i>n</i> = 23; 12 [creatine and glycerol; creatine] vs 11 [glycerol]). Age 31 ± 7 years	Glycerol 1 g·kg ⁻¹ (6 x 1 g·d ⁻¹ and 1 g 5 h pre-trial) A Creatine	30°C 70% RH WBGT 26.9°C	Rectal every 5 min (PCT)	40 min cycling @ 63% WR_{max} followed by 16.1 km TT	Sweat rate (L·h ⁻¹). Converted to WBSR (mL·min ⁻¹)

			120 g (6 x 20 g·d ⁻¹ and 10 g 5 h pre-trial) B				
			Creatine 120 g (6 x 20 g·d ⁻¹ and 10 g 5 h pre-trial) AND Glycerol 1 g·kg ⁻¹ (6 x 1 g·d ⁻¹ and 1 g 5 h pre-trial) C				
Beis et al. (14)	Intervention	Healthy males (<i>n</i> = 14). Age 27 ± 8 years	Creatine 10 g, Glycerol 1 g·kg ⁻¹ AND Glucose 75 g (6 x creatine 10 g·d ⁻¹ , glycerol 1 g·kg ⁻¹ and glucose 75 g·d ⁻¹ and 5 h pre-exercise)	35.1°C 69.4% RH WBGT 31.8°C	Gastrointestinal every 5 min (PCT)	30 min treadmill running @ 60% $\dot{V}O_{2max}$, with a 1% incline	Sweat loss (L). Converted to WBSR (mL·min ⁻¹)
Polyviou et al. (205) A and B	Double-blind, randomized, intervention	Healthy, endurance trained males (<i>n</i> = 18; 9 vs 9). Age 31.5 ± 9 years	Creatine 20 g, Glycerol 2 mg AND Glucose 150 g (7 x creatine 20 g·d ⁻¹ , glycerol 2 mg·kg ⁻¹ ·d ⁻¹ and glucose 150 g·d ⁻¹) A Creatine 20 g, Glycerol 2 mg, Glucose	30°C 70% RH WBGT 26.9°C	No method mentioned, measured every 5 min (PCT)	40 min cycling @ pre-determined work rate, followed by 16.1 km TT	Sweat loss (mL). Converted to WBSR (mL·min ⁻¹)

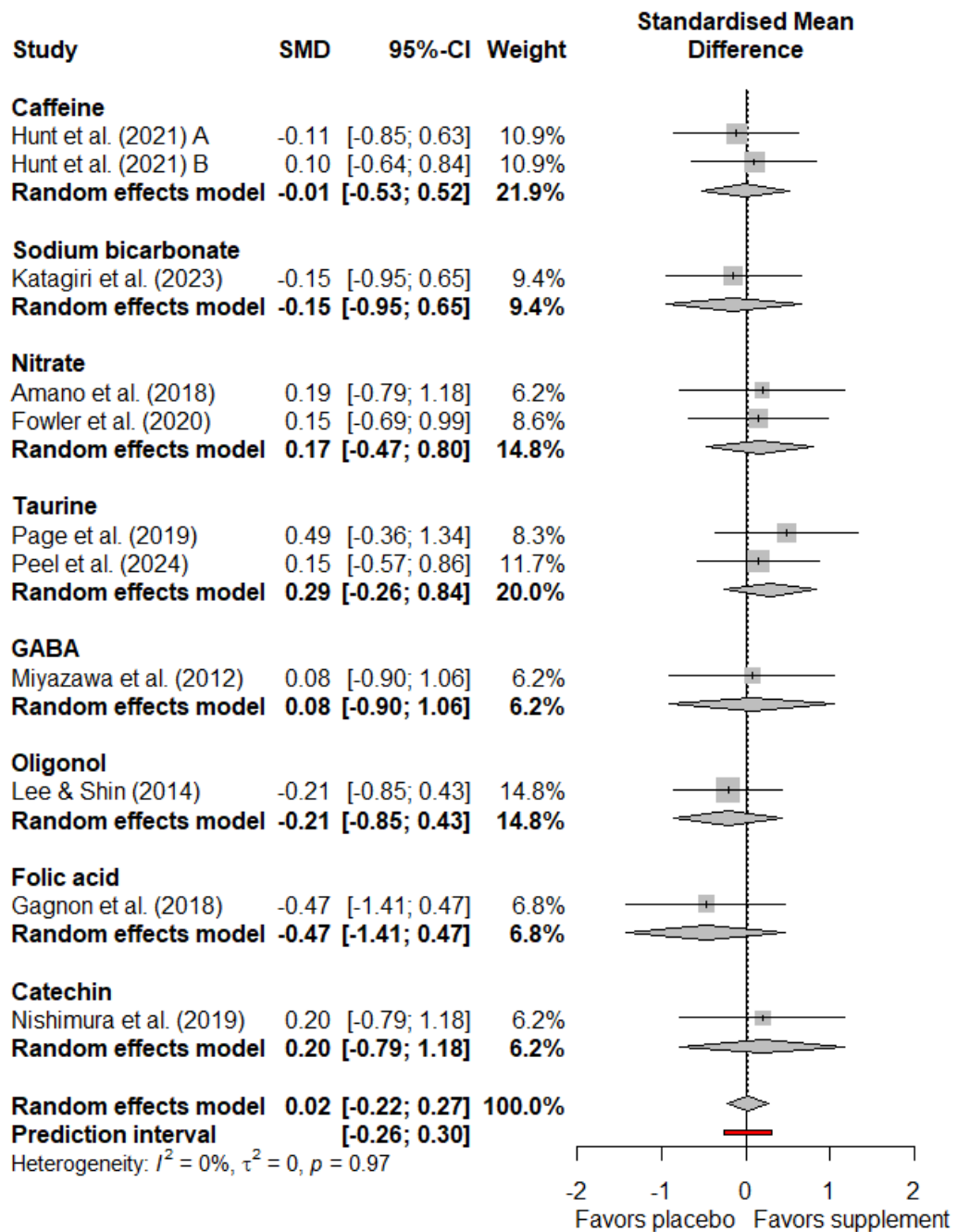
100 g AND
Alpha lipoic acid
1000 mg (7 x
creatine 20 g·d⁻¹,
glycerol 2
mg·kg⁻¹·d⁻¹, glucose
100 g·d⁻¹ and alpha
lipoic acid 1000 mg)
B

TT time-trial, *TTE* time-to-exhaustion, *IST* intermittent-sprint-test, *PPO* peak power output, *PCT* peak core temperature, *T_{core}* core temperature, *RH* relative humidity, *WBGT* wet-bulb globe temperature, *VO_{2max}* maximal oxygen uptake, *VO_{2peak}* peak oxygen uptake, *W_{max}* watt maximum, *WR_{max}* work rate maximum, *H_{prod}* heat production, *PPO* peak power output, *HR* heart rate, *L_T* lactate threshold, *GET* gas exchange threshold, *V_T* ventilatory threshold, *rpm* revolution per minute, *WBSR* whole-body sweat rate, *WBSL* whole-body sweat loss, *LSR* local sweat rate, *FFM* fat free mass, *BCAAs* branched-chain amino acids, *GABA* gamma-aminobutyric acid, *α-KG* alpha-ketoglutaric acid, 5-HMF 5-hydroxymethylfurfural, ¹ not included in peak core temperature analysis, ² not included in the sweating response analysis, ³ included in local sweat rate analysis. Data are reported as mean ± SD. The table is a reflection of the studies, as reported by the authors of the original article.



4

5 **Figure 3.** Effect of dietary supplementation on whole-body sweat rate.



6

7 **Figure 4.** Effect of dietary supplementation on local sweat rate.

8

9 **3.4 Sub-group analysis**

10 **3.4.1 Peak core temperature**

11 Sub-group analysis demonstrated a significant overall pooled effect of the different dietary supplement categories
12 on peak T_{core} ($p = 0.015$). However, the following supplements all demonstrated non-significant *trivial* effects on
13 peak T_{core} : creatine (Hedges' $g = -0.12$, 95% CI -0.453 to 0.203 , $p = 0.456$), nitrate (Hedges' $g = 0.07$, 95% CI $-$
14 0.237 to 0.381 , $p = 0.648$), L-glutamine (Hedges' $g = 0.07$, 95% CI -0.246 to 0.384 , $p = 0.667$), bovine colostrum
15 (Hedges' $g = 0.13$, 95% CI -0.327 to 0.588 , $p = 0.575$), probiotics (Hedges' $g = 0.00$, 95% CI -0.653 to 0.653 , p
16 $= 1.000$), blackcurrant extract (Hedges' $g = -0.11$, 95% CI -0.461 to 0.453 , $p = 0.986$), tyrosine (Hedges' $g = -$
17 0.04 , 95% CI -0.427 to 0.345 , $p = 0.835$), BCAAs (Hedges' $g = -0.004$, 95% CI -0.327 to 0.588 , $p = 0.575$),
18 betaine (Hedges' $g = -0.14$, 95% CI -0.749 to 0.464 , $p = 0.646$), vitamin C (Hedges' $g = -0.07$, 95% CI -0.891 to
19 0.743 , $p = 0.859$), Eurycoma longifolia Jack (Hedges' $g = 0.00$, 95% CI -0.800 to 0.800 , $p = 1.000$), polyphenols
20 (Hedges' $g = 0.00$, 95% CI -0.800 to 0.800 , $p = 1.000$), folic acid (Hedges' $g = 0.00$, 95% CI -0.924 to 0.924 , $p =$
21 1.000), amino acids (Hedges' $g = 0.13$, 95% CI -0.492 to 0.753 , $p = 0.681$), combined caffeine and taurine
22 (Hedges' $g = 0.07$, 95% CI -1.063 to 1.200 , $p = 0.906$) and combined creatine, glycerol, glucose and alpha lipoic
23 acid (Hedges' $g = 0.00$, 95% CI -0.924 to 0.924 , $p = 1.000$).

24 There were a number of caffeine-based supplements that increased T_{core} , with isolated caffeine (Hedges' $g = 0.44$,
25 95% CI 0.275 to 0.603 , $p < 0.001$) having a *small* significant positive effect and combined caffeine and ginseng
26 demonstrating a *large* significant positive effect (Hedges' $g = 1.19$, 95% CI 0.163 to 2.208 , $p = 0.023$). L-arginine
27 (Hedges' $g = 0.22$, 95% CI -0.765 to 1.203 , $p = 0.663$), sodium bicarbonate (Hedges' $g = 0.26$, 95% CI -0.309 to
28 0.829 , $p = 0.370$), beta-glucan (Hedges' $g = 0.28$, 95% CI -0.217 to 0.784 , $p = 0.268$), ginseng (Hedges' $g = 0.38$,
29 95% CI -0.554 to 1.316 , $p = 0.424$) and combined alpha-ketoglutaric acid (α -KG) and 5-hydroxymethylfurfural
30 (5-HMF; Hedges' $g = 0.35$, 95% CI -0.709 to 1.408 , $p = 0.518$) had *small* non-significant positive effects. Glycerol
31 (Hedges' $g = -0.28$, 95% CI -0.700 to 0.130 , $p = 0.179$), sodium citrate (Hedges' $g = -0.46$, 95% CI -1.261 to
32 0.341 , $p = 0.261$), GABA (Hedges' $g = -0.46$, 95% CI -1.401 to 0.480 , $p = 0.337$), vitamin E (Hedges' $g = -0.23$,
33 95% CI -0.889 to 0.423 , $p = 0.487$), curcumin (Hedges' $g = -0.28$, 95% CI -1.268 to 0.704 , $p = 0.575$), quercetin
34 (Hedges' $g = -0.24$, 95% CI -0.973 to 0.496 , $p = 0.524$), menthol (Hedges' $g = -0.46$, 95% CI -1.299 to 0.380 , p
35 $= 0.283$), Thermo Speed Extreme (Hedges' $g = -0.23$, 95% CI -0.785 to 0.327 , $p = 0.420$), Effective
36 microorganism X (Hedges' $g = -0.30$, 95% CI -1.441 to 0.841 , $p = 0.606$), whey protein (Hedges' $g = -0.36$, 95%
37 CI -1.201 to 0.486 , $p = 0.407$) and combined creatine and glycerol (Hedges' $g = -0.47$, 95% CI -1.460 to 0.530 , p
38 $= 0.359$) had *small* non-significant negative effects.

39 There were some medium-to-large effects of supplementation on peak T_{core} , such as oligonol (Hedges' $g = -0.50$,
40 95% CI -0.907 to -0.101 , $p = 0.014$) and taurine (Hedges' $g = -0.66$, 95% CI -1.296 to 0.022 , $p = 0.043$), which

41 had a *medium* significant negative effect, and combined creatine, glycerol and glucose (Hedges' $g = -0.66$, 95%
42 CI -2.187 to 0.873 , $p = 0.400$) had *medium* non-significant effects. Catechin (Hedges' $g = -0.80$, 95% CI -1.825
43 to 0.235 , $p = 0.130$) had *large* non-significant negative effects on peak T_{core} .

44

45 3.4.2 Whole-body sweat rate

46 Sub-group analysis demonstrated a non-significant overall pooled effect of the different supplement categories on
47 WBSR ($p = 0.434$). Caffeine (Hedges' $g = 0.04$, 95% CI -0.129 to 0.203 , $p = 0.660$), creatine (Hedges' $g = 0.15$,
48 95% CI -0.208 to 0.517 , $p = 0.403$), sodium citrate (Hedges' $g = -0.05$, 95% CI -0.616 to 0.525 , $p = 0.875$), L-
49 glutamine (Hedges' $g = -0.07$, 95% CI -0.473 to 0.337 , $p = 0.742$), bovine colostrum (Hedges' $g = 0.10$, 95% CI
50 -0.457 to 0.659 , $p = 0.723$), probiotics (Hedges' $g = -0.01$, 95% CI -0.666 to 0.642 , $p = 0.972$), tyrosine (Hedges'
51 $g = -0.001$, 95% CI -0.426 to 0.423 , $p = 0.995$), BCAAs (Hedges' $g = -0.08$, 95% CI -0.703 to 0.540 , $p = 0.797$),
52 betaine (Hedges' $g = -0.13$, 95% CI -0.967 to 0.706 , $p = 0.760$), L-arginine (Hedges' $g = 0.00$, 95% CI -0.980 to
53 0.980 , $p = 1.000$), vitamin C (Hedges' $g = 0.02$, 95% CI -1.141 to 1.191 , $p = 0.967$), polyphenols (Hedges' $g =$
54 0.00 , 95% CI -0.800 to 0.800 , $p = 1.000$), curcumin (Hedges' $g = -0.14$, 95% CI -1.126 to 0.838 , $p = 0.774$),
55 quercetin (Hedges' $g = 0.08$, 95% CI -0.992 to 1.156 , $p = 0.881$), menthol (Hedges' $g = 0.09$, 95% CI -0.473 to
56 0.660 , $p = 0.746$), beta-glucan (Hedges' $g = -0.14$, 95% CI -0.635 to 0.362 , $p = 0.591$), amino acids (Hedges' $g =$
57 0.08 , 95% CI -0.544 to 0.965 , $p = 0.877$) and combined α -KG and 5-HMF (Hedges' $g = -0.08$, 95% CI -1.131 to
58 0.965 , $p = 0.877$) all had *trivial* non-significant effects.

59 For WBSR, nitrate (Hedges' $g = 0.32$, 95% CI -0.083 to 0.716 , $p = 0.120$), blackcurrant extract (Hedges' $g = 0.28$,
60 95% CI -0.227 to 0.791 , $p = 0.277$), Eurycoma longifolia Jack (Hedges' $g = 0.38$, 95% CI -0.429 to 1.188 , $p =$
61 0.358), sodium bicarbonate (Hedges' $g = 0.20$, 95% CI -0.368 to 0.768 , $p = 0.490$), combined creatine and glycerol
62 (Hedges' $g = 0.48$, 95% CI -0.332 to 1.296 , $p = 0.246$), combined creatine, glycerol and glucose (Hedges' $g =$
63 0.39 , 95% CI -0.195 - 0.975 , $p = 0.192$) and combined creatine, glycerol, glucose and alpha lipoic acid (Hedges' $g =$
64 0.26 , 95% CI -0.671 to 1.187 , $p = 0.586$) had *small* non-significant positive effects. Glycerol (Hedges' $g = -$
65 0.27 , 95% CI -1.163 to 0.632 , $p = 0.562$) and oligonol (Hedges' $g = -0.25$, 95% CI -0.886 to 0.392 , $p = 0.448$)
66 had a *small* non-significant negative effect.

67 There were a number of medium-to-large effects on WBSR, including GABA (Hedges' $g = -0.78$, 95% CI -1.514
68 to -0.053 , $p = 0.036$), which had a *medium* significant negative effect and folic acid (Hedges' $g = -0.57$, 95% CI $-$
69 1.523 to 0.373 , $p = 0.235$), which had a *medium* non-significant negative effect. Taurine (Hedges' $g = 0.79$, 95%

70 CI 0.225 to 1.363, $p = 0.006$) had a *medium* significant positive effect and whey protein (Hedges' $g = -1.31$, 95%
71 CI -2.248 to -0.371, $p = 0.006$) had a *large* non-significant negative effect.

72

73 3.4.3 Local sweat rate

74 Sub-group analysis demonstrated a non-significant effect of the different supplement categories on LSR ($p =$
75 0.886). Caffeine (Hedges' $g = -0.005$, 95% CI -0.529 to 0.519, $p = 0.985$), GABA (Hedges' $g = 0.08$, 95% CI -
76 0.905 to 1.056, $p = 0.880$), nitrate (Hedges' $g = 0.17$, 95% CI -0.471 to 0.804, $p = 0.608$) and sodium bicarbonate
77 (Hedges' $g = -0.15$, 95% CI -0.950 to 0.653, $p = 0.717$) had *trivial* non-significant effects. Catechin (Hedges' $g =$
78 0.20, 95% CI -0.787 to 1.180, $p = 0.695$), and taurine (Hedges' $g = 0.29$, 95% CI -0.261 to 0.835, $p = 0.305$) had
79 *medium* non-significant positive effects. Oligonol (Hedges' $g = -0.21$, 95% CI -0.849 to 0.427, $p = 0.517$) and
80 folic acid (Hedges' $g = -0.47$, 95% CI -1.414 to 0.467, $p = 0.323$) also had *medium* non-significant effects but
81 decreased the local sweating response.

82

83 3.5 Meta-regression

84 Across the three meta-analyses, there were no significant moderating effects. The effect of several moderating
85 variables on WBSR and LSR could not be assessed due to either an insufficient number of studies included in the
86 analysis (supplement dose) or a lack of variation within the moderating variables in the included studies (e.g.
87 training, heat acclimation and hydration status).

88 **Table 2.** Meta-regression of potential moderating variables of the peak core temperature, whole-body sweat rate and local sweat rate meta-analyses.

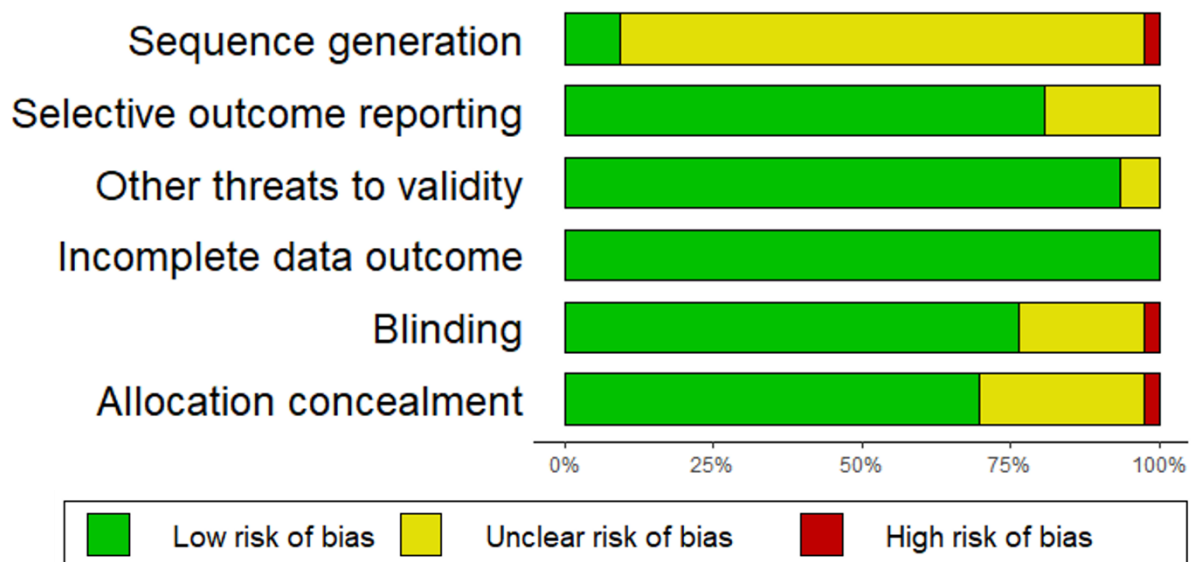
Moderator	Peak core temperature	Whole-body sweat rate	Local sweat rate
Training status	$\beta = -0.019, p = 0.854 (n = 115)$	$\beta = -0.129, p = 0.416 (n = 89)$	—
Heat acclimation status	$\beta = 0.092, p = 0.617 (n = 71)$	$\beta = -0.440, p = 0.090 (n = 65)$	—
Hydration status	$\beta = -0.190, p = 0.764 (n = 105)$	$\beta = -0.126, p = 0.876 (n = 84)$	—
Fluid ingestion during the trial	$\beta = -0.018, p = 0.889 (n = 89)$	$\beta = -0.152, p = 0.416 (n = 73)$	$\beta = 0.078, p = 0.835 (n = 6)$
Wet-bulb globe temperature	$\beta = -0.008, p = 0.596 (n = 134)$	$\beta = -0.011, p = 0.655 (n = 105)$	$\beta = -0.003, p = 0.929 (n = 11)$
Trial type	$\beta = -0.249, p = 0.128 (n = 135)$	$\beta = -0.138, p = 0.640 (n = 106)$	$\beta = -0.238, p = 0.367 (n = 11)$
Trial length	$\beta = 0.001, p = 0.645 (n = 134)$	$\beta = -0.002, p = 0.202 (n = 105)$	$\beta = -0.002, p = 0.669 (n = 11)$
Dosing duration	$\beta = -0.005, p = 0.531 (n = 135)$	$\beta = -0.003, p = 0.844 (n = 106)$	$\beta = -0.014, p = 0.261 (n = 11)$
Caffeine dose	$\beta = 0.037, p = 0.465 (n = 30)$	$\beta = -0.065, p = 0.170 (n = 26)$	—
Creatine dose	$\beta = 0.057, p = 0.551 (n = 10)$	$\beta = 0.077, p = 0.448 (n = 9)$	—

Glycerol dose	$\beta = -0.340, p = 0.837 (n = 11)$	$\beta = -2.958, p = 0.124 (n = 14)$	_____
Sodium citrate dose	$\beta = 2.157, p = 0.225 (n = 5)$	$\beta = -0.181, p = 0.911 (n = 5)$	_____
Nitrate dose	$\beta = -0.006, p = 0.895 (n = 8)$	$\beta = -0.002, p = 0.967 (n = 5)$	_____
Glutamine dose	$\beta = 0.015, p = 0.980 (n = 10)$	$\beta = -0.085, p = 0.919 (n = 4)$	_____
Tyrosine dose	$\beta = -0.003, p = 0.183 (n = 8)$	$\beta = -0.001, p = 0.865 (n = 7)$	_____
Bovine colostrum dose	$\beta = -0.001, p = 0.911 (n = 4)$	_____	_____
BCAA dose	$\beta = 0.045, p = 0.389 (n = 4)$	_____	_____

90 **3.6 Risk of bias**

91 The studies included had a generally ‘low’ or ‘unclear’ risk of bias, with only 14 studies stating randomization
92 procedures (26-28, 80-90) and three studies with pre-post intervention designs not randomizing or blinding (91-
93 93). Allocation concealment was ‘high’ in three studies (91-93); Figure 5). A number of outliers were detected in
94 the peak T_{core} meta-analysis (91, 94-103), owing to the large effects that were elicited by some supplements on
95 peak T_{core} responses, but Egger’s test showed that there was no publication bias ($p = 0.427$). Several outliers were
96 detected in the WBSR meta-analysis (104-107); however, Egger’s test indicated no publication bias ($p = 0.358$),
97 and influence analysis demonstrated no outcome changes when these were removed. No outliers or publication
98 bias ($p = 0.638$) were detected in the LSR meta-analysis (Figure 6).

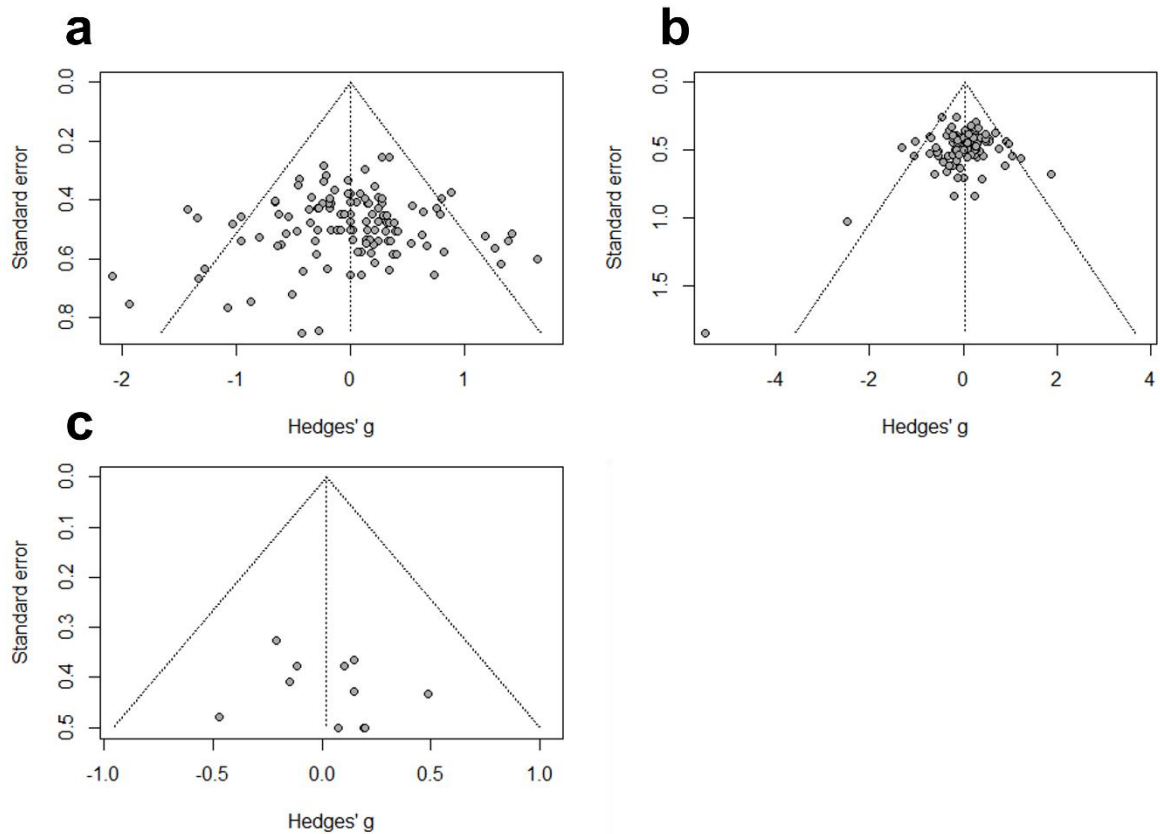
99



100

101 **Figure 5.** Risk of bias.

102



103

104 **Figure 6.** Publication bias for (a) peak core temperature, (b) whole-body sweat rate and (c) local sweat rate.

105

106 **4. Discussion**

107 The main findings of the current meta-analyses were that, overall, pooled analysis of all dietary supplements
 108 demonstrated no effect on peak T_{core} in the heat (Figure 2). However, there were differences between supplements,
 109 with caffeine, taurine and oligonol significantly affecting peak T_{core} responses, but to varying degrees and
 110 directions. Caffeine supplementation appeared to induce a thermogenic effect, while other supplements, such as
 111 taurine and oligonol lowered T_{core} responses compared to placebo. This is consistent with our previous meta-
 112 analytical findings on the thermal effects of caffeine and taurine during exercise in the heat (16), but this work has
 113 now expanded the evidence to a wider pool of supplements, across both resting and exercising conditions. Further,
 114 the additional analyses of sweating responses revealed that, collectively, dietary supplements may increase WBSR
 115 (Figure 3) but have limited effects on LSR, which is likely to be due to the smaller number of studies included in
 116 the analysis (Figure 4). Despite this, there was variation across supplements regarding their effects on sweating,
 117 with taurine demonstrating the greatest increases in WBSR and LSR, and others such as oligonol and folic acid,

118 reducing these responses. In the heat, sweating is the primary heat loss avenue, and as such, is responsible for
119 limiting thermal gain (i.e. T_{core} increases). The findings herein, which indicate that dietary supplements may
120 influence these aspects of thermoregulation, have implications for individuals exposed to hot environmental
121 conditions. Further, as demonstrated by the meta-regression analysis (Table 2), there were also no moderating
122 effects of training and heat acclimation status, hydration status, fluid ingestion during the trial and WBGT on the
123 supplements' capacity to alter T_{core} and sweating responses. Thus, the effects reported are unlikely to be altered
124 by other modifiable factors.

125

126 **4.1 Nitrate, L-arginine and folic acid**

127 In regard to WBSR and LSR, the current analysis revealed a *small* and *trivial*, non-significant positive effect of
128 nitrate, which is a supplement well-established to improve NO bioavailability (32, 108). It is theorized that NO
129 may contribute to eccrine sweat gland function, as local inhibition of NO synthase - NO's precursor - with N^G-
130 nitro-L-arginine methyl ester (L-NAME), has been shown to attenuate sweat rate during moderate exercise in the
131 heat (35, 109), though not all studies provide support for this (110). Further, NO also appears to have a role in
132 regulating cutaneous vasodilation (111-113). Interestingly, numerous studies, in isolation, reported no significant
133 increases in sweating (80, 92, 114-117), which equated to a *small* non-significant effect based on the collective
134 evidence of the current meta-analysis. Therefore, the degree to which nitrate supplementation augments the
135 sweating response is likely to be insufficient to elicit substantial thermoregulatory benefits. This is supported by
136 the findings of the peak T_{core} analysis herein, where increases in sweating did not translate to reductions in T_{core} .
137 These findings, in combination, therefore, question whether nitrate supplementation has the capacity to aid
138 thermal balance in hot environmental conditions. In line with this, supplementation with other precursors to NO,
139 specifically L-arginine and folic acid, demonstrated no significant thermoregulatory improvements, as peak T_{core}
140 was not lower and sweat rate was not increased following their supplementation. Together, these results suggest
141 that supplementation strategies aiming to increase NO bioavailability fail to enhance the sweating responses, and
142 thus, opportunity to evaporatively cool when used in a hot environment. Interestingly, many of these studies also
143 demonstrated no greater cutaneous blood flow or dry heat transfer capacity, thereby questioning any
144 thermoregulatory advantage of nitrate or NO donors in the heat (80, 93, 118). Consequently, supplementation with
145 ~4.2 to 16.8 mmol dietary nitrate, 10 g L-arginine or 5 mg folic acid cannot currently be recommended during
146 heat exposure to improve thermal balance, but do not appear to have any deleterious effects.

147

148 4.2 Caffeine

149 The peak T_{core} analysis demonstrated that caffeine and caffeine in combination with ginseng had *small* and *large*
150 significant positive effects, respectively. Substantial rises in T_{core} can cause heat strain and, ultimately, lead to heat
151 exhaustion and heat stroke if not sufficiently controlled (6, 7). This is particularly the case during heat exposure,
152 or exercise in hot conditions, when avenues of dry heat dissipation are reduced due to a smaller temperature
153 gradient between the ambient air and skin's surface (69, 119). There is evidence that caffeine supplementation
154 increases $\dot{V}O_2$ and, consequently, heat production at comparable exercise intensities compared to placebo (96,
155 120), potentially explaining this observed T_{core} rise. Indeed, the $\dot{V}O_2$ response to exercise has been reported to be
156 increased by 3% to 15% following caffeine supplementation in the heat (98, 99, 121, 122), although others have
157 demonstrated negligible differences (95, 123, 124). These results could be attributed to the methodological
158 differences between studies, as noted by John et al. (83), which calls for improved standardization of laboratory
159 procedures in studies relating to thermoregulation and caffeine supplementation. In addition to the current results,
160 regarding T_{core} , WBSR and LSR were not increased with caffeine supplementation. Given the capacity of sweating
161 to help dissipate excess metabolic heat through evaporative cooling, these *negligible* effects would place greater
162 demand upon dry heat transfer. However, it appears that caffeine-mediated reductions in cutaneous blood flow,
163 owing to vasoconstriction of the skin microvasculature, would preclude this possibility (82, 125). Thus, greater
164 heat production, coupled with reduced SkBF and no increase in sweat production, explains why heat retention
165 would ensue and the observed rises in T_{core} reported herein. Indeed, the previous meta-analytical findings support
166 this observation and have been corroborated further by a recent meta-analysis, which demonstrated a $0.1^{\circ}\text{C}\cdot\text{h}^{-1}$
167 greater T_{core} rate of rise following caffeine supplementation (126). Interestingly, combined caffeine and taurine
168 only had a *trivial* effect on peak T_{core} , theoretically due to taurine's thermolytic effect negating the additional heat
169 production elicited by caffeine. However, without $\dot{V}O_2$ data and corroboration of these findings, this remains
170 speculative. Overall, these results suggest that caffeine has an undesirable (i.e. heat gaining) effect on thermal
171 balance and questions its use in hot environmental conditions. Therefore, acute caffeine intake of between 3 to 9
172 $\text{mg}\cdot\text{kg}^{-1}$ prior to exercise, or at rest during heat exposure, should potentially be avoided due to its thermogenic
173 effect in such conditions. While caffeine intake has not been directly linked to heat illness cases, substances that
174 elevate T_{core} could result in an increased risk of heat related illnesses (127, 128).

175

176 4.3 Taurine

177 The current evaluation of sweating responses revealed that taurine had a *medium*, significant effect on peak T_{core}
178 and WBSR, as well as a *small*, non-significant positive effect on LSR. These studies suggest that ingestion of
179 taurine prior to exercise in the heat augmented the sweating response by hastening sweat onset (26) and increasing
180 sweat rate (26, 27). Theoretically, this would improve thermal balance, due to enhanced latent heat transfer and
181 reduced heat storage, delaying rises in T_{core} (1, 5). While this indicates that taurine can exert a beneficial
182 thermoregulatory effect, these findings require replication, including further insight into its mechanisms of action,
183 which are poorly understood. More thorough elucidation of taurine's effects on sweating and the consequential
184 impact on heat transfer and heat tolerance is necessary, alongside investigation of these mechanisms. However,
185 based on these studies, acute intake of $50 \text{ mg} \cdot \text{kg}^{-1}$ of taurine prior to exercise in the heat induces an earlier sweating
186 onset, greater sweating rate and lower T_{core} responses and may offer a strategy to improve thermoregulatory
187 capacity.

188

189 4.4 Tyrosine, BCAAs and GABA

190 Tyrosine and BCAAs demonstrated no significant effect on peak T_{core} or WBSR. Despite these amino acids
191 previously being reported to provide some of the greatest performance effects of any supplements during exercise
192 in the heat (16), they do not appear to confer thermoregulatory benefits. The ability to reduce central fatigue is
193 often ascribed to these supplements to explain their ergogenicity in such conditions (129-132), but no apparent link
194 to temperature regulation was found in this meta-analysis. However, tyrosine is an essential substrate for tyrosine
195 hydroxylase, which is involved in axonal catecholamine synthesis (particularly norepinephrine; (133)). Thus,
196 sufficient tyrosine availability is required to maintain catecholamine levels and facilitate sympathetic
197 vasoconstrictive effects on the subcutaneous vasculature (134). This has been reported to attenuate the rate of
198 cold-induced decline in T_{core} among those likely to have tyrosine deficiency (134), but there were no similar effects
199 reported across studies conducted in the heat. Interestingly, another amino acid, GABA, has some potential to
200 offer thermoregulatory benefits, with the peak T_{core} analysis revealing a *small*, yet non-significant thermolytic
201 effect. While one of the two studies that supplemented GABA demonstrated null peak T_{core} effects, there was a
202 slower rate of rise in T_{core} across the 30-min exercising period (135). GABA is a widely distributed
203 neurotransmitter within the central nervous system, where it acts in the hypothalamus to regulate internal body
204 temperature (136-138). Exogenous supplementation in humans is thought to increase GABA's availability in the

205 hypothalamus (139) and, thereby, influence temperature regulation (135). The hypothalamus contains cold-
206 sensitive neurons, which have a role in controlling heat production upon detection of local and peripheral
207 temperature changes (140, 141). In the animal model, suppression of these neurons appears to occur following
208 GABA administration, leading to lowered T_{core} responses in the heat (142). Indeed, the two original research
209 articles included in the current meta-analysis (135, 143) observed reductions in metabolic heat production, which
210 could in part explain the effects on T_{core} in the GABA supplementation conditions. Additionally, GABA attenuates
211 activity of the sympathetic nervous system (144, 145), which would likely suppress epinephrine and
212 norepinephrine release (146), as observed by Miyazawa et al. (143). Reductions in catecholamines have been
213 associated with slower rises in T_{core} during hyperthermic exercise (147), which is supported by the findings herein.
214 Considering these effects on heat production, it is unsurprising that there was also a *large*, significant reduction
215 in WBSR and a *trivial* effect on LSR, as it is a known driver of the thermal sweating response (5, 66, 68).
216 Therefore, while GABA appears to reduce one avenue of heat dissipation (i.e. evaporative cooling), it has created
217 a greater heat storage capacity, which would delay the onset of hyperthermic symptoms during heat stress and
218 may be effective during short duration exercise in the heat. Based on the two studies herein, the administration of
219 1 g of GABA directly prior to heat exposure (rest or exercise) appears to provide a beneficial effect on thermal
220 balance through a reduction in heat gain. However, acute tyrosine and BCAA intake would not be a useful
221 supplement in hot environmental conditions, due to the null impacts on thermoregulation.

222

223 **4.5 Glycerol and creatine**

224 The peak T_{core} analysis herein revealed *small*, non-significant negative effects for glycerol and combined creatine
225 and glycerol supplementation. *Medium*, non-significant negative effects were observed for combined creatine,
226 glycerol and glucose and *trivial*, non-significant effects for creatine and combined creatine, glycerol, glucose and
227 alpha lipoic acid. While these results demonstrate that glycerol supplementation had a *small-to-medium* effect on
228 peak T_{core} , the variation across studies decreased the certainty of these findings, rendering them non-significant.
229 When ingested, glycerol provides osmotic pressure in the plasma and intra- and extra-cellular water compartments
230 – where concentrations are evenly distributed – and thereby increase their water content (15, 148, 149). Creatine
231 acts similarly, as its transport into cells (primarily skeletal muscle) increases total body water through expansion
232 of intra- and extracellular water compartments, with even fluid distribution (150-152). This increase to total body
233 water and plasma volume expansion induces ‘hyperhydration’ and affords excess fluid to compensate for sweat

234 losses (153, 154), alongside providing greater availability to sweat glands to facilitate sweat production (155).
235 This would likely improve thermoregulatory capacity, through increased evaporative cooling, but also because
236 additional total body water enhances the specific heat carrying capacity of the tissues and blood (156-158). Here,
237 it can help transfer heat from the core to the periphery to be dissipated (159-161).

238 Creatine had a *trivial*, non-significant positive effect on WBSR, and combined creatine and glycerol, combined
239 creatine and glycerol with the addition of glucose and combined creatine and glycerol with the addition of glucose
240 and alpha lipoic acid had *small*, non-significant positive effects. The role of glycerol combined with creatine was,
241 therefore, also partially effective in promoting a sweating response but, as with the peak T_{core} responses, the effects
242 were inconsistent across studies, which increased the uncertainty of the *small* effects. Surprisingly, glycerol alone
243 had a *small*, non-significant negative effect on WBSR. Though, this was largely influenced by two glycerol
244 supplementation conditions from the same study, where much larger fluid losses in the placebo group were
245 reported (104). Collectively, it appears that these supplements may be capable of lowering T_{core} and enhancing
246 sweating responses compared to placebo, with the combination of creatine and glycerol potentially providing the
247 greatest thermoregulatory benefit. However, with the large inconsistencies between studies and non-significant
248 findings this is far from established, and further work is required to understand the heterogeneity of responses.
249 For WBSR this could be dose related, as for glycerol, Dini et al. B (104) provided the highest glycerol dose (3
250 $\text{g}\cdot\text{kg}^{-1}$) and observed a large, negative effect on WBSR. Theoretically, glycerol ingestion of a large quantity may
251 surpass concentrations that can be absorbed into the intra- and extra-cellular fluid, further elevating plasma
252 concentrations and increasing osmotic pressure. This may prevent fluid being drawn from the plasma to the sweat
253 glands, thereby decreasing sweat rate. Without further investigation into the effect of glycerol dose on the sweating
254 response, more specifically, this remains speculative. Indeed, despite the outcome from this single study, the
255 moderating effect of glycerol dose on WBSR was not significant. Overall, these findings demonstrate that lower
256 doses of glycerol (1 to $1.4\text{ g}\cdot\text{kg}^{-1}$), alone or in combination with 20 to $25\text{ g}\cdot\text{d}^{-1}$ of creatine for between 3 to 9 days,
257 appear to aid thermal balance during exercising heat stress, through hyperhydration. Additional investigation into
258 whether this supplementation strategy would provide similar benefits during passive heating is also warranted.
259 Higher doses of glycerol (e.g. $3\text{ g}\cdot\text{kg}^{-1}$), however, may reduce this capacity due to lower sweat rates, though a
260 greater understanding is necessary before providing definitive recommendations.

261

262 **4.6 Sodium citrate and sodium bicarbonate**

263 As supplements often ingested prior to high-intensity exercise to improve blood buffering capacity, both sodium
264 citrate and sodium bicarbonate have also been reported to increase plasma osmolality and plasma volume (162).
265 This could feasibly help with thermoregulation in the same manner as creatine or glycerol loading; indeed,
266 ingestion of sodium citrate had a *small*, yet non-significant negative effect on peak T_{core} , which was similar to the
267 previous supplements detailed above. However, this was not coupled with a greater WBSR. Conversely, sodium
268 bicarbonate had a *small*, non-significant positive effect on T_{core} . In these two studies, the placebo group ingested
269 sodium chloride to match the sodium content of the two conditions, as they were investigating the buffering
270 capacity of the supplement and not its effects on fluid balance (162-164). Therefore, it is likely that any potential
271 osmotic effects that could theoretically have aided thermoregulation, would be indistinguishable from the effects
272 in the placebo condition. In support of this, there was a *small*, non-significant positive effect on WBSR and no
273 effect on LSR. To establish whether sodium bicarbonate's effects on fluid balance can aid thermoregulatory
274 function in hot environmental conditions, studies would need to be conducted with a placebo group that does not
275 contain any sodium. It appears that sodium citrate can potentially improve thermoregulatory capacity in the heat,
276 though this was inconsistent across studies. Any thermolytic effect is likely due to its effects on plasma volume,
277 as expansion was observed across all studies included in the analysis, but to a larger degree in the studies which
278 demonstrated lower T_{core} responses. As there was no greater WBSR associated with the lower T_{core} response, a
279 greater heat carrying capacity of the blood may be responsible for these observed effects (156, 158). However,
280 further research is required to corroborate these findings and establish whether sodium bicarbonate can elicit the
281 same benefits. Therefore, based on the studies included in this analysis, there is evidence to suggest that acute
282 sodium citrate ingestion of ~ 100 to $600 \text{ mg}\cdot\text{kg}^{-1}$ can improve thermoregulatory capacity during exercise in the
283 heat, but more research is needed to establish these effects at rest and with sodium bicarbonate supplementation.

284

285 **4.7 Betaine**

286 Betaine is an amino acid, which acts as both an osmolyte, to assist with cell volume regulation, and as a methyl
287 group donor to convert homocysteine to methionine (165). It can be supplemented to reduce high plasma
288 concentrations of homocysteine (166) or to improve endurance and resistance exercise performance (167). Due
289 to its osmotic role, it has mechanistic potential to aid fluid balance and thermoregulation during exposure to heat
290 stress (168). However, both the current peak T_{core} and WBSR analyses demonstrated no effects. In addition to the
291 measured variables in the current meta-analysis, one of the included studies observed plasma volume expansion

292 across the study in response to betaine supplementation (169); however, the other did not (168). Together, these
293 results question whether betaine is efficacious for fluid balance when ingested prior to exercise in the heat. There
294 is also some evidence to suggest that betaine may attenuate thermal cellular stress in a similar manner to heat
295 shock proteins (170, 171) and in animal models, it has repeatedly been demonstrated to reduce T_{core} when
296 chronically supplemented (172-174). Therefore, betaine may have the capacity to improve heat tolerability, and
297 considering the limited and equivocal evidence in humans, this supplement requires further investigation.

298

299 **4.8 Anti-oxidants and anti-inflammatories**

300 In the current meta-analysis, lower peak T_{core} responses for several supplements with known anti-oxidative and
301 anti-inflammatory properties were found. Oligonol and catechin, had a *medium*, significant and a *large*, non-
302 significant negative effect on peak T_{core} , respectively. Furthermore, *small*, non-significant negative effects were
303 observed for curcumin, vitamin E and Effective microorganism X (an anti-oxidant mixture) and *trivial*, non-
304 significant negative effects for blackcurrant extract. The anti-inflammatory role of oligonol, catechins, curcumin,
305 vitamin E, Effective microorganism X is most likely responsible for the lowered T_{core} responses compared to
306 placebo, where endogenous pyrogenic cytokines (175) may be attenuated. Indeed, oligonol supplementation
307 lowered circulating levels of the pyrogenic cytokines, such as interleukins IL1- β and IL-6 (176), along with
308 reductions in serum prostaglandin E₂, a known intermediary in the development of fever (177, 178). The cytokine
309 response can be acutely lowered with anti-inflammatory substances (179), theoretically leading to decreased
310 thermal gain (180), explaining why the rate of rise in T_{core} was reduced, despite no greater WBSR or LSR.
311 However, not all of these studies observed reductions in pro-inflammatory cytokines (84, 181), despite attenuated
312 increases in T_{core} . All trials investigating oligonol and catechin – which had the greatest effects – induced heat
313 strain via hot water immersion of the lower body at rest. This is a rapid means by which to facilitate heat gain, as
314 water is highly conductive (182), yet it partially attenuates other avenues of heat dissipation, such as evaporation
315 (1). It is possible that an immersive protocol induced greater thermal strain and production of pyrogenic cytokines,
316 meaning that T_{core} responses were more readily identified between conditions. However, the rise in T_{core} within
317 these trials was less than would be expected, only reaching an average of 37.52°C across all trials; although this
318 was with tympanic measurement, which may explain the relatively low T_{core} values. Another explanation for lower
319 peak T_{core} responses is increased heat dissipation; however, sweating was only greater following catechin and
320 blackcurrant supplementation and these effects were non-significant. While this may partly explain the improved

321 thermal balance, in this instance, it appears likely that reductions in endogenous pyrogenic cytokines have an
322 important role to play in the efficacy of many of the anti-inflammatory supplements. Nevertheless, only oligonol
323 has demonstrated significant impacts on aspects of thermoregulation and, therefore, further investigation of these
324 supplements and mechanisms is required to provide definitive answers.

325 An additional role of anti-oxidants is to improve cellular oxidative capacity and, therefore, redox status (183).
326 These effects could be directly extended to sudomotor function, based on the reported relationships between
327 systemic markers of lipid peroxidation and sweat production (184). However, further research is needed to explore
328 this possibility, owing to the failure of local anti-oxidant infusion to acutely alter the local sweating response
329 during exercise-heat stress (185), which questions the likelihood that anti-oxidants play a major role in
330 thermoregulatory sweating. Indeed, a greater sweating response was not observed for the majority of these
331 supplements. A component of catechin, epicatechin, has been associated with greater cutaneous blood flow during
332 heat exposure through improved NO signaling (186, 187). The results were non-significant, but if substantiated,
333 the observed augmented sweating response may be due to the associated enhancement of skin blood demonstrated
334 in response to catechin supplementation (188). In combination, these would improve evaporative and dry heat
335 transfer, explaining the lower T_{core} response. Similarly, there is evidence that anthocyanins, a key component of
336 blackcurrants promotes production of NO, through augmented NO synthase activity (189). Furthermore,
337 *Eurycoma longifolia* Jack – another supplement with an anti-oxidative function – had a *medium*, positive non-
338 significant effect on WBSR, yet no significant change in peak T_{core} . Neither of these two latter studies measured
339 or estimated SkBF, or characterized other indices of vascular function and, therefore, it can only be speculated
340 that any potential greater WBSR observed – albeit non-significant – is in response to the aforementioned
341 mechanisms.

342 Interestingly, beta-glucan and ginseng, other supplements with anti-inflammatory properties, had *small*, non-
343 significant positive effects on peak T_{core} , and beta-glucan also had no effect on WBSR. While other endogenous
344 pyrogens were significantly reduced immediately post-exercise in the beta-glucan condition compared to placebo,
345 there was a transient elevation of macrophage inflammatory protein 1 β , which may provide a potential explanation
346 for these findings. However, without further investigation into beta-glucans' thermoregulatory effects during heat
347 exposure, any mechanistic explanations remain speculative. Ginseng is a herb with many bioactive ingredients,
348 which has been demonstrated to increase body temperature in the animal model, and may also partially explain
349 this thermogenic effect (190).

350 Other anti-oxidants; vitamin C and polyphenols had no observable effects on peak T_{core} or WBSR. Quercetin and
351 combined α -KG and 5-HMF also had no effect on WBSR, but a *small*, non-significant negative and a *small*, non-
352 significant positive effect on peak T_{core} was observed, respectively. It has been theorized that quercetin, a well
353 characterized anti-oxidant, is capable of inhibiting (via ROS scavenging) the necessary molecular signaling events
354 required to acquire the acclimated phenotype, by reducing the heat-shock factor or hypoxia-inducible factor
355 response to heat exposure (191). Acutely, anti-oxidant intake would improve redox balance and potentially aid
356 heat tolerance, but if supplemented chronically or alongside heat exposure may blunt adaptations (191, 192). A
357 similar argument can be posed for supplements with anti-inflammatory properties (193). Indeed, a greater number
358 of studies demonstrated beneficial peak T_{core} and sweating responses (e.g. catechin, oligonol, quercetin), when
359 supplemented acutely (1 day), but there is no clear consensus on dosing length and supplement efficacy within
360 the current analysis. Nevertheless, based on the required time-course of these cellular adaptations, this mechanism
361 could partially explain the lack of difference between anti-oxidants consumed over longer periods (≥ 7 days) and
362 placebo supplements in the current meta-analysis. Across the anti-oxidant and anti-inflammatory supplements in
363 the current meta-analysis, 10 of the 19 supplements were consumed repeatedly across 3 to 42 days, which means
364 that any potential thermoregulatory effects may have been masked. In summary, the use of anti-oxidants results
365 in variable responses in the heat, which could be partly explained by their multi-ingredient composition, or dosing
366 period. It is important that the specific mechanisms by which these variable effects occur should be investigated,
367 especially if chronic administration of anti-oxidants and anti-inflammatories may reduce adaptation to heat
368 exposure and exacerbate heat illness. Indeed, many of these supplements are more likely to be ingested by people
369 who require anti-oxidative or anti-inflammatory agents, such as older or clinical populations, who are also more
370 vulnerable to heat stress (194, 195) and are also less likely to tolerate increases in T_{core} (i.e. ginseng). Such
371 individuals could benefit from reductions in T_{core} and dietary supplements that may induce this (i.e. oligonol and
372 catechin), assuming that there are no other apparent side-effects.

373 The current analysis suggests that 100 to 200 mg oligonol ingested approximately 30 to 60 min pre-heat exposure
374 has a beneficial effect on thermal balance, by reducing heat gain. Although further investigation into oligonol's
375 efficacy during exercising heat stress is necessary to further elucidate its effects on thermoregulatory capacity.
376 Catechin appears to have a similar effect, though corroboration of this finding is required, as only one study has
377 been conducted thus far. There is also tentative evidence that prolonged intake of 800 mg curcumin, vitamin E,
378 70 mL Effective microorganism X (3, 42 and 7 days, respectively), may reduce T_{core} responses and 600 mg
379 blackcurrant extract and 150 mg Eurycoma longifolia Jack may improve sweat rate during exercise in the heat.

380 However, these results were non-significant and based on only one (curcumin, vitamin E, Effective microorganism
381 X and *Eurycoma longifolia* Jack) or two studies (blackcurrant extract), so these results are not conclusive. Further
382 research is required to establish these anti-inflammatory supplements' efficacy during heat exposure and their
383 effects on endogenous pyrogenic cytokines. Additional investigation into their effects when ingested acutely, at
384 rest and during more ecological valid conditions is warranted before more definitive thermoregulatory effects can
385 be established. Similarly, the potential thermogenic effects of 200 mg ginseng, 250 mg beta-glucan and combined
386 4.8 g α -KG and 60 mg 5-HMF require additional examination, as their intake cannot currently be recommended
387 based on the results herein. Further, ingestion of 250 to 1500 mg vitamin C and polyphenols does not appear to
388 influence thermoregulatory responses (T_{core} or sweat rate) during exercising heat stress and, therefore, while intake
389 is unlikely to facilitate improved thermal balance, it is also unlikely to have detrimental performance or health
390 consequences. However, establishing these effects following acute doses may reveal further impacts on
391 thermoregulatory capacity.

392

393 **4.9 L-glutamine, bovine colostrum, probiotics, whey protein and amino acids**

394 L-glutamine, bovine colostrum and probiotic supplementation all had no effects on peak T_{core} and WBSR,
395 suggesting that they confer no thermoregulatory benefit in the heat. These supplements, along with curcumin, are
396 often ingested prior to exercise in hot environmental conditions, with the aim of maintaining gastrointestinal (GI)
397 barrier integrity and reducing symptoms of GI dysfunction. Gastrointestinal injury and changes to epithelial
398 permeability are relatively common during exercising heat stress (196), which consequently, leads to translocation
399 of endotoxins and bacterial lipopolysaccharides into the central circulation, causing systemic endotoxemia (197).
400 The subsequent release of pro-inflammatory cytokines can, in turn, cause cytokemia and additional elevations in
401 T_{core} (197-199). However, evidence for these supplements' efficacy is equivocal, along with their function as
402 ergogenic aids in the heat. Favorable effects of supplementation with whey protein (106) and an amino acid
403 beverage have been demonstrated on GI permeability during exercising heat stress, where there was a *small*, non-
404 significant negative, and a *trivial* effect on peak T_{core} , respectively. Whey protein supplementation also induced a
405 *large*, significant negative effect on WBSR. In this study the whey protein condition had a lower circulatory
406 endotoxin concentration post-exercise compared with placebo (106), which theoretically may explain any T_{core}
407 differences. The large inhibitory effect on sweating was unexpected, but given the lower T_{core} – and likely heat
408 production –, the drive for sweating would be reduced (67). The only other supplement to display any potential

409 improvements to thermoregulatory capacity is curcumin – as detailed previously – which is more likely to be due
410 to its aforementioned anti-inflammatory role. Probiotics and bovine colostrum supplementation did not reduce
411 circulating endotoxin or cytokine concentrations in the studies within which these were measured (200-203) and
412 only one study which supplemented L-glutamine demonstrated reductions in endotoxins and TNF- α (204). These
413 supplements may be less effective at preventing GI injury in the heat, due to greater redistribution of blood flow
414 from the gut (GI ischemia) to the peripheral vasculature (205) and, consequently, have no influence of T_{core}
415 responses. Indeed, only a few studies identified improvements to GI barrier integrity (85, 103, 204, 206) and
416 largely attributed this improvement to upregulation of heat shock protein 70, which inhibits inflammation (103,
417 204, 207). Therefore, the long-term use of probiotics (7 to 28 days) and ~20 to 140 g bovine colostrum (7 to 14
418 days) and acute use of 0.15 to 0.9 g·kg⁻¹ L-glutamine to maintain GI barrier integrity in hot environmental
419 conditions appears to provide no thermoregulatory advantage. Whilst supplements targeting the GI tract during
420 heat stress are an area of ongoing interest, further research is required to establish other efficacious alternatives.
421 Indeed, an acute dose of whey protein (15 g) may provide an effective option (106), but without replication of
422 these results, this cannot be definitively stated.

423

424 **4.10 Menthol and Thermo Speed Extreme**

425 The oral supplementation of menthol non-significantly lowered peak T_{core} . This reduction was unanticipated but
426 the variability across studies explains the non-significant effect. Menthol is typically considered to be a non-
427 thermal cooling agent (208), which evokes the perception of cooling via transduction of the TRPM8 receptors in
428 the oral cavity (209-212) and possibly the viscera (213), without directly affecting thermal balance according to
429 current literature (214-216). However, current findings were heavily influenced by a single study's results, where
430 T_{core} was estimated by tympanic measurement (100), which can be less reliable if the correct procedures are not
431 adhered to. Therefore, there is some doubt over these results. As discussed in Barwood and colleagues' expert-led
432 consensus article (208), there are some reports of menthol causing vaso-reactivity in the skin's subcutaneous
433 vasculature when applied externally, but no consensus was reached on any form of menthol administration and
434 thermoregulatory effects. Therefore, replication of this single study may be required to confirm whether acute
435 menthol ingestion can mechanistically affect temperature regulation. Additionally, there was no effect on WBSR,
436 which supports the current consensus (208). Thermo Speed Extreme is another supplement which did not affect
437 peak T_{core} and given that this supplement contains ingredients such as caffeine (217), this finding is somewhat

438 unexpected. It is possible that the tympanic measures used within this study were insufficiently sensitive to detect
439 T_{core} changes. However, significantly greater chest T_{sk} was observed at certain time-points across the trial, which
440 could enhance dry heat dissipation to the environment, particularly as the ambient air was much cooler (26.2°C)
441 than average T_{sk} across participants (34°C). The ingredient piperine could be responsible for this likely
442 enhancement of cutaneous vasodilation, as *in-vitro* studies suggest it may have vaso-modulating effects (218).
443 This could explain the tendency towards lower T_{core} values, despite the thermogenic nature of the supplement. It
444 should be stated that this supplement would, therefore, not be appropriate for use in ambient temperatures that
445 exceed T_{sk} , where skin surface to ambient air temperature gradients, and dry heat transfer capacity are reduced.

446

447 **4.11 Moderating effects**

448 No candidate moderators, such as training and heat acclimation status, hydration status and fluid ingestion during
449 the trial, affected peak T_{core} or sweating responses to dietary supplementation. For hydration status, this is likely
450 due to the majority of papers stipulating the inclusion of hydrated participants. However, there was more variation
451 in the training (highly trained; 43% vs recreationally active; 39%) and heat acclimation (acclimated; 14% vs non-
452 acclimated; 36%) status of participants and whether fluid was ingested during the trials (ingested; 46% vs not
453 ingested; 22%), yet no moderating effects were found. Nevertheless, it would be useful for future studies to
454 consider investigating the efficacy of dietary supplements on thermoregulation among participants of different
455 training and heat acclimation statuses, given the effect of these processes on sweating and T_{core} responses (11).
456 Some supplements, such as sodium citrate, nitrate, L-glutamine and tyrosine, have only been used to assess
457 thermoregulatory responses in non-acclimated participants, which limits the wider application of these to potential
458 end-users. Whether this would augment or negate any effects seen from these supplements is of particular interest
459 and important to establish for individuals in competitive sport, military and occupational settings. In addition, all
460 other meta-regressions (WBGT, trial type and length and supplementation period) did not moderate the effect of
461 dietary supplementation on peak T_{core} or sweating. There is a large range of supplements included within the
462 current meta-analyses, each with differing underpinning mechanisms and nuances in their efficacy. It is, therefore,
463 unsurprising that there are no consistent moderating factors.

464 While these trial moderators had no significant effects in the present analysis, they still require further
465 investigation, particularly within the most efficacious supplements included here. For example, acute
466 supplementation and the use of exercise was most common, with no variation within certain supplement

467 categories. The effect of chronic supplementation of certain supplements, such as various anti-oxidants, glycerol,
468 taurine, and other amino acids (e.g. L-glutamine), on T_{core} and sweating responses in the heat is almost completely
469 unknown. Taurine has been shown to elicit various physiological effects following chronic supplementation,
470 which may be advantageous during heat exposure, such as enhanced vascular function (25) and an improved
471 endurance trained phenotype (219-221). All studies investigating the effects of L-glutamine on GI barrier integrity
472 in the heat have supplemented acutely and it is possible that a chronic dose may be more efficacious. Indeed, long-
473 term administration (2 months) has demonstrated beneficial effects on GI permeability in patients with Crohn's
474 disease (222). Longer term glycerol intake has previously elicited hyperhydration for up to 49 hours (223), but
475 whether it can be maintained over a greater period of time is currently unknown. Further research into this,
476 alongside potential side effects (e.g. hyponatremia) would help establish whether glycerol supplementation can
477 provide long-term beneficial effects on thermal tolerance. Additionally, as detailed above, the chronic and acute
478 effects of various anti-oxidants and anti-inflammatory supplements in the heat requires investigation.
479 Furthermore, establishing the efficacy of these supplements during differing trial types with differing
480 physiological demands is necessary to be able to provide practical advice and application to athletes, workers and
481 the general population. The meta-regression of WBGT demonstrated no effect, but ambient vapor pressure does
482 have an established impact on avenues of heat dissipation (69). For example, supplements which augment thermal
483 sweating (e.g. taurine) will likely be most effective in dry environments where any sweat produced can be
484 evaporated, thereby providing a cooling effect. Depending on the mechanistic actions of certain supplements,
485 beneficial thermoregulatory effects, this could have a large impact on their efficacy and ability to help individuals
486 maintain thermal equilibrium. Investigation of these above factors is important, particularly amongst the most
487 efficacious of the supplements examined within these meta-analyses.

488

489 **4.12 Limitations**

490 Within these meta-analyses, several supplements were taken in combination, such as creatine and glycerol (91,
491 224, 225), caffeine and ginseng (94), caffeine and taurine (28), combined α -KG and 5-HMF (226), whey protein
492 (106), amino acid beverage (227), Effective microorganism X (228) and Thermo Speed Extreme (217). However,
493 as only a few studies employed a co-ingestion strategy, there is limited information on the thermoregulatory
494 outcomes when using this approach across a wide range of different supplements. Herein, the combined effect of
495 creatine and glycerol was beneficial for thermal balance, while the co-ingestion of caffeine and ginseng further

496 exacerbated caffeine's thermogenic effect. As such, the former could be recommended to improve fluid balance
497 in the heat; however, the latter may pose a greater heat stress risk and potentially should be avoided in hot
498 conditions. This is perhaps unsurprising given that caffeine alone causes an increase in T_{core} . Considering these
499 differing findings and the indication that co-ingestion potentially influences the thermoregulatory responses to
500 these supplements, greater clarity across supplement types regarding these effects is certainly warranted. Indeed,
501 athletes and military personnel often combine dietary supplements (229, 230), which may increase the risk of heat
502 stress if a harmful combination is unwittingly ingested. Therefore, further research regarding the effect of dietary
503 supplement co-ingestion on thermoregulatory responses during heat exposure is necessary and represents a key
504 gap in the literature and a further lack of specific supplementation guidance for potential users.

505

506 **5. Conclusion**

507 In summary, for the first time, the effects of various dietary supplements on T_{core} and sweating responses in the
508 heat have been evaluated. The amino acids taurine and GABA, alongside whey protein, lowered peak T_{core} ,
509 indicating an improvement to thermal balance. While GABA and whey protein negatively impacted WBSR,
510 taurine increased the sweating response, demonstrating an enhancement to thermoregulatory capacity, albeit from
511 a single study. However, other amino acids, such as tyrosine and BCAAs appeared to have no meaningful effect
512 on thermoregulation. Various supplements with anti-oxidative and anti-inflammatory properties (e.g. oligonol,
513 catechin, curcumin, vitamin E and quercetin) provided beneficial effects on peak T_{core} , which may in-part be
514 explained through improved redox balance and attenuation of endogenous pyrogenic cytokines. Nevertheless, not
515 all of these supplements improved thermal balance, highlighting the need for additional research in this area. A
516 number of supplements (e.g. glycerol, creatine, sodium citrate and betaine), which appear to induce
517 hyperhydration and/or expand plasma volume, non-significantly lowered T_{core} responses. Mechanistically, this
518 may be through increasing heat carrying capacity and/or improving fluid availability to the sweat gland, as some
519 supplements (e.g. combined glycerol and creatine) also demonstrated an effect on sudomotor function. However,
520 T_{core} and sweat rate findings were inconsistent across studies and supplement types, rendering the results non-
521 significant overall. Many other supplements such as nitrate, L-arginine, folic acid (taken for their effects on NO
522 bioavailability) and L-glutamine, bovine colostrum and probiotics (taken for their effects on GI barrier integrity)
523 did not appear to provide any thermoregulatory benefit in the heat. Peak T_{core} was greater following caffeine and
524 combined caffeine and ginseng supplementation, without any increases in sweating responses. Consequently,

525 caffeine when ingested during heat exposure appears to be thermogenic and, therefore, may have potential
526 negative health implications. Several other supplements, such as ginseng, beta-glucan and combined α -KG and 5-
527 HMF also demonstrated *small* thermogenic effects, though these results were non-significant.

528 Although additional investigation is certainly required, some supplements have demonstrated the potential to
529 improve thermoregulatory capacity in the heat. However, it appears that others have null or even deleterious
530 effects on thermal balance when ingested in such conditions. These findings suggest that certain supplements,
531 such as caffeine, should possibly be avoided in hot conditions and others, such as taurine, may elicit a
532 thermoregulatory benefit. This has potential implications for those ingesting dietary supplements for their health
533 and/or performance effects during periods of heat exposure. Indeed, official guidance documents for the general
534 population, athletes and military personnel could also be updated to reflect the varying effects different dietary
535 supplements appear to have on thermoregulation, detailing which to avoid and which may be advantageous in hot
536 conditions. Additional investigation into many of these supplements is required to corroborate findings and
537 provide greater understanding of their effects. Specifically, future research should focus on the thermolytic effects
538 of various supplements such as taurine, GABA, oligonol and catechin in varying conditions, alongside further
539 mechanistic insight into these responses.

540

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545 **Author contributions**

546 JP and MW conceived and designed research, performed the searches, screening of manuscripts and study quality
547 scoring. JP performed the data analysis and prepared figures. All authors (JP, MW, MM, SH, VN and LK) assisted
548 with data interpretation, drafting, editing and revising of the manuscript, as well as reading and approving the
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550

551 **References**

- 552 1. **Gagge A, and Gonzalez R.** Handbook of Physiology. Environmental Physiology. *Bethesda, MD:*
553 *American Physiological Society* 45-84, 1996.
- 554 2. **Wenger CB.** Heat of evaporation of sweat: thermodynamic considerations. *Journal of applied*
555 *physiology* 32: 456-459, 1972.
- 556 3. **Marino FE, Mbambo Z, Kortekaas E, Wilson G, Lambert MI, Noakes TD, and Dennis SC.**
557 Advantages of smaller body mass during distance running in warm, humid environments. *Pflügers*
558 *Archiv* 441: 359-367, 2000.
- 559 4. **Sawka MN, and Young AJ.** Physiological systems and their responses to conditions of heat and
560 cold Army Research Inst Of Environmental Medicine Natick Ma Thermal and Mountain ..., 2006.
- 561 5. **Cramer MN, and Jay O.** Biophysical aspects of human thermoregulation during heat stress.
562 *Autonomic Neuroscience* 196: 3-13, 2016.
- 563 6. **Liu J, Varghese BM, Hansen A, Zhang Y, Driscoll T, Morgan G, Dear K, Gourley M, Capon A,**
564 **and Bi P.** Heat exposure and cardiovascular health outcomes: a systematic review and meta-analysis.
565 *The Lancet Planetary Health* 6: e484-e495, 2022.
- 566 7. **Székely M, Carletto L, and Garami A.** The pathophysiology of heat exposure. *Temperature* 2:
567 452-452, 2015.
- 568 8. **Benzinger T, Pratt A, and Kitzinger C.** The thermostatic control of human metabolic heat
569 production. *Proceedings of the National Academy of Sciences* 47: 730-739, 1961.
- 570 9. **Wendt D, Van Loon LJ, and Marken Lichtenbelt WD.** Thermoregulation during exercise in the
571 heat: strategies for maintaining health and performance. *Sports medicine* 37: 669-682, 2007.
- 572 10. **Périard JD, Travers GJ, Racinais S, and Sawka MN.** Cardiovascular adaptations supporting
573 human exercise-heat acclimation. *Autonomic Neuroscience* 196: 52-62, 2016.
- 574 11. **Ravanelli N, Coombs GB, Imbeault P, and Jay O.** Maximum skin wettedness after aerobic
575 training with and without heat acclimation. *Medicine and science in sports and exercise* 50: 299-307,
576 2018.
- 577 12. **Lorenzo S, Halliwill JR, Sawka MN, and Minson CT.** Heat acclimation improves exercise
578 performance. *Journal of Applied Physiology* 109: 1140-1147, 2010.
- 579 13. **Poirier MP, Gagnon D, and Kenny GP.** Local versus whole-body sweating adaptations following
580 14 days of traditional heat acclimation. *Applied Physiology, Nutrition, and Metabolism* 41: 816-824,
581 2016.
- 582 14. **Rivas E, Rao M, Castleberry T, and Ben-Ezra V.** The change in metabolic heat production is a
583 primary mediator of heat acclimation in adults. *Journal of Thermal Biology* 70: 69-79, 2017.
- 584 15. **Jardine WT, Aisbett B, Kelly MK, Burke LM, Ross ML, Condo D, Périard JD, and Carr AJ.** The
585 Effect of Pre-Exercise Hyperhydration on Exercise Performance, Physiological Outcomes and
586 Gastrointestinal Symptoms: A Systematic Review. *Sports Medicine* 53: 2111-2134, 2023.
- 587 16. **Peel JS, McNarry MA, Heffernan SM, Nevola VR, Kilduff LP, and Waldron M.** The effect of
588 dietary supplements on endurance exercise performance and core temperature in hot environments:
589 A Meta-analysis and Meta-regression. *Sports Medicine* 51: 2351-2371, 2021.
- 590 17. **Twycross-Lewis R, Kilduff LP, Wang G, and Pitsiladis Y.** The effects of creatine
591 supplementation on thermoregulation and physical (cognitive) performance: a review and future
592 prospects. *Amino Acids* 48: 1843-1855, 2016.
- 593 18. **Bailey RL, Gahche JJ, Miller PE, Thomas PR, and Dwyer JT.** Why US adults use dietary
594 supplements. *JAMA internal medicine* 173: 355-361, 2013.
- 595 19. **EFSA.** Food supplements <https://www.efsa.europa.eu/en/topics/topic/food-supplements>.
596 [18/01/2024].
- 597 20. **Kantor ED, Rehm CD, Du M, White E, and Giovannucci EL.** Trends in dietary supplement use
598 among US adults from 1999-2012. *Jama* 316: 1464-1474, 2016.
- 599 21. **Swan G.** National diet and nutrition survey. 2016.
- 600 22. **Ronis MJ, Pedersen KB, and Watt J.** Adverse effects of nutraceuticals and dietary supplements.
601 *Annual review of pharmacology and toxicology* 58: 583-601, 2018.

- 602 23. **Perkins-Kirkpatrick S, and Lewis S.** Increasing trends in regional heatwaves. *Nature*
603 *communications* 11: 3357, 2020.
- 604 24. **Schaffer SW, Ito T, and Azuma J.** Clinical significance of taurine. Springer, 2014, p. 1-5.
- 605 25. **Sun Q, Wang B, Li Y, Sun F, Li P, Xia W, Zhou X, Li Q, Wang X, and Chen J.** Taurine
606 supplementation lowers blood pressure and improves vascular function in prehypertension:
607 randomized, double-blind, placebo-controlled study. *Hypertension* 67: 541-549, 2016.
- 608 26. **Page LK, Jeffries O, and Waldron M.** Acute taurine supplementation enhances
609 thermoregulation and endurance cycling performance in the heat. *Eur J Sport Sci* 19: 1101-1109, 2019.
- 610 27. **Peel JS, McNarry MA, Heffernan SM, Nevola VR, Kilduff LP, Coates K, Dudley E, and Waldron**
611 **M.** The effect of 8-day oral taurine supplementation on thermoregulation during low-intensity exercise
612 at fixed heat production in hot conditions of incremental humidity. *European Journal of Applied*
613 *Physiology* 1-16, 2024.
- 614 28. **Yu P, Fan Y, and Wu H.** Effects of Caffeine-Taurine Co-Ingestion on Endurance Cycling
615 Performance in High Temperature and Humidity Environments. *Sports Health* 19417381241231627,
616 2024.
- 617 29. **Branch JD.** Effect of creatine supplementation on body composition and performance: a meta-
618 analysis. *International journal of sport nutrition and exercise metabolism* 13: 198-226, 2003.
- 619 30. **Gao C, Gupta S, Adli T, Hou W, Coolsaet R, Hayes A, Kim K, Pandey A, Gordon J, and Chahil**
620 **G.** The effects of dietary nitrate supplementation on endurance exercise performance and
621 cardiorespiratory measures in healthy adults: a systematic review and meta-analysis. *Journal of the*
622 *International Society of Sports Nutrition* 18: 55, 2021.
- 623 31. **McMahon NF, Leveritt MD, and Pavey TG.** The effect of dietary nitrate supplementation on
624 endurance exercise performance in healthy adults: a systematic review and meta-analysis. *Sports*
625 *Medicine* 47: 735-756, 2017.
- 626 32. **Lundberg JO, Weitzberg E, and Gladwin MT.** The nitrate–nitrite–nitric oxide pathway in
627 physiology and therapeutics. *Nature reviews Drug discovery* 7: 156-167, 2008.
- 628 33. **Moncada S, and Higgs E.** Endogenous nitric oxide: physiology, pathology and clinical relevance.
629 *European journal of clinical investigation* 21: 361-374, 1991.
- 630 34. **Fujii N, Singh MS, Halili L, Boulay P, Sigal RJ, and Kenny GP.** Cutaneous vascular and sweating
631 responses to intradermal administration of prostaglandin E1 and E2 in young and older adults: a role
632 for nitric oxide? *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*
633 310: R1064-R1072, 2016.
- 634 35. **Stapleton JM, Fujii N, Carter M, and Kenny GP.** Diminished nitric oxide-dependent sweating
635 in older males during intermittent exercise in the heat. *Experimental Physiology* 99: 921-932, 2014.
- 636 36. **Ignarro LJ, Byrns RE, Sumi D, de Nigris F, and Napoli C.** Pomegranate juice protects nitric oxide
637 against oxidative destruction and enhances the biological actions of nitric oxide. *Nitric oxide* 15: 93-
638 102, 2006.
- 639 37. **Sawka MN, Francesconi RP, Young AJ, and Pandolf KB.** Influence of hydration level and body
640 fluids on exercise performance in the heat. *Jama* 252: 1165-1169, 1984.
- 641 38. **Benjamin CJR, Porto AA, Valenti VE, Sobrinho ACdS, Garner DM, Gualano B, and Bueno**
642 **Junior CR.** Nitrate derived from beetroot juice lowers blood pressure in patients with arterial
643 hypertension: a systematic review and meta-analysis. *Frontiers in nutrition* 9: 823039, 2022.
- 644 39. **Halliwill JR.** Mechanisms and clinical implications of post-exercise hypotension in humans.
645 *Exercise and sport sciences reviews* 29: 65-70, 2001.
- 646 40. **Hormoznejad R, Zare Javid A, and Mansoori A.** Effect of BCAA supplementation on central
647 fatigue, energy metabolism substrate and muscle damage to the exercise: a systematic review with
648 meta-analysis. *Sport Sciences for Health* 15: 265-279, 2019.
- 649 41. **Holeček M.** Branched-chain amino acids in health and disease: metabolism, alterations in
650 blood plasma, and as supplements. *Nutrition & metabolism* 15: 1-12, 2018.

- 651 42. **Børsheim E, Bui Q-UT, Tissier S, Kobayashi H, Ferrando AA, and Wolfe RR.** Effect of amino acid
652 supplementation on muscle mass, strength and physical function in elderly. *Clinical nutrition* 27: 189-
653 195, 2008.
- 654 43. **Moore J, McClain A, and Hong MY.** Dietary supplement use in the United States: Prevalence,
655 trends, pros, and cons. *Nutrition Today* 55: 174-181, 2020.
- 656 44. **Hase A, Jung SE, and aan het Rot M.** Behavioral and cognitive effects of tyrosine intake in
657 healthy human adults. *Pharmacology Biochemistry and Behavior* 133: 1-6, 2015.
- 658 45. **Hensel C, Becker M, Düzel S, Demuth I, Norman K, Steinhagen-Thiessen E, Gallinat J,
659 Lindenberger U, and Kühn S.** Influence of nutritional tyrosine on cognition and functional connectivity
660 in healthy old humans. *Neuroimage* 193: 139-145, 2019.
- 661 46. **Fedewa MV, Spencer SO, Williams TD, Becker ZE, and Fuqua CA.** Effect of branched-chain
662 amino acid supplementation on muscle soreness following exercise: A meta-analysis. *International
663 Journal for Vitamin and Nutrition Research* 2019.
- 664 47. **Rahimi MH, Shab-Bidar S, Mollahosseini M, and Djafarian K.** Branched-chain amino acid
665 supplementation and exercise-induced muscle damage in exercise recovery: A meta-analysis of
666 randomized clinical trials. *Nutrition* 42: 30-36, 2017.
- 667 48. **Fernstrom JD.** Dietary precursors and brain neurotransmitter formation. *Annual review of
668 medicine* 32: 413-425, 1981.
- 669 49. **Pardridge WM.** Blood-brain barrier carrier-mediated transport and brain metabolism of amino
670 acids. *Neurochemical research* 23: 635-644, 1998.
- 671 50. **Suryawan A, Hawes JW, Harris RA, Shimomura Y, Jenkins AE, and Hutson SM.** A molecular
672 model of human branched-chain amino acid metabolism. *The American journal of clinical nutrition* 68:
673 72-81, 1998.
- 674 51. **Mishra S, Stierman B, Gahche JJ, and Potischman N.** Dietary supplement use among adults:
675 United States, 2017–2018. 2021.
- 676 52. **Heckman MA, Weil J, and De Mejia EG.** Caffeine (1, 3, 7-trimethylxanthine) in foods: a
677 comprehensive review on consumption, functionality, safety, and regulatory matters. *Journal of food
678 science* 75: R77-R87, 2010.
- 679 53. **Mitchell DC, Knight CA, Hockenberry J, Teplansky R, and Hartman TJ.** Beverage caffeine
680 intakes in the US. *Food and Chemical Toxicology* 63: 136-142, 2014.
- 681 54. **Del Coso J, Muñoz G, and Muñoz-Guerra J.** Prevalence of caffeine use in elite athletes
682 following its removal from the World Anti-Doping Agency list of banned substances. *Applied
683 physiology, nutrition, and metabolism* 36: 555-561, 2011.
- 684 55. **de Souza JG, Del Coso J, Fonseca FdS, Silva BVC, de Souza DB, da Silva Gianoni RL, Filip-
685 Stachnik A, Serrão JC, and Claudino JG.** Risk or benefit? Side effects of caffeine supplementation in
686 sport: a systematic review. *European journal of nutrition* 61: 3823-3834, 2022.
- 687 56. **Zulli A, Smith RM, Kubatka P, Novak J, Uehara Y, Loftus H, Qaradakhi T, Pohanka M, Kobyliak
688 N, and Zagatina A.** Caffeine and cardiovascular diseases: critical review of current research. *European
689 journal of nutrition* 55: 1331-1343, 2016.
- 690 57. **Collins J, Maughan RJ, Gleeson M, Bilsborough J, Jeukendrup A, Morton JP, Phillips S,
691 Armstrong L, Burke LM, and Close GL.** UEFA expert group statement on nutrition in elite football.
692 Current evidence to inform practical recommendations and guide future research. *British journal of
693 sports medicine* 2020.
- 694 58. **Maughan RJ, Burke LM, Dvorak J, Larson-Meyer DE, Peeling P, Phillips SM, Rawson ES, Walsh
695 NP, Garthe I, and Geyer H.** IOC consensus statement: dietary supplements and the high-performance
696 athlete. *International journal of sport nutrition and exercise metabolism* 28: 104-125, 2018.
- 697 59. **Thomas DT, Erdman KA, and Burke LM.** Nutrition and athletic performance. *Med Sci Sports
698 Exerc* 48: 543-568, 2016.
- 699 60. **Army Dot.** FM 7-22 Holistic Health and Fitness. edited by Army Dot. Washington D.C.:
700 Department of the Army, 2020.

- 701 61. **Organisation WH.** Climate change and extreme heat events. edited by (CCH) CCaHWorld
702 Health Organisation, 2012.
- 703 62. **Périard JD, Eijvogels TM, and Daanen HA.** Exercise under heat stress: thermoregulation,
704 hydration, performance implications, and mitigation strategies. *Physiological reviews* 2021.
- 705 63. **Sawka MN, and Montain SJ.** Fluid and electrolyte supplementation for exercise heat stress.
706 *The American journal of clinical nutrition* 72: 564S-572S, 2000.
- 707 64. **Sawka M, Latzka W, Matott R, and Montain S.** Hydration effects on temperature regulation.
708 *International journal of sports medicine* 19: S108-S110, 1998.
- 709 65. **Sawka MN, Montain SJ, and Latzka WA.** Hydration effects on thermoregulation and
710 performance in the heat. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative*
711 *Physiology* 128: 679-690, 2001.
- 712 66. **Cramer MN, and Jay O.** Selecting the correct exercise intensity for unbiased comparisons of
713 thermoregulatory responses between groups of different mass and surface area. *Journal of Applied*
714 *Physiology* 116: 1123-1132, 2014.
- 715 67. **Gagnon D, Jay O, and Kenny GP.** The evaporative requirement for heat balance determines
716 whole-body sweat rate during exercise under conditions permitting full evaporation. *The Journal of*
717 *physiology* 591: 2925-2935, 2013.
- 718 68. **Peel JS, McNarry MA, Heffernan SM, Nevola VR, Kilduff LP, and Waldron M.** Measurement of
719 thermal sweating at rest and steady-state exercise in healthy adults: Inter-day reliability and
720 relationships with components of partitional calorimetry. *Plos one* 17: e0278652, 2022.
- 721 69. **Che Muhamed AM, Atkins K, Stannard SR, Mündel T, and Thompson MW.** The effects of a
722 systematic increase in relative humidity on thermoregulatory and circulatory responses during
723 prolonged running exercise in the heat. *Temperature* 3: 455-464, 2016.
- 724 70. **Cramer MN, and Jay O.** Partitional calorimetry. *Journal of applied physiology* 126: 267-277,
725 2019.
- 726 71. **Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P, and Stewart**
727 **LA.** Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015
728 statement. *Systematic reviews* 4: 1, 2015.
- 729 72. **Budd GM.** Wet-bulb globe temperature (WBGT)—its history and its limitations. *Journal of*
730 *science and medicine in sport* 11: 20-32, 2008.
- 731 73. **Government UK.** Misuse of Drugs Act 1971. edited by Legislation U1971.
- 732 74. **WADA.** Prohibited List. 2023.
- 733 75. **Higgins JP, Thomas J, Chandler J, Cumpston M, Li T, Page MJ, and Welch VA.** *Cochrane*
734 *handbook for systematic reviews of interventions.* John Wiley & Sons, 2019.
- 735 76. **Furukawa TA, Barbui C, Cipriani A, Brambilla P, and Watanabe N.** Imputing missing standard
736 deviations in meta-analyses can provide accurate results. *Journal of clinical epidemiology* 59: 7-10,
737 2006.
- 738 77. **Harrer M, Cuijpers P, Furukawa T, and Ebert D.** Doing meta-analysis in R: a hands-on guide.
739 *PROTECT Lab Erlangen* 2019.
- 740 78. **Rothstein HR, Sutton AJ, and Borenstein M.** Publication bias in meta-analysis. *Publication bias*
741 *in meta-analysis: Prevention, assessment and adjustments* 1-7, 2005.
- 742 79. **Rosenthal R, and Rosnow RL.** *Essentials of behavioral research: Methods and data analysis.*
743 2008.
- 744 80. **Fowler R, Jeffries O, Tallent J, Theis N, Heffernan SM, McNarry MA, Kilduff L, and Waldron**
745 **M.** No thermoregulatory or ergogenic effect of dietary nitrate among physically inactive males,
746 exercising above gas exchange threshold in hot and dry conditions. *Eur J Sport Sci* 21: 370-378, 2021.
- 747 81. **Hiles AM, Flood TR, Lee BJ, Wheeler LEV, Costello R, Walker EF, Ashdown KM, Kuennen MR,**
748 **and Willems MET.** Dietary supplementation with New Zealand blackcurrant extract enhances fat
749 oxidation during submaximal exercise in the heat. *J Sci Med Sport* 23: 908-912, 2020.

- 750 82. **Hunt LA, Hospers L, Smallcombe JW, Mavros Y, and Jay O.** Caffeine alters thermoregulatory
751 responses to exercise in the heat only in caffeine-habituated individuals: a double-blind placebo-
752 controlled trial. *J Appl Physiol (1985)* 131: 1300-1310, 2021.
- 753 83. **John K, Kathuria S, Peel J, Page J, Aitkenhead R, Felstead A, Heffernan SM, Jeffries O, Tallent
754 J, and Waldron M.** Caffeine ingestion compromises thermoregulation and does not improve cycling
755 time to exhaustion in the heat amongst males. *European Journal of Applied Physiology* 1-14, 2024.
- 756 84. **Lee BJ, Flood TR, Hiles AM, Walker EF, Wheeler LEV, Ashdown KM, Willems MET, Costello R,
757 Greisler LD, Romano PA, Hill GW, and Kuennen MR.** Anthocyanin-Rich Blackcurrant Extract Preserves
758 Gastrointestinal Barrier Permeability and Reduces Enterocyte Damage but Has No Effect on Microbial
759 Translocation and Inflammation After Exertional Heat Stress. *Int J Sport Nutr Exerc Metab* 32: 265-274,
760 2022.
- 761 85. **March DS, Jones AW, Thatcher R, and Davison G.** The effect of bovine colostrum
762 supplementation on intestinal injury and circulating intestinal bacterial DNA following exercise in the
763 heat. *Eur J Nutr* 58: 1441-1451, 2019.
- 764 86. **Ogden HB, Fallowfield JL, Child RB, Davison G, Fleming SC, Delves SK, Millyard A, Westwood
765 CS, and Layden JD.** No protective benefits of low dose acute L-glutamine supplementation on small
766 intestinal permeability, epithelial injury and bacterial translocation biomarkers in response to
767 subclinical exertional-heat stress: A Randomized cross-over trial. *Temperature* 1-15, 2022.
- 768 87. **Ogden HB, Fallowfield JL, Child RB, Davison G, Fleming SC, Delves SK, Millyard A, Westwood
769 CS, and Layden JD.** Acute (L)-glutamine supplementation does not improve gastrointestinal
770 permeability, injury or microbial translocation in response to exhaustive high intensity exertional-heat
771 stress. *Eur J Sport Sci* 22: 1865-1876, 2022.
- 772 88. **Smith K, Muggeridge DJ, Easton C, and Ross MD.** An acute dose of inorganic dietary nitrate
773 does not improve high-intensity, intermittent exercise performance in temperate or hot and humid
774 conditions. *Eur J Appl Physiol* 119: 723-733, 2019.
- 775 89. **Tumilty L, Gregory N, Beckmann M, and Thatcher R.** No Influence of Low-, Medium-, or High-
776 Dose Tyrosine on Exercise in a Warm Environment. *Med Sci Sports Exerc* 52: 1404-1413, 2020.
- 777 90. **Zabriskie HA, Blumkaitis JC, Moon JM, Currier BS, Stefan R, Ratliff K, Harty PS, Stecker RA,
778 Rudnicka K, Jäger R, Roberts MD, Young K, Jagim AR, and Kerksick CM.** Yeast Beta-Glucan
779 Supplementation Downregulates Markers of Systemic Inflammation after Heated Treadmill Exercise.
780 *Nutrients* 12: 2020.
- 781 91. **Beis LY, Polyviou T, Malkova D, and Pitsiladis YP.** The effects of creatine and glycerol
782 hyperhydration on running economy in well trained endurance runners. *Journal of the International
783 Society of Sports Nutrition* 8: 24-32, 2011.
- 784 92. **Cramer MN, Hieda M, Huang M, Morales G, and Crandall CG.** Dietary nitrate supplementation
785 does not influence thermoregulatory or cardiovascular strain in older individuals during severe
786 ambient heat stress. *Exp Physiol* 105: 1730-1741, 2020.
- 787 93. **Gagnon D, Romero SA, Cramer MN, Kouda K, Poh PYS, Ngo H, Jay O, and Crandall CG.** Folic
788 acid supplementation does not attenuate thermoregulatory or cardiovascular strain of older adults
789 exposed to extreme heat and humidity. *Exp Physiol* 103: 1123-1131, 2018.
- 790 94. **Bandyopadhyay A, Ping FW, and Keong CC.** Effects of acute supplementation of caffeine and
791 Panax ginseng on endurance running performance in a hot and humid environment. *J Hum Ergol
792 (Tokyo)* 40: 63-72, 2011.
- 793 95. **Cheuvront SN, Ely BR, Kenefick RW, Michniak-Kohn BB, Rood JC, and Sawka MN.** No effect of
794 nutritional adenosine receptor antagonists on exercise performance in the heat. *Am J Physiol Regul
795 Integr Comp Physiol* 296: R394-401, 2009.
- 796 96. **Ely BR, Ely MR, and Cheuvront SN.** Marginal effects of a large caffeine dose on heat balance
797 during exercise-heat stress. *Int J Sport Nutr Exerc Metab* 21: 65-70, 2011.
- 798 97. **Lyons T, Riedesel M, Meuli L, and Chick T.** Effects of glycerol-induced hyperhydration prior to
799 exercise in the heat on sweating and core temperature. *Medicine and Science in Sports and Exercise*
800 22: 477-483, 1990.

- 801 98. **Millard-Stafford ML, Cureton KJ, Wingo JE, Trilk J, Warren GL, and Buyckx M.** Hydration
802 during exercise in warm, humid conditions: effect of a caffeinated sports drink. *Int J Sport Nutr Exerc*
803 *Metab* 17: 163-177, 2007.
- 804 99. **Ping WC, Keong CC, and Bandyopadhyay A.** Effects of acute supplementation of caffeine on
805 cardiorespiratory responses during endurance running in a hot & humid climate. *Indian J Med Res* 132:
806 36-41, 2010.
- 807 100. **Riera F, Trong TT, Sinnapah S, and Hue O.** Physical and perceptual cooling with beverages to
808 increase cycle performance in a tropical climate. *PLoS One* 9: e103718, 2014.
- 809 101. **Roelands B, Buysse L, Pauwels F, Delbeke F, Deventer K, and Meeusen R.** No effect of caffeine
810 on exercise performance in high ambient temperature. *Eur J Appl Physiol* 111: 3089-3095, 2011.
- 811 102. **Sims ST, van VLIET L, Cotter J, and Rehrer N.** Sodium loading aids fluid balance and reduces
812 physiological strain of trained men exercising in the heat. *Medicine and science in sports and exercise*
813 39: 123, 2007.
- 814 103. **Zuhl MN, Lanphere KR, Kravitz L, Mermier CM, Schneider S, Dokladny K, and Moseley PL.**
815 Effects of oral glutamine supplementation on exercise-induced gastrointestinal permeability and tight
816 junction protein expression. *Journal of Applied Physiology* 116: 183-191, 2014.
- 817 104. **Dini M, Corbianco S, Rossi B, and Lucacchini A.** Hyperhydrating with glycerol: effects on
818 thermoregulation, hydration and athletic performance during specific exergonic exercise in a warm-
819 humid environment. *Sport Sciences for Health* 2: 1-7, 2007.
- 820 105. **Marino FE, Kay D, and Cannon J.** Glycerol hyperhydration fails to improve endurance
821 performance and thermoregulation in humans in a warm humid environment. *Pflügers Archiv* 446:
822 455-462, 2003.
- 823 106. **Snipe RMJ, Khoo A, Kitic CM, Gibson PR, and Costa RJS.** Carbohydrate and protein intake
824 during exertional heat stress ameliorates intestinal epithelial injury and small intestine permeability.
825 *Applied Physiology, Nutrition and Metabolism* 42: 1283-1292, 2017.
- 826 107. **Wingo JE, Casa DJ, Berger EM, Dellis WO, Knight JC, and McClung JM.** Influence of a pre-
827 exercise glycerol hydration beverage on performance and physiologic function during mountain-bike
828 races in the heat. *Journal of athletic training* 39: 169, 2004.
- 829 108. **Lundberg JO, Weitzberg E, Shiva S, and Gladwin MT.** The nitrate–nitrite–nitric oxide pathway
830 in mammals. *Nitrite and nitrate in human health and disease* 21-48, 2011.
- 831 109. **Fujii N, McGinn R, Stapleton JM, Paull G, Meade RD, and Kenny GP.** Evidence for
832 cyclooxygenase-dependent sweating in young males during intermittent exercise in the heat. *The*
833 *Journal of Physiology* 592: 5327-5339, 2014.
- 834 110. **Fujii N, McGinn R, Paull G, Stapleton JM, Meade RD, and Kenny GP.** Cyclooxygenase inhibition
835 does not alter methacholine-induced sweating. *Journal of Applied Physiology* 117: 1055-1062, 2014.
- 836 111. **Kellogg Jr D, Crandall C, Liu Y, Charkoudian N, and Johnson J.** Nitric oxide and cutaneous active
837 vasodilation during heat stress in humans. *Journal of applied physiology* 85: 824-829, 1998.
- 838 112. **McNamara TC, Keen JT, Simmons GH, Alexander LM, and Wong BJ.** Endothelial nitric oxide
839 synthase mediates the nitric oxide component of reflex cutaneous vasodilatation during dynamic
840 exercise in humans. *The Journal of Physiology* 592: 5317-5326, 2014.
- 841 113. **Shastry S, Dietz NM, Halliwill JR, Reed AS, and Joyner MJ.** Effects of nitric oxide synthase
842 inhibition on cutaneous vasodilation during body heating in humans. *Journal of applied physiology* 85:
843 830-834, 1998.
- 844 114. **Amano T, Okushima D, Breese BC, Bailey SJ, Koga S, and Kondo N.** Influence of dietary nitrate
845 supplementation on local sweating and cutaneous vascular responses during exercise in a hot
846 environment. *European Journal of Applied Physiology* 118: 1579-1588, 2018.
- 847 115. **Kent GL, Dawson B, Cox GR, Abbiss CR, Smith KJ, Croft KD, Lim ZX, Eastwood A, Burke LM,**
848 **and Peeling P.** Effect of dietary nitrate supplementation on thermoregulatory and cardiovascular
849 responses to submaximal cycling in the heat. *Eur J Appl Physiol* 118: 657-668, 2018.

850 116. **Kuennen M, Jansen L, Gillum T, Granados J, Castillo W, Nabiyyar A, and Christmas K.** Dietary
851 nitrate reduces the O₂ cost of desert marching but elevates the rise in core temperature. *Eur J Appl*
852 *Physiol* 115: 2557-2569, 2015.

853 117. **McQuillan JA, Casadio JR, Dulson DK, Laursen PB, and Kilding AE.** The Effect of Nitrate
854 Supplementation on Cycling Performance in the Heat in Well-Trained Cyclists. *Int J Sports Physiol*
855 *Perform* 13: 50-56, 2018.

856 118. **Tyler C, Coffey T, Hodges G, Tyler CJ, Coffey TRM, and Hodges GJ.** Acute L-arginine
857 supplementation has no effect on cardiovascular or thermoregulatory responses to rest, exercise, and
858 recovery in the heat. *European Journal of Applied Physiology* 116: 363-371, 2016.

859 119. **Kenny GP, Wilson TE, Flouris AD, and Fujii N.** Heat exhaustion. *Handbook of clinical neurology*
860 157: 505-529, 2018.

861 120. **Bell DG, and McLellan TM.** Exercise endurance 1, 3, and 6 h after caffeine ingestion in caffeine
862 users and nonusers. *Journal of applied physiology* 93: 1227-1234, 2002.

863 121. **Falk B, Burstein R, Rosenblum J, Shapiro Y, Zylber-Katz E, and Bashan N.** Effects of caffeine
864 ingestion on body fluid balance and thermoregulation during exercise. *Can J Physiol Pharmacol* 68:
865 889-892, 1990.

866 122. **Pitchford NW, Fell JW, Leveritt MD, Desbrow B, and Shing CM.** Effect of caffeine on cycling
867 time-trial performance in the heat. *J Sci Med Sport* 17: 445-449, 2014.

868 123. **Beaumont RE, and James LJ.** Effect of a moderate caffeine dose on endurance cycle
869 performance and thermoregulation during prolonged exercise in the heat. *J Sci Med Sport* 20: 1024-
870 1028, 2017.

871 124. **Roti MW, Casa DJ, Pumerantz AC, Watson G, Judelson DA, Dias JC, Ruffin K, and Armstrong**
872 **LE.** Thermoregulatory responses to exercise in the heat: chronic caffeine intake has no effect. *Aviat*
873 *Space Environ Med* 77: 124-129, 2006.

874 125. **Daniels JW, Molé PA, Shaffrath JD, and Stebbins CL.** Effects of caffeine on blood pressure,
875 heart rate, and forearm blood flow during dynamic leg exercise. *Journal of applied physiology* 85: 154-
876 159, 1998.

877 126. **Naulleau C, Jeker D, Pancrate T, Claveau P, Deshayes TA, Burke LM, and Goulet ED.** Effect of
878 Pre-Exercise Caffeine Intake on Endurance Performance and Core Temperature Regulation During
879 Exercise in the Heat: A Systematic Review with Meta-Analysis. *Sports Medicine* 52: 2431-2445, 2022.

880 127. **Nichols AW.** Heat-related illness in sports and exercise. *Current reviews in musculoskeletal*
881 *medicine* 7: 355-365, 2014.

882 128. **Westwood CS, Fallowfield JL, Delves SK, Nunns M, Ogden HB, and Layden JD.** Individual risk
883 factors associated with exertional heat illness: A systematic review. *Experimental Physiology* 106: 191-
884 199, 2021.

885 129. **Newsholme E, and Blomstrand dE.** Tryptophan, 5-hydroxytryptamine and a possible
886 explanation for central fatigue. In: *Fatigue* Springer, 1995, p. 315-320.

887 130. **Nybo L.** Hyperthermia and fatigue. *Journal of applied physiology* 104: 871-878, 2008.

888 131. **Nybo L, Rasmussen P, and Sawka MN.** Performance in the heat—physiological factors of
889 importance for hyperthermia-induced fatigue. *Comprehensive Physiology* 4: 657-689, 2011.

890 132. **Tumilty L, Davison G, Beckmann M, and Thatcher R.** Oral tyrosine supplementation improves
891 exercise capacity in the heat. *Eur J Appl Physiol* 111: 2941-2950, 2011.

892 133. **Fernstrom JD, and Fernstrom MH.** Tyrosine, phenylalanine, and catecholamine synthesis and
893 function in the brain. *The Journal of nutrition* 137: 1539S-1547S, 2007.

894 134. **Lang JA, Krajek AC, Schwartz KS, and Rand JE.** Oral L-tyrosine supplementation improves core
895 temperature maintenance in older adults. *Med Sci Sports Exerc* 52: 928-934, 2020.

896 135. **Miyazawa T, Kawabata T, Okazaki K, Suzuki T, Imai D, Hamamoto T, Matsumura S, and**
897 **Miyagawa T.** Oral administration of γ -aminobutyric acid affects heat production in a hot environment
898 in resting humans. *J Physiol Anthropol* 31: 3, 2012.

899 136. **Quéva C, Bremner-Danielsen M, Edlund A, Jonas Ekstrand A, Elg S, Erickson S, Johansson T,**
900 **Lehmann A, and Mattsson JP.** Effects of GABA agonists on body temperature regulation in GABAB
901 (1)-/- mice. *British journal of pharmacology* 140: 315-322, 2003.

902 137. **Watanabe M, Maemura K, Kanbara K, Tamayama T, and Hayasaki H.** GABA and GABA
903 receptors in the central nervous system and other organs. *International review of cytology* 213: 1-47,
904 2002.

905 138. **Yakimova K, Sann H, Schmid HA, and Pierau F-K.** Effects of GABA agonists and antagonists on
906 temperature-sensitive neurones in the rat hypothalamus. *The Journal of physiology* 494: 217-230,
907 1996.

908 139. **Cavagnini F, Invitti C, Pinto M, Maraschini C, Di Landro A, Dubini A, and Marelli A.** Effect of
909 acute and repeated administration of gamma aminobutyric acid (GABA) on growth hormone and
910 prolactin secretion in man. *European Journal of Endocrinology* 93: 149-154, 1980.

911 140. **Hori T.** An update on thermosensitive neurons in the brain: from cellular biology to thermal
912 and non-thermal homeostatic functions. *The Japanese Journal of Physiology* 41: 1-22, 1991.

913 141. **Nakayama T.** Thermosensitive neurons in the brain. *The Japanese Journal of Physiology* 35:
914 375-389, 1985.

915 142. **Ishiwata T, Saito T, Hasegawa H, Yazawa T, Kotani Y, Otokawa M, and Aihara Y.** Changes of
916 body temperature and thermoregulatory responses of freely moving rats during GABAergic
917 pharmacological stimulation to the preoptic area and anterior hypothalamus in several ambient
918 temperatures. *Brain research* 1048: 32-40, 2005.

919 143. **Miyazawa T, Kawabata T, Suzuki T, Imai D, Hamamoto T, Yoshikawa T, and Miyagawa T.** Effect
920 of oral administration of GABA on temperature regulation in humans during rest and exercise at high
921 ambient temperature. *Osaka City Med J* 55: 99-108, 2009.

922 144. **Deuchars SA, Milligan CJ, Stornetta RL, and Deuchars J.** GABAergic neurons in the central
923 region of the spinal cord: a novel substrate for sympathetic inhibition. *Journal of Neuroscience* 25:
924 1063-1070, 2005.

925 145. **Wible Jr JH, DiMicco JA, and Luft FC.** Hypothalamic GABA and sympathetic regulation in
926 spontaneously hypertensive rats. *Hypertension* 14: 623-628, 1989.

927 146. **Goldstein DS, Mccarty R, Polinsky RJ, and Kopin IJ.** Relationship between plasma
928 norepinephrine and sympathetic neural activity. *Hypertension* 5: 552-559, 1983.

929 147. **Mora-Rodriguez R, Gonzalez-Alonso J, Below PR, and Coyle EF.** Plasma catecholamines and
930 hyperglycaemia influence thermoregulation in man during prolonged exercise in the heat. *The Journal*
931 *of physiology* 491: 529-540, 1996.

932 148. **Nelson JL, and Robergs RA.** Exploring the potential ergogenic effects of glycerol
933 hyperhydration. *Sports Medicine* 37: 981-1000, 2007.

934 149. **Suvi S, Mooses M, Timpmann S, Medijainen L, Narõškina D, Unt E, and Ööpik V.** Impact of
935 sodium citrate ingestion during recovery after dehydrating exercise on rehydration and subsequent 40-
936 km cycling time-trial performance in the heat. *Appl Physiol Nutr Metab* 43: 571-579, 2018.

937 150. **Persky AM, Brazeau GA, and Hochhaus G.** Pharmacokinetics of the dietary supplement
938 creatine. *Clinical pharmacokinetics* 42: 557-574, 2003.

939 151. **Powers ME, Arnold BL, Weltman AL, Perrin DH, Mistry D, Kahler DM, Kraemer W, and Volek**
940 **J.** Creatine supplementation increases total body water without altering fluid distribution. *Journal of*
941 *athletic training* 38: 44, 2003.

942 152. **Watson G, Casa DJ, Fiala KA, Hile A, Roti MW, Healey JC, Armstrong LE, and Maresh CM.**
943 Creatine Use and Exercise Heat Tolerance in Dehydrated Men. *Journal of Athletic Training (National*
944 *Athletic Trainers' Association)* 41: 18-29, 2006.

945 153. **Anderson M, Cotter J, Garnham A, Casley D, and Febbraio MA.** Effect of glycerol-induced
946 hyperhydration on thermoregulation and metabolism during exercise in the heat. *International Journal*
947 *of Sport Nutrition and Exercise Metabolism* 11: 315-333, 2001.

- 948 154. **Coutts A, Reaburn P, Mummery K, and Holmes M.** The effect of glycerol hyperhydration on
949 Olympic distance triathlon performance in high ambient temperatures. *International journal of sport*
950 *nutrition and exercise metabolism* 12: 105-119, 2002.
- 951 155. **Nielsen B, Rowell LB, and Bonde-Petersen F.** Cardiovascular responses to heat stress and
952 blood volume displacements during exercise in man. *European journal of applied physiology and*
953 *occupational physiology* 52: 370-374, 1984.
- 954 156. **Kay D, and Marino FE.** Fluid ingestion and exercise hyperthermia: implications for
955 performance, thermoregulation, metabolism and the development of fatigue. *Journal of sports*
956 *sciences* 18: 71-82, 2000.
- 957 157. **Kilduff LP, Georgiades E, James N, Minnion RH, Mitchell M, Kingsmore D, Hadjicharlambous**
958 **M, and Pitsiladis YP.** The effects of creatine supplementation on cardiovascular, metabolic, and
959 thermoregulatory responses during exercise in the heat in endurance-trained humans. *Int J Sport Nutr*
960 *Exerc Metab* 14: 443-460, 2004.
- 961 158. **Sawka MN.** Physiological consequences of hypohydration: exercise performance and
962 thermoregulation. *Medicine and science in sports and exercise* 24: 657-670, 1992.
- 963 159. **Chato J.** Heat transfer to blood vessels. 1980.
- 964 160. **Keller KH, and Seiler Jr L.** An analysis of peripheral heat transfer in man. *Journal of applied*
965 *physiology* 30: 779-786, 1971.
- 966 161. **Morimoto T.** Thermoregulation and Body Fluids: Role of Blood Volume and Central Venous
967 Pressure. *The Japanese journal of physiology* 40: 165-179, 1990.
- 968 162. **Siegler JC, Carr AJ, Jardine WT, Convit L, Cross R, Chapman D, Burke LM, and Ross M.** The
969 hyperhydration potential of sodium bicarbonate and sodium citrate. *International journal of sport*
970 *nutrition and exercise metabolism* 32: 74-81, 2021.
- 971 163. **Katagiri A, Fujii N, Dobashi K, Lai Y-F, Tsuji B, and Nishiyasu T.** Sodium bicarbonate reduces
972 ventilation without altering core temperature threshold or sensitivity of hyperthermia-induced
973 hyperventilation in exercising humans. *American Journal of Physiology-Regulatory, Integrative and*
974 *Comparative Physiology* 2023.
- 975 164. **Katagiri A, Kitadai Y, Miura A, Fukuba Y, Fujii N, Nishiyasu T, and Tsuji B.** Sodium bicarbonate
976 ingestion mitigates the heat-induced hyperventilation and reduction in cerebral blood velocity during
977 exercise in the heat. *Journal of Applied Physiology* 131: 1617-1628, 2021.
- 978 165. **Lever M, and Slow S.** The clinical significance of betaine, an osmolyte with a key role in methyl
979 group metabolism. *Clinical biochemistry* 43: 732-744, 2010.
- 980 166. **McRae MP.** Betaine supplementation decreases plasma homocysteine in healthy adult
981 participants: a meta-analysis. *Journal of chiropractic medicine* 12: 20-25, 2013.
- 982 167. **Cholewa JM, Guimaraes-Ferreira L, and Zanchi NE.** Effects of betaine on performance and
983 body composition: a review of recent findings and potential mechanisms. *Amino acids* 46: 1785-1793,
984 2014.
- 985 168. **Willingham BD, Rentería LI, Ragland TJ, and Ormsbee MJ.** The effects of betaine
986 supplementation on fluid balance and heat tolerance during passive heat stress in men. *Physiol Rep*
987 11: e15792, 2023.
- 988 169. **Armstrong LE, Casa DJ, Roti MW, Lee EC, Craig SA, Sutherland JW, Fiala KA, and Maresch CM.**
989 Influence of betaine consumption on strenuous running and sprinting in a hot environment. *J Strength*
990 *Cond Res* 22: 851-860, 2008.
- 991 170. **Dangi SS, Dangi SK, Chouhan V, Verma M, Kumar P, Singh G, and Sarkar M.** Modulatory effect
992 of betaine on expression dynamics of HSPs during heat stress acclimation in goat (*Capra hircus*). *Gene*
993 575: 543-550, 2016.
- 994 171. **Willingham BD, Ragland TJ, and Ormsbee MJ.** Betaine supplementation may improve heat
995 tolerance: Potential mechanisms in humans. *Nutrients* 12: 2939, 2020.
- 996 172. **Attia Y, Hassan R, and Qota E.** Recovery from adverse effects of heat stress on slow-growing
997 chicks in the tropics 1: Effect of ascorbic acid and different levels of betaine. *Tropical animal health and*
998 *production* 41: 807-818, 2009.

- 999 173. **DiGiacomo K, Simpson S, Leury BJ, and Dunshea FR.** Dietary betaine impacts the physiological
1000 responses to moderate heat conditions in a dose dependent manner in sheep. *Animals* 6: 51, 2016.
- 1001 174. **Zulkifli I, Mysahra S, and Jin L.** Dietary supplementation of betaine (Betafin®) and response to
1002 high temperature stress in male broiler chickens. *Asian-australasian journal of animal sciences* 17: 244-
1003 249, 2004.
- 1004 175. **Vybíral S, Bárčayová L, Pešanová Z, and Janský L.** Pyrogenic effects of cytokines (IL-1 β , IL-6,
1005 TNF- α) and their mode of action on thermoregulatory centers and functions. *Journal of Thermal*
1006 *Biology* 30: 19-28, 2005.
- 1007 176. **Shin Y-O, Lee J-B, Min Y-K, and Yang H-M.** Effect of oligonol intake on cortisol and cytokines,
1008 and body temperature after leg immersion into hot water. *Food Science and Biotechnology* 20: 659-
1009 663, 2011.
- 1010 177. **Coceani F, Bishai I, Lees J, and Sirko S.** Prostaglandin E2 and fever: a continuing debate. *The*
1011 *Yale journal of biology and medicine* 59: 169, 1986.
- 1012 178. **Shin YO, Lee JB, Song YJ, Min YK, and Yang HM.** Oligonol supplementation attenuates body
1013 temperature and the circulating levels of prostaglandin E2 and cyclooxygenase-2 after heat stress in
1014 humans. *J Med Food* 16: 318-323, 2013.
- 1015 179. **Moreland LW.** Cytokines as targets for anti-inflammatory agents. *Annals of the New York*
1016 *Academy of Sciences* 1182: 88-96, 2009.
- 1017 180. **Bradford CD, Cotter JD, Thorburn MS, Walker RJ, and Gerrard DF.** Exercise can be pyrogenic
1018 in humans. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 292:
1019 R143-R149, 2007.
- 1020 181. **Szymanski MC, Gillum TL, Gould LM, Morin DS, and Kuennen MR.** Short-term dietary
1021 curcumin supplementation reduces gastrointestinal barrier damage and physiological strain responses
1022 during exertional heat stress. *J Appl Physiol (1985)* 124: 330-340, 2018.
- 1023 182. **Parsons K.** *Human thermal environments: the effects of hot, moderate, and cold environments*
1024 *on human health, comfort and performance.* CRC press, 2007.
- 1025 183. **Gulcin İ.** Antioxidants and antioxidant methods: An updated overview. *Archives of toxicology*
1026 94: 651-715, 2020.
- 1027 184. **Hoeldtke RD, Bryner KD, and VanDyke K.** Oxidative stress and autonomic nerve function in
1028 early type 1 diabetes. *Clinical Autonomic Research* 21: 19-28, 2011.
- 1029 185. **Fujii N, Meade RD, Schmidt MD, King KE, Boulay P, Ruzicka M, Amano T, and Kenny GP.** The
1030 effect of acute intradermal administration of ascorbate on heat loss responses in older adults with
1031 uncomplicated controlled hypertension. *Experimental Physiology* 107: 834-843, 2022.
- 1032 186. **Brossette T, Hundsdörfer C, Kröncke K-D, Sies H, and Stahl W.** Direct evidence that (-)-
1033 epicatechin increases nitric oxide levels in human endothelial cells. *European journal of nutrition* 50:
1034 595-599, 2011.
- 1035 187. **Schroeter H, Heiss C, Balzer J, Kleinbongard P, Keen CL, Hollenberg NK, Sies H, Kwik-Urbe C,**
1036 **Schmitz HH, and Kelm M.** (-)-Epicatechin mediates beneficial effects of flavanol-rich cocoa on vascular
1037 function in humans. *Proceedings of the National Academy of Sciences* 103: 1024-1029, 2006.
- 1038 188. **Nishimura R, Nishimura N, Iwase S, Takeshita M, Katashima M, Katsuragi Y, and Sato M.**
1039 Effects of catechin-enriched ion beverage intake on thermoregulatory function in a hot environment.
1040 *J Physiol Sci* 69: 39-45, 2019.
- 1041 189. **Matsumoto H, Takenami E, Iwasaki-Kurashige K, Osada T, Katsumura T, and Hamaoka T.**
1042 Effects of blackcurrant anthocyanin intake on peripheral muscle circulation during typing work in
1043 humans. *European journal of applied physiology* 94: 36-45, 2005.
- 1044 190. **Park E-Y, Kim M-H, Kim E-H, Lee E-K, Park I-S, Yang D-C, and Jun H-S.** Efficacy comparison of
1045 Korean ginseng and American ginseng on body temperature and metabolic parameters. *The American*
1046 *journal of Chinese medicine* 42: 173-187, 2014.
- 1047 191. **Kuennen M, Gillum T, Dokladny K, Bedrick E, Schneider S, and Moseley P.** Thermotolerance
1048 and heat acclimation may share a common mechanism in humans. *Am J Physiol Regul Integr Comp*
1049 *Physiol* 301: R524-533, 2011.

- 1050 192. **Pastor R, and Tur JA.** Antioxidant supplementation and adaptive response to training: a
1051 systematic review. *Current pharmaceutical design* 25: 1889-1912, 2019.
- 1052 193. **Lilja M, Mandić M, Apró W, Melin M, Olsson K, Rosenborg S, Gustafsson T, and Lundberg TR.**
1053 High doses of anti-inflammatory drugs compromise muscle strength and hypertrophic adaptations to
1054 resistance training in young adults. *Acta Physiologica* 222: e12948, 2018.
- 1055 194. **Kenny GP, Yardley J, Brown C, Sigal RJ, and Jay O.** Heat stress in older individuals and patients
1056 with common chronic diseases. *Cmaj* 182: 1053-1060, 2010.
- 1057 195. **Vandentorren S, Bretin P, Zeghnoun A, Mandereau-Bruno L, Croisier A, Cochet C, Ribéron J,**
1058 **Siberan I, Declercq B, and Ledrans M.** August 2003 heat wave in France: risk factors for death of elderly
1059 people living at home. *The European Journal of Public Health* 16: 583-591, 2006.
- 1060 196. **Chantler S, Griffiths A, Matu J, Davison G, Jones B, and Deighton K.** The effects of exercise on
1061 indirect markers of gut damage and permeability: a systematic review and meta-analysis. *Sports*
1062 *Medicine* 51: 113-124, 2021.
- 1063 197. **Bosenberg A, Brock-Utne J, Gaffin S, Wells M, and Blake G.** Strenuous exercise causes
1064 systemic endotoxemia. *Journal of applied physiology* 65: 106-108, 1988.
- 1065 198. **Lim CL, and Mackinnon LT.** The roles of exercise-induced immune system disturbances in the
1066 pathology of heat stroke: the dual pathway model of heat stroke. *Sports Medicine* 36: 39-64, 2006.
- 1067 199. **Selkirk GA, McLellan TM, Wright HE, and Rhind SG.** Mild endotoxemia, NF- κ B translocation,
1068 and cytokine increase during exertional heat stress in trained and untrained individuals. *American*
1069 *Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 295: R611-R623, 2008.
- 1070 200. **Gill SK, Allerton DM, Ansley-Robson P, Hemmings K, Cox M, and Costa RJS.** Does Short-Term
1071 High Dose Probiotic Supplementation Containing Lactobacillus casei Attenuate Exertional-Heat Stress
1072 Induced Endotoxaemia and Cytokinaemia? *International Journal of Sport Nutrition & Exercise*
1073 *Metabolism* 26: 268-275, 2016.
- 1074 201. **Morrison SA, Cheung SS, and Cotter JD.** Bovine colostrum, training status, and gastrointestinal
1075 permeability during exercise in the heat: a placebo-controlled double-blind study. *Appl Physiol Nutr*
1076 *Metab* 39: 1070-1082, 2014.
- 1077 202. **Osborne JO, Stewart IB, Beagley KW, Borg DN, and Minett GM.** Acute glutamine
1078 supplementation does not improve 20-km self-paced cycling performance in the heat. *Eur J Appl*
1079 *Physiol* 119: 2567-2578, 2019.
- 1080 203. **Shing C, Peake J, Lim C, Briskey D, Walsh N, Fortes M, Ahuja K, and Vitetta L.** Effects of
1081 probiotics supplementation on gastrointestinal permeability, inflammation and exercise performance
1082 in the heat. *European Journal of Applied Physiology* 114: 93-103, 2014.
- 1083 204. **Zuhl M, Dokladny K, Mermier C, Schneider S, Salgado R, and Moseley P.** The effects of acute
1084 oral glutamine supplementation on exercise-induced gastrointestinal permeability and heat shock
1085 protein expression in peripheral blood mononuclear cells. *Cell Stress Chaperones* 20: 85-93, 2015.
- 1086 205. **Yeh YJ, Law LYL, and Lim CL.** Gastrointestinal response and endotoxemia during intense
1087 exercise in hot and cool environments. *European journal of applied physiology* 113: 1575-1583, 2013.
- 1088 206. **Pugh J, Sage S, Hutson M, Doran D, Fleming S, Highton J, Morton J, Close G, Pugh JN, Doran**
1089 **DA, Fleming SC, Morton JP, and Close GL.** Glutamine supplementation reduces markers of intestinal
1090 permeability during running in the heat in a dose-dependent manner. *European Journal of Applied*
1091 *Physiology* 117: 2569-2577, 2017.
- 1092 207. **Nava RC, Zuhl MN, Moriarty TA, Amorim FT, Bourbeau KC, Welch AM, McCormick JJ, King KE,**
1093 **and Mermier CM.** The Effect of Acute Glutamine Supplementation on Markers of Inflammation and
1094 Fatigue During Consecutive Days of Simulated Wildland Firefighting. *J Occup Environ Med* 61: e33-e42,
1095 2019.
- 1096 208. **Barwood M, Gibson O, Gillis DJ, Jeffries O, Morris N, Pearce J, Ross M, Stevens C, Rinaldi K,**
1097 **and Kounalakis S.** Menthol as an Ergogenic aid for the Tokyo 2021 Olympic games: an Expert-Led
1098 consensus statement using the modified Delphi method. *Sports Medicine* 50: 1709-1727, 2020.

- 1099 209. **Andersen HH, Olsen RV, Møller HG, Eskelund PW, Gazerani P, and Arendt-Nielsen L.** A review
1100 of topical high-concentration L-menthol as a translational model of cold allodynia and hyperalgesia.
1101 *European journal of pain* 18: 315-325, 2014.
- 1102 210. **Liu Y, Mikrani R, He Y, Baig MMFA, Abbas M, Naveed M, Tang M, Zhang Q, Li C, and Zhou X.**
1103 TRPM8 channels: A review of distribution and clinical role. *European Journal of Pharmacology* 882:
1104 173312, 2020.
- 1105 211. **McKemy DD, Neuhauser WM, and Julius D.** Identification of a cold receptor reveals a general
1106 role for TRP channels in thermosensation. *Nature* 416: 52-58, 2002.
- 1107 212. **Peier AM, Moqrich A, Hergarden AC, Reeve AJ, Andersson DA, Story GM, Earley TJ, Dragoni
1108 I, McIntyre P, and Bevan S.** A TRP channel that senses cold stimuli and menthol. *Cell* 108: 705-715,
1109 2002.
- 1110 213. **Harrington AM, Hughes PA, Martin CM, Yang J, Castro J, Isaacs NJ, Blackshaw LA, and Brierley
1111 SM.** A novel role for TRPM8 in visceral afferent function. *Pain*® 152: 1459-1468, 2011.
- 1112 214. **Flood TR, Waldron M, and Jeffries O.** Oral L-menthol reduces thermal sensation, increases
1113 work-rate and extends time to exhaustion, in the heat at a fixed rating of perceived exertion. *European
1114 journal of applied physiology* 117: 1501-1512, 2017.
- 1115 215. **Jeffries O, Goldsmith M, and Waldron M.** L-Menthol mouth rinse or ice slurry ingestion during
1116 the latter stages of exercise in the heat provide a novel stimulus to enhance performance despite
1117 elevation in mean body temperature. *European journal of applied physiology* 118: 2435-2442, 2018.
- 1118 216. **Jeffries O, and Waldron M.** The effects of menthol on exercise performance and thermal
1119 sensation: A meta-analysis. *Journal of science and medicine in sport* 22: 707-715, 2019.
- 1120 217. **Pokora I, Wolowski Ł, and Wyderka P.** The effect of a single dose of the Thermo Speed
1121 Extreme (Olimp) thermogenic supplement on circulatory functions and body temperatures at rest in
1122 male and female subjects. *Baltic Journal of Health & Physical Activity* 11: 11-25, 2019.
- 1123 218. **Taqvi SIH, Shah AJ, and Gilani AH.** Blood pressure lowering and vasomodulator effects of
1124 piperine. *Journal of cardiovascular pharmacology* 52: 452-458, 2008.
- 1125 219. **Ahmadian M, Dabidi Roshan V, and Ashourpore E.** Taurine supplementation improves
1126 functional capacity, myocardial oxygen consumption, and electrical activity in heart failure. *Journal of
1127 dietary supplements* 14: 422-432, 2017.
- 1128 220. **Lee H, Paik I, and Park T.** Effects of dietary supplementation of taurine, carnitine or glutamine
1129 on endurance exercise performance and fatigue parameters in athletes. *Korean Journal of Nutrition*
1130 36: 711-719, 2003.
- 1131 221. **Zhang M, Izumi I, Kagamimori S, Sokejima S, Yamagami T, Liu Z, and Qi B.** Role of taurine
1132 supplementation to prevent exercise-induced oxidative stress in healthy young men. *Amino acids* 26:
1133 203-207, 2004.
- 1134 222. **Benjamin J, Makharia G, Ahuja V, Anand Rajan K, Kalaivani M, Gupta SD, and Joshi YK.**
1135 Glutamine and whey protein improve intestinal permeability and morphology in patients with Crohn's
1136 disease: a randomized controlled trial. *Digestive diseases and sciences* 57: 1000-1012, 2012.
- 1137 223. **Koenigsberg PS, Martin KK, Hlava HR, and Riedesel ML.** Sustained hyperhydration with
1138 glycerol ingestion. *Life sciences* 57: 645-653, 1995.
- 1139 224. **Easton C, Turner S, and Pitsiladis YP.** Creatine and glycerol hyperhydration in trained subjects
1140 before exercise in the heat. *Int J Sport Nutr Exerc Metab* 17: 70-91, 2007.
- 1141 225. **Polyviou TP, Pitsiladis YP, Wu Chean L, Pantazis T, Hambly C, Speakman JR, and Malkova D.**
1142 Thermoregulatory and cardiovascular responses to creatine, glycerol and alpha lipoic acid in trained
1143 cyclists. *Journal of the International Society of Sports Nutrition* 9: 29-40, 2012.
- 1144 226. **Klarod K, Gatterer H, Frontull V, Philippe M, and Burtscher M.** Effects of short-term
1145 antioxidant supplementation on oxidative stress and exercise performance in the heat and the cold.
1146 *International journal of physiology, pathophysiology and pharmacology* 7: 98, 2015.
- 1147 227. **Costa RJ, Henningsen K, Gaskell SK, Alcock R, Mika A, Rauch C, Chevront SN, Blazy P, and
1148 Kenefick R.** Amino acid-based beverage interventions ameliorate exercise-induced gastrointestinal

1149 syndrome in response to exertional-heat stress: The heat exertion amino acid technology (HEAAT)
1150 study. *International Journal of Sport Nutrition and Exercise Metabolism* 33: 230-242, 2023.

1151 228. **Taylor L, Lee BJ, Gibson OR, Midgley AW, Watt P, Mauger A, and Castle P.** Effective
1152 microorganism – X attenuates circulating superoxide dismutase following an acute bout of intermittent
1153 running in hot, humid conditions. *Research in Sports Medicine* 24: 130-144, 2016.

1154 229. **Baylis A, Cameron-Smith D, and Burke LM.** Inadvertent doping through supplement use by
1155 athletes: assessment and management of the risk in Australia. *International journal of sport nutrition*
1156 *and exercise metabolism* 11: 365-383, 2001.

1157 230. **Casey A, Hughes J, Izard RM, and Greeves JP.** Supplement use by UK-based British Army
1158 soldiers in training. *British journal of nutrition* 112: 1175-1184, 2014.

1159