

## LIPID MAPS: Powering discovery in lipidomics

Lauren Cockayne<sup>1</sup>, Matthew J. Conroy<sup>1</sup>, Chetin Baloglu<sup>2</sup>, Eoin Fahy<sup>3</sup>, Gerhard Hagn<sup>4</sup>, Oswald Quehenberger<sup>5</sup>, Aaron M. Armando<sup>5</sup>, Gabriele Lombardi Bendoula<sup>6</sup>, Jean-Marie Galano<sup>7</sup>, Ángel Sánchez-Illana<sup>7,8</sup>, Thierry Durand<sup>7</sup>, Nadja Kampschulte<sup>9</sup>, Paul D. Kennedy<sup>10</sup>, Miguel Gijón<sup>10</sup>, Hiroshi Tsugawa<sup>11,12</sup>, Makoto Arita<sup>11</sup>, Kirk Maxey<sup>10</sup>, Meghan Truskowski<sup>10</sup>, Ondrej Kuda<sup>13</sup>, Shazia Khan<sup>14</sup>, Natalie Z. M. Homer<sup>14</sup>, Yuki Matsuzawa<sup>12</sup>, Rosario Domingues<sup>15</sup>, Peter J. Meikle<sup>16</sup>, Corey Giles<sup>16</sup>, Kevin Huynh<sup>16</sup>, Robert C. Murphy<sup>17</sup>, Zidan Wang<sup>18</sup>, Yu Xia<sup>18</sup>, Xue Li Guan<sup>19</sup>, Kim Ekroos<sup>20</sup>, Gerhard Liebisch<sup>21</sup>, Alfred H. Merrill, Jr<sup>22</sup>, Andrea F. Lopez-Clavijo<sup>23</sup>, Dominic Campopiano<sup>24</sup>, Craig E. Wheelock<sup>4,25</sup>, Shankar Subramaniam<sup>3</sup>, Robert Andrews<sup>1</sup>, Laura Goracci<sup>26</sup>, Zhixu Ni<sup>27</sup>, Maria Fedorova<sup>6</sup>, Simon Andrews<sup>2</sup>, William Griffiths<sup>28</sup>, Ruth Andrew<sup>14</sup>, Edward A. Dennis<sup>5,29</sup>, Valerie B. O'Donnell<sup>1\*</sup>

<sup>1</sup>Systems Immunity Research Institute, School of Medicine, Cardiff University, CF14 4XN, UK. <sup>2</sup>Babraham Institute, Babraham Research Campus, Cambridge, CB22 3AT, UK. <sup>3</sup>Department of Bioengineering, University of California, San Diego, La Jolla, CA 92093, USA. <sup>4</sup>Unit of Integrative Metabolomics, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden. <sup>5</sup>Department of Pharmacology, University of California San Diego, La Jolla, CA 92093, USA. <sup>6</sup>Center of Membrane Biochemistry and Lipid Research, University Hospital and Faculty of Medicine Carl Gustav Carus of TU Dresden, Dresden, Germany. <sup>7</sup>Institut des Biomolécules Max Mousseron (IBMM), Pôle Chimie Balard Recherche, CNRS, Université de Montpellier, ENSCN, Montpellier, France. <sup>8</sup>Department of Analytical Chemistry, University of Valencia, Burjassot, Spain. <sup>9</sup>School of Mathematics and Natural Sciences, University of Wuppertal, Wuppertal, Germany. <sup>10</sup>Cayman Chemical Company, Ann Arbor, MI 48108, USA. <sup>11</sup>RIKEN Center for Integrative Medical Sciences (IMS), Yokohama, Kanagawa, Japan. <sup>12</sup>Department of Biotechnology and Life Science, Tokyo University of Agriculture and Technology, Koganei, Tokyo, Japan. <sup>13</sup>Institute of Physiology of the Czech Academy of Sciences, Videnska 1083, Prague 14200, Czech Republic. <sup>14</sup>Institute of Neurosciences and Cardiovascular Research, University of Edinburgh, Edinburgh, UK. <sup>15</sup>Department of Chemistry, University of Aveiro, Aveiro, Portugal. <sup>16</sup>Baker Heart and Diabetes Institute, Melbourne, Victoria, 3004, Australia. <sup>17</sup>Department of Pharmacology, University of Colorado Denver, Denver, CO 80045, USA. <sup>18</sup>MOE Key Laboratory of Bioorganic Phosphorus Chemistry & Chemical Biology, Department of Chemistry, Tsinghua University, Beijing, 100084, China. <sup>19</sup>Lee Kong Chian School of Medicine, Nanyang Technological University, Singapore; and Novo Nordisk Foundation Centre for Metabolic Disease Research, Faculty of Health and Medical Sciences, University of Copenhagen, Denmark. <sup>20</sup>Lipidomics Consulting Ltd., Esbo, Finland. <sup>21</sup>Institute of Clinical Chemistry and Laboratory Medicine, University Hospital Regensburg, Regensburg, Germany. <sup>22</sup>School of Biological Sciences, Georgia Institute of Technology, Atlanta, GA 30338, USA. <sup>23</sup>CRUK Cambridge Institute, Cambridge University, Cambridge, UK. <sup>24</sup>School of Chemistry, University of Edinburgh, Scotland, UK. <sup>25</sup>Department of Respiratory Medicine and Allergy, Karolinska University Hospital, Stockholm, Sweden. <sup>26</sup>Department of Chemistry, Biology, and Biotechnology, University of Perugia, Perugia, Italy. <sup>27</sup>Institute of Data and Information (iDI), Tsinghua Shenzhen International Graduate School, 518055 Shenzhen, China. <sup>28</sup>Swansea University Medical School, Singleton Park, Swansea SA2 8PP, Wales, UK. <sup>29</sup>Department of Biochemistry and Molecular Biophysics and Department of Chemistry, University of California, San Diego, La Jolla, CA 92093, USA.

\*Corresponding author. Email: o-donnellvb@cardiff.ac.uk

## **Abstract**

As lipidomics approaches its 25<sup>th</sup> anniversary, we explore how lipid research has matured over the years, while highlighting emerging innovations that are expanding our ability to study these diverse, life-critical biomolecules. In particular, we showcase the community-driven, open-access databases, software, and educational resources made freely available through the ELIXIR Core Data Resource LIPID MAPS for the benefit of both established and new researchers.

Lipid species and their concentrations change in response to internal and external stimuli, serving as valuable indicators of health and disease. About 25 years ago, the advent of benchtop mass spectrometry (MS) drove the emergence of lipidomics, with the LIPID MAPS consortium ([www.lipidmaps.org](http://www.lipidmaps.org)) established to meet the growing need for standardized nomenclature, systematic structural classification, and the organization of experimental data. Here, we provide an update on state-of-the-art innovations in lipidomics, highlighting how LIPID MAPS supports members of the community to apply these new approaches to their own research.

In 2019 (1), lipid researchers were anticipating how large-scale, global high-throughput analysis of lipids might contribute to or drive the field of precision medicine. Since then, methods enabling the quantitation of hundreds-to-thousands of lipids driven by innovations in chromatography and MS/MS have revolutionized lipidomics of large cohorts and tissues. Quantification of multiple molecular species across diverse lipid families in a single sample is now robust and becoming more routine, with increased annotation confidence. A lipid-based clinical score for cardiovascular

disease was developed, and stratification of pancreatic cancer was also achieved, using serum or plasma (2, 3). Clinical translation of targeted lipidomics has been achieved for some diseases; for example, sphingolipidomics in the screening of newborns for metachromatic leukodystrophy, whereby a sulfatide acts as a biomarker (4). Aiming toward translation, NIST-1950 plasma was adopted as reference material, and ring trials for ceramides and bile acids were initiated by the International Lipidomics Society (ILS) and the Singapore Lipidomics Group (SLING), with the latter reporting in 2024 (5).

New lipidomics innovations continue to be driven by technological and methodological advances. These include “enhanced fragmentation” achieved by oxygen attachment dissociation (OAD), electron-activated dissociation (EAD), ultraviolet-photodissociation (UV-PD), and ozone-induced dissociation (OzID), which can also determine stereochemistry. These methods generate substantially more structural information than is generated by collision-induced dissociation (CAD/CID), and coupled with high-resolution MS, they greatly increase the molecular diversity of detectable lipids that can be curated. However, we generally lack reference spectra for these “enhanced fragmentation” modalities. A second area of development is molecular networking, whereby data-dependent analysis of globally sourced MS/MS data is expanding the numbers of known lipids, including, for example, new polyamine bile amidates (6). Here, a challenge lies in the annotation of species using rule-based approaches unique to lipids. Last, driven by community-agreed needs to support data quality, a substantial focus has been on standardizing lipidomics, by developing the ILS reporting checklist and technical recommendations for oxylipin analysis (7, 8), which are aligned with and supported by the data and resources provided by LIPID MAPS.

Working alongside experimental data depositories (Metabolomics Workbench and MetaboLights), small-molecule databases (ChEBI, SwissLipids, and PubChem), and supported by communities contributing data and establishing reporting standards (ILS and EpiLipidNET), LIPID MAPS provides databases, tools, and resources to enable large-scale lipidomics, integration of multi-omics (systems biology), and smaller scale, targeted lipid research. Indicating its size and scale, LIPID MAPS was accessed by >182,000 users in 2025, from almost every country in the world, with >4.1 million pageviews (Fig. 1). In 2024, LIPID MAPS became an ELIXIR Core Data Resource and an ELIXIR Deposition Database and joined the Global Biodata Coalition.

Many lipids from diverse biological sources are not fully structurally characterized and thus remain missing from databases. This is a growing concern, especially with the increasing use of enhanced fragmentation methods, and is particularly relevant for “epilipids,” which are modified by enzymatic, non-enzymatic, or environmental factors and are detectable by MS but lack full structural annotation. Although these can be included in resources for molecular networking, such as the Global Natural Products Social Molecular Networking database (GNPS), they often lack structural descriptions. Developed in collaboration with the EU COST Action (CA19105) network EpiLipidNET, the LIPID MAPS database Partial Spectra DB provides >450 annotated and expert-curated MS/MS spectra and associated metadata, including for mammalian neutral glycosphingolipids, microalgal glycosyldiacylglycerols, and bacterial acylaminosugars. Further supporting efforts to structurally identify lipids, ~1800 interactive, downloadable MS/MS spectra for oxylipins and sterols were added to the LIPID MAPS Standards Spectra DB with standardized metadata. Stable identifiers for reporting in publications were provided in a new Shorthand DB to identify lipids described at the species/molecular species levels. Updates to lipid shorthand

nomenclature were implemented, including the expansion of bile acid conjugate notation to include amino acid conjugates using three-letter codes and modifying taurine conjugate notation from “;T” to “;Tau” to eliminate ambiguity.

A large number of new features were added to the LIPID MAPS Structure Database (LMSD) since the last database update (9). These include biological information provided by Cayman Chemical for >1500 lipids. LMSD now hosts links from individual lipids to raw data from thousands of lipidomics studies held on Metabolomics Workbench. Links to nuclear magnetic resonance (NMR) spectra (NP-MRD) and MS/MS data on MassBankEU were included. The open-access lipidomics software MS-DIAL 5 incorporated LMSD search functionality, as did Lipostar. LIPID MAPS reached a milestone of 50,000 lipid structures curated in LMSD, including structures discovered by enhanced fragmentation and through molecular networking. LIPID MAPS made available two large archives of historical GC/MS data to aid in the annotation of spectra obtained with this approach. About 1300 steroid spectra generated by Jan Sjövall at the Karolinska Institutet, accompanied by handwritten structural depictions and supplementary notations, were uploaded, whereas a collection of >700 spectra for prostaglandins, leukotrienes, and related fatty acids authored by Cecil Pace-Asciak was reproduced (10).

Several new tools have been added to LIPID MAPS, including: (i) a standalone tool enabling the direct calculation and visualization of isotope patterns for user-defined molecular formulae; (ii) an Android version of the Lipid Calculator App, complementing the previously updated iOS application available from LIPID MAPS; (iii) a steroid structure drawing tool that accommodates >20 sterol cores, including steroid hormones, bile acids, and plant sterols; and (iv) a

multifunctional tool enabling the generation of structures for cholesterol esters and many other lipids.

As lipidomics continues to expand in scale, complexity, and clinical relevance, sustained community engagement and data sharing will be essential to ensure that the emerging lipid diversity can be confidently annotated, interpreted, and translated. By evolving in step with new technologies and standards, LIPID MAPS is positioned to remain a central, enabling infrastructure for the next phase of discovery-driven and translational lipidomics.

## References and Notes

1. V. B. O'Donnell, E. A. Dennis, M. J. O. Wakelam, S. Subramaniam, LIPID MAPS: Serving the next generation of lipid researchers with tools, resources, data, and training. *Science Signaling* **12**, eaaw2964 (2019).
2. J. Wu, C. Giles, A. Dakic, H. B. Beyene, K. Huynh, T. Wang, T. Meikle, G. Olshansky, A. Salim, T. Duong, G. F. Watts, J. Hung, J. Hui, G. Cadby, J. Beilby, J. Blangero, E. K. Moses, J. E. Shaw, D. J. Magliano, D. Zhu, J. Y. Yang, S. M. Grieve, A. Wilson, C. K. Chow, S. T. Vernon, M. P. Gray, G. A. Figtree, M. J. Carrington, M. Inouye, T. H. Marwick, P. J. Meikle, Lipidomic Risk Score to Enhance Cardiovascular Risk Stratification for Primary Prevention. *Journal of the American College of Cardiology* **84**, 434–446 (2024).
3. D. Wolrab, R. Jirásko, E. Cífková, M. Höring, D. Mei, M. Chocholoušková, O. Peterka, J. Idkowiak, T. Hrnčiarová, L. Kuchař, R. Ahrends, R. Brumarová, D. Friedecký, G. Vivo-Truyols, P. Škrha, J. Škrha, R. Kučera, B. Melichar, G. Liebisch, R. Burkhardt, M. R. Wenk, A. Cazenave-Gassiot, P. Karásek, I. Novotný, K. Greplová, R. Hrstka, M. Holčapek, Lipidomic profiling of human serum enables detection of pancreatic cancer. *Nature Communications* **13**, 124 (2022).
4. S. Bekri, A. Bley, H. A. Brown, C. Chanson, H. J. Church, M. H. Gelb, X. Hong, N. Janzen, D. C. Kasper, T. Mechtler, G. Morton, S. Murko, P. Oliva, A. Tebani, T. H. Y. Wu, Higher precision, first tier newborn screening for metachromatic leukodystrophy using 16:1-OH-sulfatide. *Molecular Genetics and Metabolism* **142**, 108436 (2024).
5. F. Torta, N. Hoffmann, B. Burla, I. Alecu, M. Arita, T. Bamba, S. A. L. Bennett, J. Bertrand-Michel, B. Brügger, M. P. Cala, D. Camacho-Muñoz, A. Checa, M. Chen, M. Chocholoušková, M. Cinel, E. Chu-Van, B. Colsch, C. Coman, L. Connell, B. C. Sousa, A. M. Dickens, M. Fedorova, F. F. Eiriksson, H. Gallart-Ayala, M. Ghorasaini, M. Giera, X. L. Guan, M. Haid, T. Hankemeier, A. Harms, M. Höring, M. Holčapek, T. Hornemann, C. Hu, A. J. Hülsmeier, K. Huynh, C. M. Jones, J. Ivanisevic, Y. Izumi, H. C. Köfeler, S.

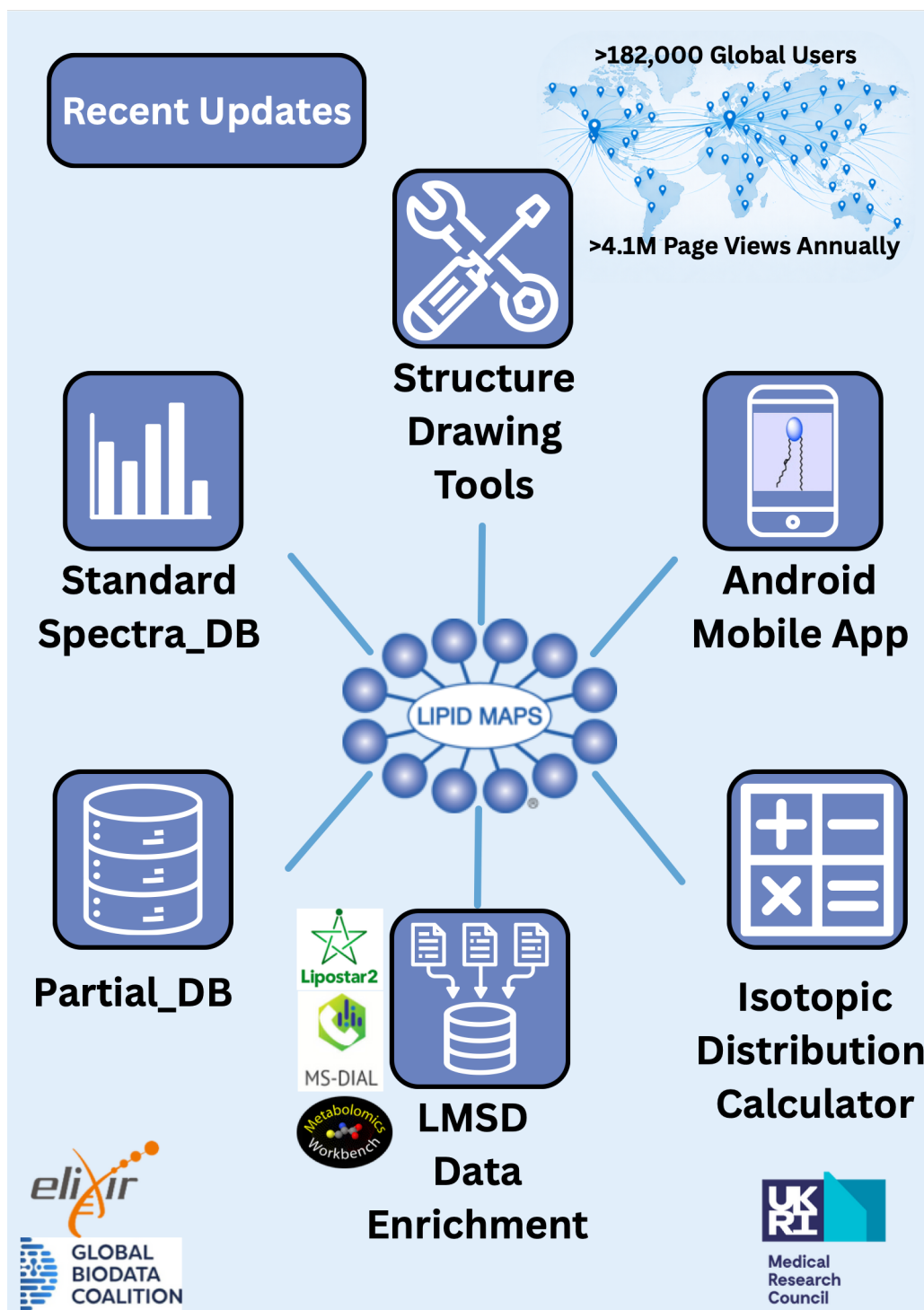
- M. Lam, M. Lange, J. C. Lee, G. Liebisch, K. Lippa, A. F. Lopez-Clavijo, M. Manzi, M. R. Martinefski, R. G. H. Math, S. Mayor, P. J. Meikle, M. E. Monge, M. H. Moon, S. Muralidharan, A. Nicolaou, T. Nguyen-Tran, V. B. O'Donnell, M. Orešič, A. Ramanathan, F. Riols, D. Saigusa, T. B. Schock, H. Schwartz-Zimmermann, G. Shui, M. Singh, M. Takahashi, M. Thorsteinsdóttir, N. Tomiyasu, A. Tournadre, H. Tsugawa, V. J. Tyrrell, G. van der Gugten, M. O. Wakelam, C. E. Wheelock, D. Wolrab, G. Xu, T. Xu, J. A. Bowden, K. Ekroos, R. Ahrends, M. R. Wenk, Concordant inter-laboratory derived concentrations of ceramides in human plasma reference materials via authentic standards. *Nature Communications* **15**, 8562 (2024).
6. I. Mohanty, H. Mannocho-Russo, J. V. Schweer, Y. El Abiead, W. Bittremieux, S. Xing, R. Schmid, S. Zuffa, F. Vasquez, V. B. Muti, J. Zemlin, O. E. Tovar-Herrera, S. Moraís, D. Desai, S. Amin, I. Koo, C. W. Turck, I. Mizrahi, P. M. Kris-Etherton, K. S. Petersen, J. A. Fleming, T. Huan, A. D. Patterson, D. Siegel, L. R. Hagey, M. Wang, A. T. Aron, P. C. Dorrestein, The underappreciated diversity of bile acid modifications. *Cell* **187**, 1801–1818.e1820 (2024).
  7. D. Kopczynski, C. S. Ejsing, J. G. McDonald, T. Bamba, E. S. Baker, J. Bertrand-Michel, B. Brügger, C. Coman, S. R. Ellis, T. J. Garrett, W. J. Griffiths, X. L. Guan, X. Han, M. Höring, M. Holčapek, N. Hoffmann, K. Huynh, R. Lehmann, J. W. Jones, R. Kaddurah-Daouk, H. C. Köfeler, P. J. Meikle, T. O. Metz, V. B. O'Donnell, D. Saigusa, D. Schwudke, A. Shevchenko, F. Torta, J. A. Vizcaíno, R. Welti, M. R. Wenk, D. Wolrab, Y. Xia, K. Ekroos, R. Ahrends, G. Liebisch, The lipidomics reporting checklist a framework for transparency of lipidomic experiments and repurposing resource data. *Journal of Lipid Research* **65**, (2024).
  8. N. H. Schebb, N. Kampschulte, G. Hagn, K. Plitzko, S. W. Meckelmann, S. Ghosh, R. Joshi, J. Kuligowski, D. Vuckovic, M. T. Botana, Á. Sánchez-Illana, F. Zandkarimi, A. Das, J. Yang, L. Schmidt, A. Checa, H. M. Roche, A. M. Armando, M. L. Edin, F. B. Lih, J. J. Aristizabal-Henao, S. Miyamoto, F. Giuffrida, A. Moussaieff, R. Domingues, M. Rothe, C. Hinz, U. S. Das, K. M. Rund, A. Y. Taha, R. K. Hofstetter, M. Werner, O. Werz, A. S. Kahnt, J. Bertrand-Michel, P. Le Faouder, R. Gurke, D. Thomas, F. Torta, I. Milic, I. H. K. Dias, C. M. Spickett, D. Biagini, T. Lomonaco, H. Idborg, J.-Y. Liu, M. Fedorova, D. A. Ford, A. Barden, T. A. Mori, P. D. Kennedy, K. Maxey, J. Ivanisevic, H. Gallart-Ayala, C. Gladine, M. Wenk, J.-M. Galano, T. Durand, K. D. Stark, C. Barbas, U. Garscha, S. L. Gelhaus, U. Ceglarek, N. Flamand, J. L. Griffin, R. Ahrends, M. Arita, D. C. Zeldin, F. J. Schopfer, O. Quehenberger, R. Julian, A. Nicolaou, I. A. Blair, M. P. Murphy, B. D. Hammock, B. Freeman, G. Liebisch, C. N. Serhan, H. C. Köfeler, P.-J. Jakobsson, D. Steinhilber, M. H. Gelb, M. Holčapek, R. Andrew, M. Giera, G. A. FitzGerald, R. C. Murphy, J. W. Newman, E. A. Dennis, K. Ekroos, G. L. Milne, M. A. Gijón, H. W. Vesper, C. E. Wheelock, V. B. O'Donnell, Technical recommendations for analyzing oxylipins by liquid chromatography–mass spectrometry. *Science Signaling* **18**, eadw1245 (2025).
  9. M. J. Conroy, R. M. Andrews, S. Andrews, L. Cockayne, Edward A. Dennis, E. Fahy, C. Gaud, William J. Griffiths, G. Jukes, M. Kolchin, K. Mendivelso, Andrea F. Lopez-Clavijo, C. Ready, S. Subramaniam, Valerie B. O'Donnell, LIPID MAPS: update to databases and tools for the lipidomics community. *Nucleic Acids Research* **52**, D1677–D1682 (2023).

10. C. R. Pace-Asciak, Mass spectra of prostaglandins and related products. *Adv Prostaglandin Thromboxane Leukot Res* **18**, 1–565 (1989).

**Acknowledgments:** We gratefully acknowledge the following for support with the curation of spectra for LIPID MAPS: A. Faqehi, A. Rutter, G. Naredo-Gonzalez, J. Weatherill, N. Denver, I. Stasinopoulos, S. G. Denham, R. Morgan, J. Simpson, S. Laforest, G. Just (University of Edinburgh, UK), T. Melo, B. Neves, T. Conde (University of Aviero, Portugal), M. A. Chromik, R. Kirchhoff, A. Löwen, K. Mosel, K. Plitzko, L. Scholz, L. Wende, and M. Wiebel (University of Wuppertal, Germany). **Funding:** We gratefully acknowledge Partnership Grant funding from the Medical Research Council/UKRI for LIPID MAPS (MR/Y000064/1). LIPID MAPS receives financial support from Cayman Chemical, Avanti Research, and Larodan. **Competing interests:** The authors declare that they have no competing interests.

**Fig. 1. Schematic summarizing updates to the site and the global reach of LIPID MAPS.**

Usage data were derived from Google Analytics.



**Figure 1. Schematic showing a summary of recent updates and global reach of LIPID MAPS.** User figures are from Google Analytics.